

Package: ROTS (via r-universe)

September 29, 2026

Type Package

Title Reproducibility-Optimized Test Statistic

Version 2.4.0

Author Fatemeh Seyednasrollah, Tomi Suomi, Laura L. Elo

Maintainer Tomi Suomi <tomi.suomi@utu.fi>

Description Calculates the Reproducibility-Optimized Test Statistic (ROTS) for differential testing in omics data.

biocViews Software, GeneExpression, DifferentialExpression, Microarray, RNASeq, Proteomics, ImmunoOncology

License GPL (>= 2)

LazyData yes

LinkingTo Rcpp

Imports Rcpp, stats, Biobase, methods, BiocParallel, lme4, survival

Suggests testthat

Depends R (>= 3.6)

NeedsCompilation yes

Config/pak/sysreqs cmake make

Repository Bioconductor 3.23

Date/Publication 2026-04-28 12:42:34 UTC

RemoteUrl <https://github.com/bioc/ROTS>

RemoteRef RELEASE_3_23

RemoteSha a2b50206bab429a357ec81854c6a8226a78ceaba

Contents

affySpikeIn	2
coxphROTS	2
lmeROTS	4
lmROTS	5

plot.regROTS	6
plot.ROTS	7
results	8
ROTS	9
summary.regROTS	11
summary.ROTS	12
upsSpikeIn	13
Index	15

affySpikeIn	<i>Gene expression data from the Affymetrix spike-in experiment</i>
-------------	---

Description

The Affymetrix HG-U95Av2 spike-in data contains two groups of 12 samples. In this carefully controlled experiment, it is known that 14 genes are differentially expressed between the two sample groups, while the rest of the genes are equally expressed. In order to reduce the running time of the vignette example, this package uses a subset of 1000 genes of the original dataset including all the spiked genes with 5 samples from each group. The preprocessed RMA-normalized data were obtained from the Bioconductor 2.2 package DEBS. For more details about the spike-in experiment, see <http://www.affymetrix.com>.

Usage

affySpikeIn

Format

- affySpikeIn a numeric matrix of gene expression levels with 12626 rows (genes) and 24 columns (samples).
- affySpikeIn.L a vector indicating the sample groups.
- affySpikeIn.gnames a character vector containing the names of the genes.
- spikedgene a numeric vector indicating the locations of the 14 spiked genes.

coxphROTS	<i>Reproducibility-Optimized Test Statistic (ROTS)</i>
-----------	--

Description

Calculates the reproducibility-optimized test statistic (ROTS) for ranking genes in order of evidence for differential expression using Cox proportional hazards regression model.

Usage

coxphROTS(formula, data, metadata, B = 100, K = NULL, seed = NULL, a1 = NULL, a2 = NULL, terms = NULL, BPPARAM = bpparam())

Arguments

formula	A string representing a formula that follows the <code>coxph</code> syntax.
data	A numeric data matrix in which rows correspond to genes and columns correspond to samples.
metadata	A numeric data matrix in which rows correspond to samples and columns correspond to explanatory variables.
B	An integer specifying the number of bootstrap and permutation resamplings (default 100).
K	An integer indicating the largest top list size considered. If no value is given, 1/4 of the features are used.
seed	An integer seed for the random number generator.
a1, a2	Non-negative parameters.
terms	Terms to be optimized (default all).
BPPARAM	BiocParallel back-end if other than default.

Details

The reproducibility-optimization procedure ROTS enables the selection of a suitable gene ranking statistic directly from the given dataset. This function allows the use of Cox proportional hazards model as a basis for reproducibility-optimization procedure.

Value

`coxphROTS` returns an object of class `regROTS`, which is a list of ROTS results for each of the variables in the formula. They contain the following components:

d	the value of the optimized ROTS-statistic for each gene.
coef	coefficients of the model for each gene.
pvalue	the corresponding p-values.
FDR	the corresponding FDR-values.
a1	the optimized parameter a1.
a2	the optimized parameter a2.
k	the optimized top list size.
R	the optimized reproducibility value.
Z	the optimized reproducibility Z-score.
ztable	the full reproducibility table.

