

# Package: crisprBase (via r-universe)

May 29, 2026

**Version** 1.16.0

**Date** 2025-10-27

**Title** Base functions and classes for CRISPR gRNA design

**Depends** utils, methods, R (>= 4.1)

**Imports** BiocGenerics, Biostrings, GenomicRanges, graphics, IRanges, S4Vectors, stringr

**Suggests** BiocStyle, knitr, rmarkdown, testthat

**biocViews** CRISPR, FunctionalGenomics

**Description** Provides S4 classes for general nucleases, CRISPR nucleases, CRISPR nickases, and base editors. Several CRISPR-specific genome arithmetic functions are implemented to help extract genomic coordinates of spacer and protospacer sequences. Commonly-used CRISPR nuclease objects are provided that can be readily used in other packages. Both DNA- and RNA-targeting nucleases are supported.

**License** MIT + file LICENSE

**Encoding** UTF-8

**RoxygenNote** 7.3.3

**VignetteBuilder** knitr

**BugReports** <https://github.com/crisprVerse/crisprBase/issues>

**URL** <https://github.com/crisprVerse/crisprBase>

**Collate** 'AllGenerics.R' 'rebase.R' 'Nuclease-class.R'  
'Nickase-class.R' 'CrisprNuclease-class.R'  
'CrisprNickase-class.R' 'BaseEditor-class.R' 'arithmetics.R'  
'data.R' 'utils.R' 'converters.R' 'annotateMismatches.R'

**LazyData** true

**Config/pak/sysreqs** libicu-dev zlib1g-dev

**Repository** <https://bioc-release.r-universe.dev>

**Date/Publication** 2026-04-28 12:57:37 UTC

**RemoteUrl** <https://github.com/bioc/crisprBase>

**RemoteRef** RELEASE\_3\_23

**RemoteSha** 00f5ba957ff3ff7b5b69591c332dc36395d9c44f

## Contents

annotateMismatches . . . . .	2
AsCas12a . . . . .	3
baseEditorName . . . . .	4
BE4max . . . . .	6
CasRx . . . . .	7
CrisprNickase-class . . . . .	7
Csm . . . . .	10
enAsCas12a . . . . .	11
extractPamFromTarget . . . . .	11
extractProtospacerFromTarget . . . . .	12
getAvailableCrisprNucleases . . . . .	13
getCutSiteFromPamSite . . . . .	14
getCutSiteRanges . . . . .	14
getEditingSiteFromPamSite . . . . .	15
getPamRanges . . . . .	16
getProtospacerRanges . . . . .	17
getTargetRanges . . . . .	18
MAD7 . . . . .	19
motifs . . . . .	20
nickaseName . . . . .	23
plotEditingWeights . . . . .	27
restrictionEnzymes . . . . .	28
SaCas9 . . . . .	28
spacerLength . . . . .	29
SpCas9 . . . . .	32
SpG . . . . .	33
<b>Index</b>	<b>34</b>

---

annotateMismatches	<i>Annotate mismatches between spacer and protospacer sequences</i>
--------------------	---

---

### Description

Annotate mismatches between spacer and protospacer sequences.

### Usage

```
annotateMismatches(spacers, protospacers, rnase = FALSE)
```

**Arguments**

spacers	A character vector specifying spacer sequences (gRNA).
protospacers	A character vector specifying protospacer sequences (target DNA).
rnase	Is it for an RNase? FALSE by default. If TRUE, spacers and protospacers are expected to be the reverse complement of each other.

**Value**

A data.frame storing spacer and protospacer columns, as well as number of mismatches, and positions for the different mismatches, if any. Positions are relative to the 5' end of the spacer sequences. For RNases (e.g. CasRx), this means that a mismatch at position 1 corresponds to the last nucleotide of the protospacer sequence.

**Author(s)**

Jean-Philippe Fortin

**Examples**

```
spacers <- c("CCGGAGCGAGTTGCAGTAAGCAG",
            "GCCGGAGCGAGTTGCAGTAAGCA",
            "GGCCGGAGCGAGTTGCAGTAAGC")

protospacers=c("CTGCTTACTGCAACTCGCTCTGG",
              "TGCTTAATGCAACCCGCTCCGGC",
              "GCTTACTGCAACTCGCTCCGGC")

ann <- annotateMismatches(spacers,
                        protospacers,
                        rnase=TRUE)
```

---

AsCas12a

*AsCas12a CrisprNuclease object*

---

**Description**

CrisprNuclease object for the Wildtype Acidaminococcus Cas12a (AsCas12a) nuclease.

**Usage**

```
data(AsCas12a, package="crisprBase")
```

**Format**

CrisprNuclease object.

**Details**

The AsCas12a nuclease recognizes TTTV PAM sequences. Spacer sequences must be located downstream of PAM sequences.

