

# Package: DESpace (via r-universe)

June 2, 2026

**Type** Package

**Title** DESpace: a framework to discover spatially variable genes and differential spatial patterns across conditions

**Version** 2.4.0

**Description** Intuitive framework for identifying spatially variable genes (SVGs) and differential spatial variable pattern (DSP) between conditions via edgeR, a popular method for performing differential expression analyses. Based on pre-annotated spatial clusters as summarized spatial information, DESpace models gene expression using a negative binomial (NB), via edgeR, with spatial clusters as covariates. SVGs are then identified by testing the significance of spatial clusters. For multi-sample, multi-condition datasets, we again fit a NB model via edgeR, incorporating spatial clusters, conditions and their interactions as covariates. DSP genes-representing differences in spatial gene expression patterns across experimental conditions-are identified by testing the interaction between spatial clusters and conditions.

**biocViews** Spatial, SingleCell, RNASeq, Transcriptomics, GeneExpression, Sequencing, DifferentialExpression,StatisticalMethod, Visualization

**License** GPL-3

**Depends** R (>= 4.5.0)

**Imports** edgeR, limma, dplyr, stats, Matrix, SpatialExperiment, ggplot2, SummarizedExperiment, S4Vectors, BiocGenerics, data.table, assertthat, terra, sf, spatstat.explore, spatstat.geom, ggforce, ggnewscale, patchwork, BiocParallel, methods, scales, scuttle

**Suggests** knitr, rmarkdown, testthat, BiocStyle, muSpaData, ExperimentHub, spatialLIBD, purrr, reshape2, tidyverse, concaveman

**VignetteBuilder** knitr

**RoxygenNote** 7.3.3

**ByteCompile** true**Encoding** UTF-8**URL** <https://github.com/peicai/DESpace>,  
<https://peicai.github.io/DESpace/>**BugReports** <https://github.com/peicai/DESpace/issues>**Config/pak/sysreqs** libabsl-dev cmake libfontconfig1-dev libfreetype6-dev libgdal-dev gdal-  
bin libgeos-dev libmagick++-dev gsfonts libicu-dev libssl-dev libproj-dev libsqlite3-  
dev libudunits2-dev zlib1g-dev**Repository** <https://bioc-release.r-universe.dev>**Date/Publication** 2026-04-28 13:00:19 UTC**RemoteUrl** <https://github.com/bioc/DESpace>**RemoteRef** RELEASE\_3\_23**RemoteSha** 93c47a3230327e28ab98b73f05f9ad094ec14c19

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dsp_test	<i>dsp_test</i>
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## Description

'dsp\_test' identifies differential spatial pattern (DSP) genes between conditions from spatially-resolved transcriptomics data, provided spatial clusters are available.

## Usage

```
dsp_test(
  spe,
  design = NULL,
  cluster_col,
  sample_col,
```

```

    condition_col,
    test = "QLF",
    min_counts = 20,
    min_non_zero_spots = 10,
    min_pct_cells = 0.5,
    filter_gene = FALSE,
    filter_cluster = TRUE,
    verbose = FALSE
)

```

### Arguments

spe	SpatialExperiment or SingleCellExperiment.
design	Matrix or array. Numeric design matrix for a regression-like model created by 'model.matrix' function.
cluster_col	Character. Column name of spatial clusters in colData(spe).
sample_col	Character. Column name of sample ids in colData(spe). Sample ids must be either a factor or character.
condition_col	Character. Column name of condition ids in colData(spe).
test	Character. Either 'QLF' or 'LRT'. Default is 'QLF'. Specifies whether to perform quasi-likelihood F-tests ('edgeR::glmQLFTest') or likelihood ratio tests ('edgeR::glmLRT'). Quasi-likelihood tests apply empirical Bayes moderation to genewise dispersions, account for intra-sample variability, and provide more conservative error control.
min_counts	Numeric. Minimum number of counts per sample (across all spots) for a gene to be analyzed.
min_non_zero_spots	Numeric. Minimum number of non-zero spots per sample, for a gene to be analyzed.
min_pct_cells	Numeric. Minimum percentage of cells required for each cluster to be included in the analysis across the specified conditions. Default value is 0.5 (i.e., 0.5% of total cells per cluster per condition).
filter_gene	Logical. If TRUE, <code>dsp_test</code> filters genes by requiring them to be expressed in at least 'min_non_zero_spots' cells and have at least 'min_counts' counts per sample across all locations.
filter_cluster	Logical. When set to TRUE, <code>dsp_test</code> excludes clusters that are insufficiently represented in the dataset. Only clusters meeting the 'min_pct_cells' threshold (i.e., containing at least the specified percentage of cells across all conditions) will be retained for analysis.
verbose	Logical. If TRUE, <code>svg_test</code> returns additional components containing the underlying edgeR model object. By default (test = "QLF"), these are the 'edgeR::glmQLFTest' and 'edgeR::glmQLFit' results. When test = "LRT", the returned objects correspond to 'edgeR::glmFit' and 'edgeR::glmLRT'.