Author(s)

Tomi Suomi, Laura L. Elo
 Maintainer: Tomi Suomi <tomi.suomi@utu.fi>

Examples

```
## ROTS-statistic
## Not run: rots.out <- coxphROTS(Surv(time, event) ~ exprs, data, metadata)
```

lmeROTS

*Reproducibility-Optimized Test Statistic (ROTS)***Description**

Calculates the reproducibility-optimized test statistic (ROTS) for ranking genes in order of evidence for differential expression using linear mixed-effects model

Usage

```
lmeROTS(formula, data, metadata, B = 100, K = NULL, seed = NULL,
a1 = NULL, a2 = NULL, terms = NULL, BPPARAM = bpparam())
```

Arguments

formula	A string representing a formula that follows the lme4 syntax.
data	A numeric data matrix in which rows correspond to genes and columns correspond to samples.
metadata	A numeric data matrix in which rows correspond to samples and columns correspond to explanatory variables.
B	An integer specifying the number of bootstrap and permutation resamplings (default 100).
K	An integer indicating the largest top list size considered. If no value is given, 1/4 of the features are used.
seed	An integer seed for the random number generator.
a1, a2	Non-negative parameters.
terms	Terms to be optimized (default all).
BPPARAM	BiocParallel back-end if other than default.

Details

The reproducibility-optimization procedure ROTS enables the selection of a suitable gene ranking statistic directly from the given dataset. This function allows the use of linear mixed-effects models as a basis for reproducibility-optimization procedure.

Value

lmeROTS returns an object of class regROTS, which is a list of ROTS results for each of the variables in the formula. They contain the following components:

d	the value of the optimized ROTS-statistic for each gene.
coef	coefficients of the model for each gene.
pvalue	the corresponding pvalues.
FDR	the corresponding FDR-values.

a1	the optimized parameter a1.
a2	the optimized parameter a2.
k	the optimized top list size.
R	the optimized reproducibility value.
Z	the optimized reproducibility Z-score.
ztable	the full reproducibility table.

Author(s)

Tomi Suomi, Laura L. Elo
 Maintainer: Tomi Suomi <tomi.suomi@utu.fi>

Examples

```
## ROTS-statistic
## Not run: rots.out <- lmROTS(exprs ~ Group + Age + (1|Individual), data, metadata)
```

lmROTS	<i>Reproducibility-Optimized Test Statistic (ROTS)</i>
--------	--

Description

Calculates the reproducibility-optimized test statistic (ROTS) for ranking genes in order of evidence for differential expression using linear model

Usage

```
lmROTS(formula, data, metadata, B = 100, K = NULL, seed = NULL,
a1 = NULL, a2 = NULL, terms = NULL, BPPARAM = bpparam())
```

Arguments

formula	A string representing a formula that follows the <code>lm</code> syntax.
data	A numeric data matrix in which rows correspond to genes and columns correspond to samples.
metadata	A numeric data matrix in which rows correspond to samples and columns correspond to explanatory variables.
B	An integer specifying the number of bootstrap and permutation resamplings (default 100).
K	An integer indicating the largest top list size considered. If no value is given, 1/4 of the features are used.
seed	An integer seed for the random number generator.
a1, a2	Non-negative parameters.
terms	Terms to be optimized (default all).
BPPARAM	BiocParallel back-end if other than default.

Details

The reproducibility-optimization procedure ROTS enables the selection of a suitable gene ranking statistic directly from the given dataset. This function allows the use of linear models as a basis for reproducibility-optimization procedure.

Value

lmeROTS returns an object of class regROTS, which is a list of ROTS results for each of the variables in the formula. They contain the following components:

d	the value of the optimized ROTS-statistic for each gene.
coef	coefficients of the model for each gene.
pvalue	the corresponding pvalues.
FDR	the corresponding FDR-values.
a1	the optimized parameter a1.
a2	the optimized parameter a2.
k	the optimized top list size.
R	the optimized reproducibility value.
Z	the optimized reproducibility Z-score.
ztable	the full reproducibility table.