---

baseEditorName	<i>An S4 class to represent a base editor</i>
----------------	---

---

**Description**

An S4 class to represent a base editor

**Usage**

```
baseEditorName(object)

baseEditorName(object) <- value

editingWeights(object, ...)

editingStrand(object, ...)

editingStrand(object) <- value

BaseEditor(
  CrisprNuclease,
  baseEditorName = NA_character_,
  editingStrand = c("original", "opposite"),
  editingWeights = NULL,
  scale = TRUE
)

## S4 method for signature 'BaseEditor'
show(object)

## S4 method for signature 'BaseEditor'
baseEditorName(object)

## S4 replacement method for signature 'BaseEditor'
baseEditorName(object) <- value

## S4 method for signature 'BaseEditor'
editingWeights(object, substitutions = NULL)

setEditingWeights(object, value, scale = TRUE)

## S4 method for signature 'BaseEditor'
```

```

editingStrand(object)

## S4 replacement method for signature 'BaseEditor'
editingStrand(object) <- value

```

### Arguments

object	BaseEditor object.
value	Value to replaced with.
...	Additional arguments for class-specific methods
CrisprNuclease	A CrisprNuclease object.
baseEditorName	String specifying base editor name.
editingStrand	String indicating which strand with respect to the target protospacer sequence will be edited. Must be either "original" or "opposite". "original" by default.
editingWeights	Numeric matrix of editing weights. Column names must be indicating relative position to the PAM site. Row names must be of the form "X2Y" where "X" represents the origin base, and "Y" represents the substituted base. For instance, "C2T" indicates the row corresponding to C to T editing.
scale	Logical indicating if weights should be scaled to have a maximum of 1. TRUE by default. Should be set to FALSE if editing weights are already representing editing probabilities.
substitutions	Character vector indicating which substitutions should be returned.

### Value

A BaseEditor object

### Functions

- BaseEditor(): Create a BaseEditor object

### Slots

baseEditorName	Name of the base editor.
editingWeights	Matrix of editing weights.
editingStrand	String indicating which strand with respect to the target protospacer sequence will be edited. Must be either "original" or "opposite". "original" by default.

### Constructors

Use the constructor `link{BaseEditor}` to create a BaseEditor object.

### Accessors

baseEditorName:	To get the name of the base editor.
editingWeights:	To return the matrix of editing weights.
editingStrand:	To return the editing strand.

**Setters**

baseEditorName<-: To change the name of the base editor.  
 editingWeights<-: To change the matrix of editing weights.  
 editingStrand<-: To change the editing strand.

**Examples**

```
# Creating an object for BE4max (C to T editor)
# based on experimental weights

ws <- c(0.7, 0.7, 0.8, 1.8, 1, 2, 1.4, 1.2, 2.3, 1.3, 2.4, 2.2, 3.4,
        2.2, 2.1, 3.5, 5.8, 16.2, 31.8, 63.2, 90.3, 100, 87, 62, 31.4,
        16.3, 10, 5.6, 3.3, 1.9, 1.8, 2.4, 1.7, 0.5, 0.2, 0.1)
ws <- matrix(ws, nrow=1, ncol=length(ws))
rownames(ws) <- "C2T"
colnames(ws) <- -36:-1
data(SpCas9, package="crisprBase")
BE4max <- BaseEditor(SpCas9,
                    baseEditorName="BE4max",
                    editingStrand="original",
                    editingWeights=ws)
metadata(BE4max)$description_base_editor <- "BE4max cytosine base editor."
```

---

 BE4max

*BE4max BaseEditor object*


---

**Description**

BaseEditor for the cytosine base editor CRISPR/Cas9 system BE4max. Editing weights were obtained from <https://doi.org/10.1016/j.cell.2020.05.037>

**Usage**

```
data(BE4max, package="crisprBase")
```

**Format**

BaseEditor object.

**Details**

BaseEditor for the cytosine base editor CRISPR/Cas9 system BE4max. Editing weights were obtained from <https://doi.org/10.1016/j.cell.2020.05.037>.

---

CasRx	<i>CasRx CrisprNuclease object</i>
-------	------------------------------------

---

**Description**

CrisprNuclease object for the Cas13d-NLS from *Ruminococcus flavefaciens* strain XPD3002 nuclease (RNase).

**Usage**

```
data(CasRx, package="crisprBase")
```

**Format**

CrisprNuclease object.

**Details**

The CasRx nuclease was derived from Cas13d *Ruminococcus flavefaciens* strain XPD3002. See [10.1016/j.cell.2018.02.033](https://doi.org/10.1016/j.cell.2018.02.033).

---

CrisprNickase-class	<i>An S4 class to represent a CRISPR nickase.</i>
---------------------	---

---

**Description**

An S4 class to represent a CRISPR nickase.