**Value**

A list containing:

- "gene\_results": a data frame with the main edgeR test results.
- "estimated\_y": a DGEList object containing the estimated common dispersion, which can be reused to speed up computation when testing individual clusters.
- "glmQLFit" (only if verbose = TRUE): a DGEGLM object containing the full statistics from 'edgeR::glmQLFit' (when test = "QLF").
- "glmQLFTest" (only if verbose = TRUE): a DGELRT-like object containing the full statistics from 'edgeR::glmQLFTest' (when test = "QLF").
- "glmFit" (only if verbose = TRUE): a DGEGLM object containing the full statistics from 'edgeR::glmFit' (when test = "LRT").
- "glmLRT" (only if verbose = TRUE): a DGELRT object containing the full statistics from 'edgeR::glmLRT' (when test = "LRT").

**See Also**

[svg\\_test](#), [individual\\_svg](#), [individual\\_dsp](#), [FeaturePlot](#), [top\\_results](#)

**Examples**

```
## Load the example multi-sample multi-group spe object
spe <- muSpaData::Wei22_example()
# Fit the model via \link{dsp_test} function.
set.seed(123)
results_dsp <- dsp_test(spe = spe,
                        cluster_col = "Banksy_smooth",
                        sample_col = "sample_id",
                        condition_col = "condition",
                        verbose = FALSE)

# dsp_test returns of an object:
# "gene_results": a dataframe contains main edgeR test results.

# We visualize differential results:
head(results_dsp, 3)
```

---

FeaturePlot

*FeaturePlot*

---

**Description**

Plot spatial gene expression. This function is a modified version of the [featurePlot](#) function from 'BayesSpace' R package. In comparison to the original BayesSpace function, this function allows plotting multiple genes simultaneously and drawing an outline around a specified cluster. To draw outlines, the [reconstructShapeDensityImage](#) function from the 'sosta' R package has been adapted. Compared to the original 'sosta' function, this version allows the use of a SingleCellExperiment object, which cannot be used with 'spatialCoords()'.

**Usage**

```

FeaturePlot(
  spe,
  feature,
  coordinates = NULL,
  concave_hull = FALSE,
  sf_dim = 200,
  assay.type = "logcounts",
  annotation_cluster = FALSE,
  annotation_title = NULL,
  platform = "Visium",
  cluster_col = NULL,
  cluster = NULL,
  legend_cluster = FALSE,
  legend_exprs = FALSE,
  diverging = FALSE,
  low = NULL,
  high = NULL,
  mid = NULL,
  color = NULL,
  linewidth = 0.4,
  linecolor = NULL,
  label = FALSE,
  ncol = 3,
  title = FALSE,
  title_size = 10,
  point_size = 0.5
)

```

**Arguments**

spe	SpatialExperiment or SingleCellExperiment. If feature is specified and is a string, it must exist as a row in the specified assay of spe.
feature	Feature vector used to color each cell. May be the name of a gene/row in an assay of spe, or a vector of continuous values.
coordinates	Column names for the spatial coordinates of cells stored in colData(spe). If specified, these coordinates will be used. If not, the function defaults to using 'row' and 'col' in colData(spe) if they exist. Otherwise, it will use spatialCoords(spe) if 'spe' is a SpatialExperiment object and spatialCoords(spe) is not NULL.
concave_hull	A logical value (TRUE or FALSE). For Visium or ST platforms, 'concave_hull' is automatically set to TRUE. If TRUE, the function uses <a href="#">geom_mark_hull</a> to outline cluster boundaries (recommended for non-discontinuous clusters). If FALSE, the adapted <a href="#">reconstructShapeDensityImage</a> is used for complex cluster shapes.
sf_dim	A numeric value for the x-dimension of the reconstruction (default is 200). A

