Package: VaSP (via r-universe)

November 25, 2026

Type Package

Version 1.26.0

Title Quantification and Visualization of Variations of Splicing in Population

Description Discovery of genome-wide variable alternative splicing events from short-read RNA-seq data and visualizations of gene splicing information for publication-quality multi-panel figures in a population. (Warning: The visualizing function is removed due to the dependent package Sushi deprecated. If you want to use it, please change back to an older version.)

URL https://github.com/yuhuihui2011/VaSP

BugReports https://github.com/yuhuihui2011/VaSP/issues

License GPL (>= 2.0)

Depends R (>= 4.0), ballgown

Imports IRanges, GenomicRanges, S4Vectors, parallel, matrixStats, GenomicAlignments, GenomeInfoDb, Rsamtools, cluster, stats, graphics, methods

Suggests knitr, rmarkdown

VignetteBuilder knitr

biocViews RNASeq, AlternativeSplicing, DifferentialSplicing, StatisticalMethod, Visualization, Preprocessing, Clustering, DifferentialExpression, KEGG, ImmunoOncology

Encoding UTF-8

LazyData false

RoxygenNote 7.1.1

Config/pak/sysreqs
make libbz2-dev liblzma-dev libpng-dev libxml2-dev libssl-dev xz-utils zlib1g-dev

Repository Bioconductor 3.24

Date/Publication 2026-11-23 22:44:04 UTC

RemoteUrl https://github.com/bioc/VaSP

RemoteRef RELEASE_3_24

RemoteSha 65ac90860d6fb8a8f1fb9825a44b4cf279f249f0

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`BMfinder` *Discover bimodal distrubition features*

Description

Find bimodal distrubition features and divide the samples into 2 groups by k-means clustering.

Usage

```
BMfinder(x, p.value = 0.01, maf = 0.05, miss = 0.05, fold = 2, log = FALSE,
        cores = detectCores() - 1)
```

Arguments

`x`	a numeric matrix with feature rows and sample columns, e.g., splicing score matrix from `spliceGenome` or `spliceGene` function.
`p.value`	p.value threshold for bimodal distrubition test
`maf`	minor allele frequency threshold in k-means clustering
`miss`	missing grouping rate threshold in k-means clustering
`fold`	fold change threshold between the two groups
`log`	whether the scores are to be logarithmic. If TRUE, all the scores are log2 tranformed before k-means clustering: x = log2(x+1).
`cores`	threads to be used. This value is passed to **?mclapply** in **parallel** package

Details

The matrix contains 1, 2 and NA, and values of 'x' in group 2 are larger than group 1.

Value

a matrix with feature rows and sample columns.

Examples

```
data(rice.bg)
score<-spliceGene(rice.bg,'MSTRG.183',junc.type='score')
score<-round(score,2)
as<-BMfinder(score,cores=1) # 4 bimodal distrubition features found

##compare
as
score[rownames(score)%in%rownames(as),]
```

---

getDepth *Get Read Depth*

Description

Get read depth from a BAM file (in bedgraph format)

Usage

```
getDepth(x, chrom, start, end)
```

Arguments

x	path to a BAM file
chrom	chromosome of a region to be searched
start	start position
end	end position

Value

a data.frame in bedgraph file format.

Examples

```
path <- system.file('extdata',package='VaSP')
bam_files<-list.files(path,'bam$')
bam_files

depth<-getDepth(file.path(path, bam_files[1]), 'Chr1',
                start=1171800, end=1179400)
head(depth)

# library(Sushi)
# plotBedgraph(depth,'Chr1',chromstart=1171800, chromend=1179400,yaxt='s')
# mtext('Depth',side=2,line=2.5,cex=1.2,font=2)
# labelgenome('Chr1',1171800,1179400,side=1,scipen=20,n=5,scale='Kb')
```

getGeneinfo *Get Gene Informaton from a ballgown object*

Description

Get gene informaton from a ballgown object by genes or by genomic regions

Usage

```
getGeneinfo(genes = NA, bg, chrom, start, end, samples = sampleNames(bg),
            trans.select = NA)
```

Arguments

genes	a character vector specifying gene IDs in 'bg'. Any values other than NA override genomic region (chrom, start, stop)
bg	ballgown object
chrom	chromosome of a region
start	start postion
end	stop postion
samples	names of samples. The transcrpts in these samples are subjected to 'trans.select'
trans.select	logical expression-like string, indicating transcript rows to select from a matrix of transcript coverages: NA value keeps all transcripts.

Value

a data.frame in bed-like file format

Examples

```
data(rice.bg)
unique(geneIDs(rice.bg))

gene_id <- c('MSTRG.181', 'MSTRG.182', 'MSTRG.183')
geneinfo <- getGeneinfo(genes=gene_id,rice.bg)
trans <- table(geneinfo$name) # show how many exons each transcript has
trans

# library(Sushi)
# chrom = geneinfo$chrom[1]
# chromstart = min(geneinfo$start) - 1e3
# chromend = max(geneinfo$stop) + 1e3
# color = rep(SushiColors(2)(length(trans)), trans)

# par(mar=c(3,1,1,1))
# plotGenes(geneinfo, chrom, chromstart, chromend, col = color, bheight = 0.2,
#           bentline = FALSE, plotgenetype = 'arrow', labeloffset = 0.5)
# labelgenome(chrom, chromstart , chromend, side = 1, n = 5, scale = 'Kb')
```

`rice.bg` *Rice ballgown object*

Description

Small ballgown object created with a subset of rice RNAseq data, for demonstration purposes

Format

a ballgown object with 33 transcripts and 6 samples

Details

The raw RNA-seq data were screened and trimmed using Trimmomatic (Bolger et al., 2014) and RNA-seq mapping, transcript assembly, and quantification were conducted with HISAT, StringTie, and Ballgown by following the method described by Pertea et al. (Pertea et al., 2016). The rice.bg is a subset ballgown object with 33 transcripts and 6 samples (Yu et al., 2021).