Author(s)

Tomi Suomi, Laura L. Elo
 Maintainer: Tomi Suomi <tomi.suomi@utu.fi>

Examples

```
## ROTS-statistic
## Not run: rots.out <- lmeROTS(exprs ~ Group + Age, data, metadata)
```

plot.regROTS	<i>Plotting of a regROTS object</i>
--------------	-------------------------------------

Description

Plots the regROTS objects created with the ROTS package.

Usage

```
## S3 method for class 'regROTS'
plot(x, fdr=0.05, type="reproducibility", features=NULL, labels=FALSE, ...)
```

Arguments

x	A regROTS object created from differential expression testing run by lmeROTS or lmROTS.
fdr	Selected cutoff for FDR value.
type	Type of plot to be created. Options are: 'volcano', 'reproducibility', 'pvalue'.
features	Option to plot only selected variables from the output.
labels	Option to print labels for differentially expressed features.
...	Other arguments passed to the plot function.

Details

This function plots the results from a ROTS object using given false discovery rate threshold.

Value

Plots the results from ROTS object.

Author(s)

Tomi Suomi, Laura L. Elo
 Maintainer: Tomi Suomi <tomi.suomi@utu.fi>

Examples

```
## Run analysis and plot ROTS object
## Not run: rots.out <- lmROTS(exprs ~ Group + Age, data, metadata)
## Not run: plot(rots.out, type="reproducibility")
```

plot.ROTS	<i>Plotting of a ROTS object</i>
-----------	----------------------------------

Description

Plots the ROTS objects created with the ROTS package.

Usage

```
## S3 method for class 'ROTS'
plot(x, type="reproducibility", fdr=0.05, labels=FALSE, ...)
```

Arguments

x	A ROTS object created from differential expression testing run by ROTS.
fdr	Selected cutoff for FDR value.
type	Type of plot to be created. Options are: 'volcano', 'heatmap', 'ma', 'reproducibility', 'pvalue', 'pca'.
labels	Option to print labels for differentially expressed features.
...	Other arguments passed to the plot function.

Details

This function plots the results from a ROTS object using given false discovery rate threshold.

Value

Plots the results from ROTS object.

Author(s)

Fatemeh Seyednasrollah, Tomi Suomi, Laura L. Elo

Maintainer: Tomi Suomi <tomi.suomi@utu.fi>

See Also

[affySpikeIn](#)

Examples

```
## ROTS-statistic for the Affymetrix spike-in data.
rots.out <- ROTS(data = affySpikeIn, groups = c(rep(0,5), rep(1,5)),
  B = 100, K = 500 , seed = 1234)
## Plotting of the ROTS results.
plot(rots.out, type="volcano")
```

results

Gathering results from a regROTS object

Description

Gathers the results from regROTS objects created with the ROTS package.

Usage

```
results(object, order=FALSE, pvalue=NULL, FDR=NULL, coef=NULL, logfc=NULL)
```

Arguments

object	A regROTS object created from differential expression testing run by lmeROTS or lmROTS.
order	Ordering of results based on significance (default FALSE).
pvalue	Optional p-value cutoff for filtering the results.
FDR	Optional false discovery rate cutoff for filtering the results.
coef	Optional model coefficient cutoff for filtering the results (regROTS).
logfc	Optional log fold change cutoff for filtering the results (ROTS).

Details

This is a convenience function to gather results from regROTS objects.

Value

A list of reproducibility-optimized results for each variable in the model.

Author(s)

Tomi Suomi, Laura L. Elo

Maintainer: Tomi Suomi <tomi.suomi@utu.fi>

Examples

```
## Run and plot lmeROTS-statistic
## Not run: rots.out <- lmeROTS("Group + Age + (1|Individual)", data, metadata)
## Not run: results <- results(rots.out)
```

ROTS

Reproducibility-Optimized Test Statistic (ROTS)

Description

Calculates the reproducibility-optimized test statistic (ROTS) for ranking genes in order of evidence for differential expression in two-group or multi-group comparisons.