	lower value speeds up computation but reduces accuracy. Used only when 'concave_hull' is FALSE.
assay.type	String indicating which assay in spe the expression vector should be taken from.
annotation_cluster	A logical value. TRUE or FALSE. If TRUE, annotated spatial clusters are plotted alongside expression plots. If FALSE, clusters are not displayed.
annotation_title	A character string for the title of the annotated spatial clusters. Applied only when 'annotation_cluster' is TRUE.
platform	A character string specifying the spatial sequencing platform. If "Visium" or "ST", a hexagonal spot layout will be used. Otherwise, points will be plotted.
cluster_col	Column name of spatial clusters in colData(spe).
cluster	Names of the spatial clusters used for drawing a boundary around a group of points that belong to the specify cluster. It can be NULL, "all"/"ALL", or a vector of cluster names.
legend_cluster	A logical value. TRUE or FALSE, indicating whether to plot the legend for the shaped clusters (TRUE), or not (FALSE). Only used when 'cluster_col' and 'cluster' are specified, and is supported only when 'concave_hull' is set to TRUE.
legend_exprs	A logical value. TRUE or FALSE, indicating whether to plot the legend for the expression level (TRUE), or not (FALSE).
diverging	A logical value. If TRUE, uses a diverging color gradient in <a href="#">FeaturePlot</a> (e.g., for fold change). If FALSE, uses a sequential gradient (e.g., for expression).
low, mid, high	Optional hex codes for low, mid, and high values of the color gradient used for continuous cell values.
color	Optional hex code to set color of borders around cells. Set to NA to remove borders.
linewidth	The width of the boundary line around the cluster. The default ('0.4') size of the boundary line is one.
linecolor	The colors of the boundary lines around the cluster. If unspecified, the default color scheme is used.
label	A logical. TRUE or FALSE. Adding a label and an arrow pointing to a group.
ncol	The dimensions of the grid to create. By default, 1, if the length of feature equals to 1, and 3, otherwise.
title	A logical. TRUE or FALSE. If true, the title name of each (subplot) is the gene name.
title_size	Title font size.
point_size	Point size.

**Value**

Returns a ggplot object.

**See Also**

[svg\\_test](#), [individual\\_svg](#), [top\\_results](#), [dsp\\_test](#), [individual\\_dsp](#)

**Examples**

```
# load the input data:
data("LIBD_subset", package = "DESpace")

# load pre-computed results (obtained via `svg_test`)
data("results_svg_test", package = "DESpace")

# Visualize the gene expression of the top three genes
feature = results_svg_test$gene_results$gene_id[seq_len(3)]
FeaturePlot(LIBD_subset, feature, coordinates = c("array_row", "array_col"),
            ncol = 3, title = TRUE)
```

---

individual_dsp	<i>individual_dsp</i>	
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---

**Description**

DESpace can also be used to reveal the specific areas of the tissue affected by DSP genes; i.e., spatial clusters that are particularly over/under abundant compared to the average signal across conditions. This function can be used to identify SVGs among conditions for each individual cluster.

**Usage**

```
individual_dsp(
  spe,
  cluster_col,
  sample_col,
  condition_col,
  test = "QLF",
  min_counts = 20,
  min_non_zero_spots = 10,
  min_pct_cells = 0.5,
  filter_gene = TRUE,
  filter_cluster = TRUE
)
```

**Arguments**

<code>spe</code>	SpatialExperiment or SingleCellExperiment.
<code>cluster_col</code>	Character. Column name of spatial clusters in <code>colData(spe)</code> .
<code>sample_col</code>	Character. Column name of sample ids in <code>colData(spe)</code> . Sample ids must be either a factor or character.