Source

The raw RNA-seq data were from the project of variation in transcriptional responses to salt stress in rice (SRA Accession: SRP106054)

References

Yu, H., Du, Q., Campbell, M., Yu, B., Walia, H. and Zhang, C. (2021), Genome-wide discovery of natural variation in pre-mRNA splicing and prioritising causal alternative splicing to salt stress response in rice. New Phytol. https://doi.org/10.1111/nph.17189

Bolger, A.M., Lohse, M., and Usadel, B. (2014). Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30, 2114-2120.

Pertea, M., Kim, D., Pertea, G.M., Leek, J.T., and Salzberg, S.L. (2016). Transcript-level expression analysis of RNA-seq experiments with HISAT, StringTie and Ballgown. Nat Protoc 11, 1650-1667.

Examples

```
data(rice.bg)
rice.bg
# ballgown instance with 33 transcripts and 6 samples
```

spliceGene *Calculate Splicing Scores for One Gene*

Description

Calculate splicing Scores from ballgown object for a given gene. This function can only calculate one gene. Please use function spliceGenome to obtain genome-wide splicing scores.

Usage

```
spliceGene(bg, gene, samples = sampleNames(bg), junc.type = c("score", "count"),
           trans.select = "rowMaxs(x)>=1", junc.select = "rowMaxs(x)>=5")
```

Arguments

bg	ballgown object
gene	a character string specifying gene id
samples	names of samples
junc.type	type of junction estimate ('score' for junction score; 'count' for junction read count)
trans.select	logical expression-like string, indicating transcript rows to select from a matrix of transcript coverages: NA value keeps all transcripts. e.g. use trans.select='rowMaxs(x)>=1' to filter the transcrpts with the maximium coverage among all the samples less than 1.
junc.select	logical expression-like string, indicating junction rows to select from a matrix of junction counts: NA value keeps all junctions. e.g. use junc.select='rowMaxs(x)>=5' to filter the junctions with the maximium read count among all the samples less than 5.

Details

score = junction count/gene-level per base read coverage. Row functions for matrices are useful to select transcripts and junctions. See matrixStats package.

Value

a matrix of junction scores with intron rows and sample columns.

References

Yu, H., Du, Q., Campbell, M., Yu, B., Walia, H. and Zhang, C. (2021), Genome-wide discovery of natural variation in pre-mRNA splicing and prioritising causal alternative splicing to salt stress response in rice. New Phytol. https://doi.org/10.1111/nph.17189

See Also

spliceGenome, which calculates splicing scores in whole genome.

Examples

```
data(rice.bg)
rice.bg
head(geneIDs(rice.bg))

score<-spliceGene(rice.bg,'MSTRG.183',junc.type='score')
count<-spliceGene(rice.bg,'MSTRG.183',junc.type='count')

## compare
tail(score)
tail(count)

## get intron structrue
intron<-structure(rice.bg)$intron
intron[intron$id%in%rownames(score)]
```

`spliceGenome` *Calculate Genome-wide Splicing Scores*

Description

Calculate splicing scores from ballgown objects for all genes.

Usage

```
spliceGenome(bg, gene.select = "rowQuantiles(x,probs = 0.05)>=1",
            intron.select = "rowQuantiles(x,probs = 0.95)>=5")
```

Arguments

`bg`	ballgown object
`gene.select`	logical expression-like string, indicating genes to select from a matrix of gene-level coverages: NA value keeps all genes. e.g. gene.select = 'rowQuantiles(x,probs = 0.05)>=1' keeps the genes with the read coverage greater than or equal to 1 in at least 95 (0.05 quantile). Used to filter low expressed genes.
`intron.select`	logical expression-like string, indicating introns to select from a matrix of junction counts: NA value keeps all introns. e.g. intron.select = 'rowQuantiles(x,probs = 0.95)>=5' keeps the introns with the read count greater than or euqal to 5 in at least 5 (0.95 quantile). Used to filter introns with very few junction reads supporting.

Details

score = junction count/gene-level per base read coverage. Row functions for matrices in `matrixStats` package are useful to select genes and introns.

Value

a list of two elelments: 'score' is matrix of intron splicing scores with intron rows and sample columns and 'intron' is a GRanges object of intron structure. See structure in **ballgown** package

References

Yu, H., Du, Q., Campbell, M., Yu, B., Walia, H. and Zhang, C. (2021), Genome-wide discovery of natural variation in pre-mRNA splicing and prioritising causal alternative splicing to salt stress response in rice. New Phytol. https://doi.org/10.1111/nph.17189

See Also

spliceGene, which calculates splicing scores in one gene.

Examples

```
data(rice.bg)
rice.bg

splice<-spliceGenome(rice.bg,gene.select=NA,intron.select=NA)
names(splice)

head(splice$score)
splice$intron
```

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