Usage

```
ROTS(data, groups, B = 1000, K = NULL, paired = FALSE, seed = NULL,
a1 = NULL, a2 = NULL, log = TRUE, progress = FALSE, verbose = TRUE,
time = NULL, event = NULL)
```

Arguments

data	A numeric data matrix or an ExpressionSet instance, in which rows correspond to genes and columns correspond to samples.
groups	A vector indicating the sample groups.
B	An integer specifying the number of bootstrap and permutation resamplings (default 1000).
K	An integer indicating the largest top list size considered. If no value is given, 1/4 of the features are used.
paired	A logical indicating whether a paired test is performed. The samples are expected to be in the same order in both groups.
seed	An integer seed for the random number generator.
a1, a2	Non-negative parameters. See details section for further information.

log	A logical (default TRUE) indicating whether input data is log2 scaled. This information is only used to calculate log fold change.
progress	A logical indicating if additional progress bars are shown.
verbose	A logical indicating if messages are shown.
time	Time to event (for survival data).
event	Censoring information (for survival data): 1=event, 0=censored.

Details

The reproducibility-optimization procedure ROTS enables the selection of a suitable gene ranking statistic directly from the given dataset. In two-group comparisons, the statistic is optimized among a family of t-type statistics $d = m/(a_1 + a_2 * s)$, where m is the absolute difference between the group averages, s is the pooled standard error, and a_1 and a_2 are the non-negative parameters to be optimized. Two special cases of this family are the ordinary t-statistic ($a_1=0$, $a_2=1$) and the signal log-ratio ($a_1=1$, $a_2=0$). The optimality is defined in terms of maximal overlap of top-ranked genes in group-preserving bootstrap datasets. Importantly, besides the group labels, no a priori information about the properties of the data is required and no fixed cutoff for the gene rankings needs to be specified. For more details about the reproducibility-optimization procedure, see Elo et al. (2008).

The user is given the option to adjust the largest top list size considered in the reproducibility calculations, since lowering this size can markedly reduce the computation time. In large data matrices with thousands of rows, we generally recommend using a size of several thousands. In smaller data matrices, and especially if there are many rows with only a few non-missing entries, the size of K should be decreased accordingly.

ROTS tolerates a moderate number of missing values in the data matrix by effectively ignoring their contribution during the operation of the procedure. However, each row of the data matrix must contain at least two values in both groups. The rows containing only a few non-missing values should be removed; or alternatively, the missing data entries can be imputed using, e.g., the K-nearest neighbors imputation, which is implemented in the Bioconductor package `impute`. ROTS assumes the input data matrix is log2 transformed (the default for log parameter is set to TRUE). Although, this only affects fold change values, we recommend setting log parameter to FALSE if the input matrix is not log transformed to avoid downstream confusions.

If the parameter values a_1 and a_2 are set by the user, then no optimization is performed but the statistic and FDR-values are calculated for the given parameters. The false discovery rate (FDR) for the optimized test statistic is calculated by permuting the sample labels. The results for all the genes can be obtained by setting the FDR cutoff to 1.

Value

ROTS returns an object of class ROTS, which is a list containing the following components

data	the expression data matrix.
B	the number of bootstrap and permutation resamplings.
d	the value of the optimized ROTS-statistic for each gene.
pvalue	the corresponding pvalues.
FDR	the corresponding FDR-values.

a1	the optimized parameter a1.
a2	the optimized parameter a2.
k	the optimized top list size.
R	the optimized reproducibility value.
Z	the optimized reproducibility Z-score.

print prints the optimized parameters a1 and a2, the optimized top list size and the corresponding reproducibility values.

summary summarizes the results of a ROTS analysis. If fdr and num.genes are not specified, then the optimized parameters a1 and a2, the optimized top list size and the corresponding reproducibility values are shown. If fdr or num.genes is specified, then also the gene-specific information is shown for the genes at the specified FDR-level or top list size, respectively.

Author(s)

Fatemeh Seyednasrollah, Tomi Suomi, Laura L. Elo
 Maintainer: Tomi Suomi <tomi.suomi@utu.fi>

References

Suomi T, Seyednasrollah F, Jaakkola MK, Faux T, Elo LL.
 ROTS: An R package for reproducibility-optimized statistical testing.
 PLoS Comput Biol 2017; 13: e1005562.