condition_col	Character. Column name of condition ids in colData(spe).
test	Character. Either 'QLF' or 'LRT'. Default is 'QLF'. Specifies whether to perform quasi-likelihood F-tests ('edgeR::glmQLFTest') or likelihood ratio tests ('edgeR::glmLRT'). Quasi-likelihood tests apply empirical Bayes moderation to genewise dispersions, account for intra-sample variability, and provide more conservative error control.
min_counts	Numeric. Minimum number of counts per sample (across all spots) for a gene to be analyzed.
min_non_zero_spots	Numeric. Minimum number of non-zero spots per sample, for a gene to be analyzed.
min_pct_cells	Numeric. Minimum percentage of cells required for each cluster to be included in the analysis across the specified conditions. Default value is 0.5 (i.e., 0.5% of total cells per cluster per condition).
filter_gene	Logical. If TRUE, <a href="#">dsp_test</a> filters genes: genes have to be expressed in at least 'min_non_zero_spots' spots, and a gene requires at least 'min_counts' counts per sample (across all locations).
filter_cluster	Logical. When set to TRUE, <a href="#">dsp_test</a> excludes clusters that are insufficiently represented in the dataset. Only clusters meeting the 'min_pct_cells' threshold (i.e., containing at least the specified percentage of cells across all conditions) will be retained for analysis.

### Value

A list of results, with one result per spatial cluster in each element. Specifically, each item in the list is a "gene\_results" dataframe which contains main edgeR test results.

### See Also

[top\\_results](#), [svg\\_test](#), [dsp\\_test](#), [FeaturePlot](#)

### Examples

```
# load the input data:
spe <- muSpaData::Wei22_example()
set.seed(123)
results_individual_dsp <- individual_dsp(spe,
                                         cluster_col = "Banksy_smooth",
                                         sample_col = "sample_id",
                                         condition_col = "condition")

# We visualize results for the cluster '3'
results <- results_individual_dsp[['3']]
head(results,3)
```

---

individual\_svg            *individual\_svg*

---

### Description

DESpace can also be used to reveal the specific areas of the tissue affected by SVGs; i.e., spatial clusters that are particularly over/under abundant compared to the average signal. This function can be used to identify SVGs for each individual cluster.

### Usage

```
individual_svg(
  spe,
  cluster_col,
  sample_col = "sample_id",
  edgeR_y = NULL,
  min_counts = 20,
  min_non_zero_spots = 10,
  filter_gene = TRUE,
  replicates = FALSE,
  BPPARAM = NULL
)
```

### Arguments

spe	SpatialExperiment or SingleCellExperiment.
cluster_col	Column name of spatial clusters in colData(spe).
sample_col	Column name of sample ids in colData(spe).
edgeR_y	Pre-estimated dispersion; if it's null, compute dispersion.
min_counts	Minimum number of counts per sample (across all spots) for a gene to be analyzed.
min_non_zero_spots	Minimum number of non-zero spots per sample, for a gene to be analyzed.
filter_gene	A logical. If TRUE, <code>svg_test</code> filters genes: genes have to be expressed in at least 'min_non_zero_spots' spots, and a gene requires at least 'min counts' counts per sample (across all locations).
replicates	Single sample or multi-sample test.
BPPARAM	An optional parameter passed internally to <code>bplapply</code> . We suggest using as many cores as the number of spatial clusters. If unspecified, the script does not run in parallel. Note that parallel coding performs better only when dispersion estimations are not provided beforehand. Moreover, parallelizing the script will increase the memory requirement; if memory is an issue, leave 'BPPARAM' unspecified and, hence, avoid parallelization.

## Details

For every spatial cluster we test, edgeR would normally re-compute the dispersion estimates based on the specific design of the test. However, this calculation represents the majority of the overall computing time. Therefore, to speed-up calculations, we propose to use the dispersion estimates which were previously computed for the gene-level tests. This introduces a minor approximation which, in our benchmarks, does not lead to decreased accuracy. If you want to use pre-computed gene-level dispersion estimates, set `edgeR_y` to `'estimated_y'`. Alternatively, if you want to re-compute dispersion estimates (significantly slower, but marginally more accurate option), leave `edgeR_y` empty.

## Value

A list of results, with one result per spatial cluster in each element. Specifically, each item in the list is a "gene\_results" dataframe which contains main edgeR test results.

## See Also

[top\\_results](#), [svg\\_test](#), [FeaturePlot](#)

## Examples

```
# load the input data:
data("LIBD_subset", package = "DESpace")
LIBD_subset

# load pre-computed results (obtained via `svg_test`)
data("results_svg_test", package = "DESpace")

# svg_test returns of a list of 2 objects:
# "gene_results": a dataframe contains main edgeR test results;
# "estimated_y": a DGEList object contains the estimated common dispersion,
# which can later be used to speed-up calculation when testing individual clusters.

# We visualize differential results:
head(results_svg_test$gene_results, 3)

# Individual cluster test: identify SVGs for each individual cluster
# set parallel computing; we suggest using as many cores as the number of spatial clusters.
# Note that parallelizing the script will increase the memory requirement;
# if memory is an issue, leave 'BPPARAM' unspecified and, hence, avoid parallelization.
set.seed(123)
results_individual_svg <- individual_svg(LIBD_subset,
                                         edgeR_y = results_svg_test$estimated_y,
                                         cluster_col = "layer_guess_reordered")

# We visualize results for the cluster 'WM'
results_WM <- results_individual_svg[[7]]
head(results_WM, 3)
```

---

LIBD_subset	<i>Subset from the human DLPFC 10x Genomics Visium dataset of the spatialLIBD package</i>
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---

## Description

Subset from the human DLPFC 10x Genomics Visium dataset of the spatialLIBD package

## Arguments

LIBD\_subset contains a [SpatialExperiment-class](#) object, representing a subset of the sample 151673 from the full real data of the spatialLIBD package. Below the code used to subset the original dataset.

## Value

A spatial experiment object

## Author(s)

Peiyong Cai <peiyong.cai@uzh.ch>, Simone Tiberi <simone.tiberi@unibo.it>

## See Also

[svg\\_test](#), [individual\\_svg](#)

## Examples

```
# Connect to ExperimentHub
# ehub <- ExperimentHub::ExperimentHub()
# Download the example spe data
# spe_all <- spatialLIBD::fetch_data(type = "spe", eh = ehub)
# Select one sample only:
# LIBD_subset <- spe_all[, colData(spe_all)$sample_id == '151673']
# Select small set of random genes for faster runtime
# set.seed(123)
# sel_genes <- sample(dim(LIBD_subset)[1],500)
# LIBD_subset <- LIBD_subset[sel_genes,]
# keep_col <- c("array_row","array_col","layer_guess_reordered")
# library(SingleCellExperiment)
# LIBD_subset <- SpatialExperiment(assay = list(counts = assay(LIBD_subset),
#                                           logcounts = logcounts(LIBD_subset)),
#                                colData = colData(LIBD_subset)[keep_col])
# save(LIBD_subset, file = "./DESpace/data/LIBD_subset.RData")
```

---

results\_individual\_svg

*Results from [individual\\_svg](#) function*

---

## Description

Results from [individual\\_svg](#) function

## Arguments

results\_individual\_svg

contains a [list](#) object, with the results obtained applying [individual\\_svg](#) function to an external dataset from the [spatialLIBD](#) package. Below the code used to obtain 'results\_individual\_svg'.

## Value

A List of 7 elements - one element for each spatial cluster

## Author(s)

Peiyong Cai <[peiyong.cai@uzh.ch](mailto:peiyong.cai@uzh.ch)>, Simone Tiberi <[simone.tiberi@unibo.it](mailto:simone.tiberi@unibo.it)>

## See Also

[svg\\_test](#), [individual\\_svg](#)

## Examples

```
# load the input data:
# data("LIBD_subset", package = "DESpace")
# LIBD_subset
# load pre-computed results (obtained via `svg_test`)
# data("results_svg_test", package = "DESpace")
# results_svg_test

# Function `individual_svg()` can be used to identify SVGs for each individual cluster.
# Parameter `spatial_cluster` indicates the column names of `colData(spe)`
# containing spatial clusters.
# set.seed(123)
# results_individual_svg <- individual_svg(LIBD_subset,
#                                         edgeR_y = results_svg_test$estimated_y,
#                                         spatial_cluster = "layer_guess_reordered")
# save(results_individual_svg, file = "./DESpace/data/results_individual_svg.RData")
```

---

results\_svg\_test      *Results from [svg\\_test](#) function*

---

### Description

Results from [svg\\_test](#) function

### Arguments

results\_svg\_test

contains a [list](#) object, with the results obtained applying [svg\\_test](#) function to an external dataset from the [spatialLIBD](#) package. Below the code used to obtain 'results\_svg\_test'.