See Also

[affySpikeIn](#)

Examples

```
## ROTS-statistic for the Affymetrix spike-in data.
rots.out <- ROTS(data = affySpikeIn, groups = c(rep(0,5), rep(1,5)),
  B = 100, K = 500 , seed = 1234)
## Summary of the ROTS results.
rots.summary <- summary(rots.out, fdr = 0.05)
```

summary.regROTS	<i>Summary of a regROTS object</i>
-----------------	------------------------------------

Description

Summarizes the differential testing results from ROTS package.

Usage

```
## S3 method for class 'regROTS'
summary(object, fdr=0.05, ...)
```

Arguments

object	A regROTS object created from differential expression testing run by ROTS.
fdr	Selected cutoff for FDR value.
...	Other arguments passed to the summary function.

Details

This function returns the summary information of regROTS object.

Value

Summary of reproducibility-optimisation.

Author(s)

Tomi Suomi, Laura L. Elo
 Maintainer: Tomi Suomi <tomi.suomi@utu.fi>

Examples

```
## Run and get summary of lmeROTS-statistic
## Not run: rots.out <- lmeROTS("Group + Age + (1|Individual)", data, metadata)
## Not run: summary(rots.out)
```

summary.ROTS	<i>Summary of a ROTS object</i>
--------------	---------------------------------

Description

Summarizes the differential testing results from ROTS package.

Usage

```
## S3 method for class 'ROTS'
summary(object, fdr=NULL, num.genes=NULL, verbose=TRUE, ...)
```

Arguments

object	a ROTS object created from differential expression testing run by ROTS.
fdr	selected cutoff for FDR value.
num.genes	selected cutoff number for number of differentially detected features.
verbose	If TRUE (default), summary function will print out 10 first detections which fulfill the cutoff criteria.
...	other arguments passed to the summary function.

Details

This function returns the summary information (including row number, test-statistic, pvalue and FDR value) for the selected features.

Value

Returns a matrix where the rows are the selected features and columns are the Row number, ROTS-statistic, pvalue and FDR.

Author(s)

Fatemeh Seyednasrollah, Tomi Suomi, Laura L. Elo
Maintainer: Tomi Suomi <tomi.suomi@utu.fi>

References

Suomi T, Seyednasrollah F, Jaakkola MK, Faux T, Elo LL.
ROTS: An R package for reproducibility-optimized statistical testing.
PLoS Comput Biol 2017; 13: e1005562.

See Also

[affySpikeIn](#)

Examples

```
## ROTS-statistic for the Affymetrix spike-in data.  
rots.out <- ROTS(data = affySpikeIn, groups = c(rep(0,5), rep(1,5)),  
  B = 100, K = 500 , seed = 1234)  
## Summary of the ROTS results.  
rots.summary <- summary(rots.out, fdr = 0.05)
```

upsSpikeIn

*Protein expression data from the CPTAC Technology Assessment
(Study 6)*

Description

The data contains preprocessed expression values from CPTAC Technology Assessment, where equimolar concentrations of 48 human proteins (Sigma UPS 1) are spiked in five different concentrations into yeast proteome background and then processed. The example data contains only two groups from the study 6.

Usage

upsSpikeIn

Format

upsSpikeIn A numeric matrix of protein expressions.

Index

* datasets

affySpikeIn, [2](#)

upsSpikeIn, [13](#)

* math

coxphROTS, [2](#)

lmeROTS, [4](#)

lmROTS, [5](#)

plot.regROTS, [6](#)

plot.ROTS, [7](#)

results, [8](#)

ROTS, [9](#)

summary.regROTS, [11](#)

summary.ROTS, [12](#)

affySpikeIn, [2](#), [8](#), [11](#), [13](#)

coxphROTS, [2](#)

lmeROTS, [4](#)

lmROTS, [5](#)

plot.regROTS, [6](#)

plot.ROTS, [7](#)

print.ROTS (ROTS), [9](#)

results, [8](#)

ROTS, [9](#)

ROTS-package (ROTS), [9](#)

spikedgene (affySpikeIn), [2](#)

summary.regROTS, [11](#)

summary.ROTS, [12](#)

upsSpikeIn, [13](#)