### Value

Large List of 2 elements:

- "gene\_results": a dataframe contains main edgeR test results;
- "estimated\_y": a DGEList object contains the estimated common dispersion,

### Author(s)

Peiyong Cai <peiyong.cai@uzh.ch>, Simone Tiberi <simone.tiberi@unibo.it>

### See Also

[svg\\_test](#)

### Examples

```
# load the input data:
# data("LIBD_subset", package = "DESpace")
# LIBD_subset
#
# Fit the model via `svg_test` function.
# Parameter `spe` specifies the input `SpatialExperiment` or `SingleCellExperiment` object,
# while `cluster_col` defines the column names of `colData(spe)` containing spatial clusters.
# To obtain all statistics, set `verbose` to `TRUE`.
#
# set.seed(123)
# results_svg_test <- svg_test(spe = LIBD_subset,
#                             cluster_col = "layer_guess_reordered",
#                             verbose = FALSE)
#
# save(results_svg_test, file = "./DESpace/data/results_svg_test.RData")
```

---

svg_test	<i>svg_test</i>
----------	-----------------

---

## Description

'svg\_test' identifies spatially variable genes (SVGs) from spatially-resolved transcriptomics data, provided spatial clusters are available.

## Usage

```
svg_test(
  spe,
  cluster_col,
  sample_col = NULL,
  replicates = FALSE,
  min_counts = 20,
  min_non_zero_spots = 10,
  filter_gene = TRUE,
  verbose = FALSE
)
```

## Arguments

spe	SpatialExperiment or SingleCellExperiment.
cluster_col	Column name of spatial clusters in colData(spe).
sample_col	Column name of sample ids in colData(spe).
replicates	A logical, indicating whether biological replicates are provided (TRUE) or not (FALSE). If biological replicates are provided, <code>svg_test</code> performs a joint test across all replicates, searching for SVGs with consistent spatial patterns across samples.
min_counts	Minimum number of counts per sample (across all spots) for a gene to be analyzed.
min_non_zero_spots	Minimum number of non-zero spots per sample, for a gene to be analyzed.
filter_gene	A logical. If TRUE, <code>svg_test</code> filters genes: genes have to be expressed in at least 'min_non_zero_spots' spots, and a gene requires at least 'min counts' counts per sample (across all locations).
verbose	A logical. If TRUE, <code>svg_test</code> returns two more results: 'DGEGLM' and 'DGELRT' objects contain full statistics from 'edgeR::glmFit' and 'edgeR::glmLRT'.

## Details

If 'sample\_col' is not specified and 'replicates == FALSE', `svg_test` assumed that data comes from an individual sample, and performs SV testing on it.

If 'sample\_col' is provided and 'replicates == FALSE', `svg_test` tests each sample individually and returns a list of results for each sample.

If 'sample\_col' is provided and 'replicates == TRUE', `svg_test` performs a joint multi-sample test.

### Value

A list of results:

- "gene\_results": a dataframe contains main edgeR test results;
- "estimated\_y": a DGEList object contains the estimated common dispersion, which can later be used to speed-up calculation when testing individual clusters.
- "glmFit" (only if verbose = TRUE): a DGEGLM object contains full statistics from "edgeR::glmFit".
- "glmLRT" (only if verbose = TRUE): a DGELRT object contains full statistics from "edgeR::glmLRT".

### See Also

[top\\_results](#), [individual\\_svg](#), [FeaturePlot](#), [dsp\\_test](#), [individual\\_dsp](#)

### Examples

```
# load the input data:
data("LIBD_subset", package = "DESpace")
LIBD_subset

# Fit the model via \code{\link{svg_test}} function.
set.seed(123)
results_svg_test <- svg_test(spe = LIBD_subset,
                           cluster_col = "layer_guess_reordered",
                           verbose = FALSE)

# svg_test returns of a list of 2 objects:
# "gene_results": a dataframe contains main edgeR test results;
# "estimated_y": a DGEList object contains the estimated common dispersion,
# which can later be used to speed-up calculation when testing individual clusters.

# We visualize differential results:
head(results_svg_test$gene_results, 3)
```

---

top\_results

*top\_results*

---

### Description

Filter significant results. `top_results` returns the significant results obtained via `svg_test` and `individual_svg`. It can also be used to merge gene- and cluster-level results into a single object.

**Usage**

```
top_results(
  gene_results = NULL,
  cluster_results,
  cluster = NULL,
  select = "both",
  high_low = NULL
)
```

**Arguments**

gene_results	Results returned from <a href="#">svg_test</a> .
cluster_results	Results returned from <a href="#">individual_svg</a> .
cluster	A character indicating the cluster(s) whose results have to be returned. Results from all clusters are returned by default ("NULL").
select	A character indicating what results should be returned ("FDR", "logFC", or "both"). Only used if "cluster_results" are provided. By default ("both"), both FDR and logFC are returned.
high_low	A character indicating whether to filter results or not. Only used if "cluster_results" are provided, and one cluster is specified in "cluster" parameter. By default (NULL), all results are returned in a single data.frame. If "high" or "HIGH", we only return SVGs with average abundance in "cluster" higher than in the rest of the tissue (i.e., logFC > 0). If "low" or "LOW", we only return SVGs with average abundance in "cluster" lower than in the rest of the tissue (i.e., logFC < 0). If "both" or "BOTH", then both "high" and "low" results are returned, but in two separate data.frames.

**Value**

A [data.frame](#) object or a list of [data.frame](#) with results.

- When only "cluster\_results" is provided, results are reported as a [data.frame](#) with columns for gene names (gene\_id), spatial clusters affected by SV (Cluster), cluster-specific likelihood ratio test statistics (LR), cluster-specific average (across spots) log-2 counts per million (logCPM), cluster-specific log2-fold changes (logFC), cluster-specific raw p-values (PValue), and Benjamini-Hochberg adjusted p-values (FDR) for each spatial cluster.
- When "gene\_results" and "cluster\_results" are given, results are reported as a [data.frame](#) that merges gene- and cluster-level results.
- If "cluster" is specified, the function returns a subset [data.frame](#) for the given cluster, which contains cluster name, gene name, LR, logCPM, logFC, PValue and FDR, ordered by FDR for the specified cluster.
- If "high\_low" is set, the function returns a list of [data.frame](#) that contains subsets of results for genes with higher and/or lower expression in the given cluster compared to the rest of the tissue.

**See Also**

[svg\\_test](#), [individual\\_svg](#), [FeaturePlot](#), [dsp\\_test](#), [individual\\_dsp](#)

**Examples**

```
# load pre-computed results (obtained via `svg_test`)
data("results_svg_test", package = "DESpace")

# svg_test returns of a list of 2 objects:
# "gene_results": a dataframe contains main edgeR test results;
# "estimated_y": a DGEList object contains the estimated common dispersion,
# which can later be used to speed-up calculation when testing individual clusters.

# We visualize differential results:
head(results_svg_test$gene_results, 3)

# load pre-computed results (obtained via `individual_svg`)
data("results_individual_svg", package = "DESpace")
# Function `individual_svg()` can be used to identify SVGs for each individual cluster.
# `individual_svg()` returns a list containing the results of individual clusters.
# For each cluster, results are reported as a data.frame,
# where columns for each cluster, results are reported as a data.frame,
# where columns contain gene names (`genes`), likelihood ratio (`LR`),
# log2-fold changes (`logFC`) and adjusted p-value (`FDR`).
#
# Combine gene-and cluster-level results
merge_res = top_results(results_svg_test$gene_results,
                        results_individual_svg)

head(merge_res,3)
# 'select = "FDR"' can be used to visualize adjusted p-values for each spatial cluster.
merge_res = top_results(results_svg_test$gene_results,
                        results_individual_svg, select = "FDR")

head(merge_res,3)
# Specify the cluster of interest and check top genes detected by svg_test.
results_WM_both = top_results(cluster_results = results_individual_svg,
                              cluster = "WM", high_low = "both")

head(results_WM_both$high_genes, 3)
head(results_WM_both$low_genes, 3)
```

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