**Usage**

```
CrisprNickase(
  nickaseName,
  nickingStrand = c("original", "opposite"),
  pams = NA_character_,
  weights = rep(1, length(pams)),
  metadata = list(),
  pam_side = NA_character_,
  spacer_gap = 0L,
  spacer_length = NA_integer_
)

## S4 method for signature 'CrisprNickase'
show(object)

## S4 method for signature 'CrisprNickase'
pamLength(object)
```

```

## S4 method for signature 'CrisprNickase'
spacerLength(object)

## S4 replacement method for signature 'CrisprNickase'
spacerLength(object) <- value

## S4 method for signature 'CrisprNickase'
pamSide(object)

## S4 replacement method for signature 'CrisprNickase'
pamSide(object) <- value

## S4 method for signature 'CrisprNickase'
spacerGap(object)

## S4 replacement method for signature 'CrisprNickase'
spacerGap(object) <- value

## S4 method for signature 'CrisprNickase'
hasSpacerGap(object)

## S4 method for signature 'CrisprNickase'
targetLength(object)

## S4 method for signature 'CrisprNickase'
pams(object, primary = TRUE, ignore_pam = FALSE, as.character = FALSE)

## S4 method for signature 'CrisprNickase'
pamIndices(object)

## S4 method for signature 'CrisprNickase'
spacerIndices(object)

## S4 method for signature 'CrisprNickase'
prototypeSequence(object, primary = TRUE)

```

### Arguments

<code>nickaseName</code>	Name of the CRISPR nickase.
<code>nickingStrand</code>	String specifying with strand with respect to the motif sequence (5' to 3') is nicked. Must be either "original" (default) or "opposite".
<code>pams</code>	Character vector of PAM sequence motifs written from 5' to 3'. If the point of cleavage has been determined, the precise site is marked with ^. Only letters in the IUPAC code are accepted. For nickases that cleave away from their recognition sequence, the cleavage sites are indicated in parentheses. See details for more information.

weights	Optional numeric vector specifying relative weights of the PAM sequences to specify cleavage probabilities.
metadata	Optional list providing global metadata information.
pam_side	String specifying the side of the PAM sequence with respect to the protospacer sequence. Must be either '3prime' (e.g. Cas9) or '5prime' (e.g. Cas12a)
spacer_gap	Integer specifying the length (in nucleotides) between the spacer sequence and the PAM sequence (e.g. 0 for Cas9 and Cas12a).
spacer_length	Integer specifying the length of the spacer sequence
object	<a href="#">CrisprNickase</a> object.
value	For spacerLength<- and gapLength<-, must be a non-negative integer. For pamSide, must be either '5prime' or '3prime'.
primary	Should only the PAM sequence with the heighest weight be returned? If no cleavage weights are stored in the <a href="#">CrisprNickase</a> object, all sequences are returned. TRUE by default.
ignore_pam	Should all possible k-mer sequences for a given PAM length be returned, ir-respectively of the PAM sequence motifs stored in the <a href="#">CrisprNickase</a> object? FALSE by default.
as.character	Should the PAM sequences be returned as a character vector? FALSE by default.

**Value**

A [CrisprNickase](#) object

**Functions**

- [CrisprNickase\(\)](#): Create a [CrisprNickase](#) object

**Slots**

`pam_side` String specifying the side of the PAM sequence with respect to the protospacer sequence. Must be either '3prime' (e.g. SpCas9) or '5prime' (e.g. AsCas12a)

`spacer_length` Integer specifying the length of the spacer sequence

`spacer_gap` Integer specifying the length (in nucleotides) between the spacer sequence and the PAM sequence (e.g. 0 for SpCas9 and AsCas12a).

**Constructors**

Use the constructor `link{CrisprNickase}` to create a [CrisprNickase](#) object.

**Accessors**

`nickaseName`: To get the name of the CRISPR nickase.

`spacerLength`: To return the length of the spacer sequence.

`targetLength`: To return the length of the target sequence (protospacer + pam).

**pamLength:** To return the length of the PAM sequence.

**pamSide:** To return the side of the PAM sequence with respect to the spacer sequence.

**spacerGap:** To return the length of the gap between the PAM and spacer sequences.

**pams:** To return the list of PAM sequences.

### Setters

**spacerGap<-:** To change the length of the gap between the PAM and spacer sequences.

**pamSide<-:** To change the side of the PAM sequence with respect to the protospacer sequence.

**spacerLength<-:** To change the length of the spacer sequence.

### Utility functions for genomic arithmetics

**pamIndices:** To return the relative coordinates of the PAM sequence within the protospacer sequence.

**spacerIndices:** To return the relative coordinates of the spacer sequence within the protospacer sequence.

### Examples

```
Cas9D10A <- CrisprNickase("Cas9D10A",
  nickingStrand="opposite",
  pams=c("(3)NGG", "(3)NAG", "(3)NGA"),
  weights=c(1, 0.2593, 0.0694),
  metadata=list(description="D10A-mutated Streptococcus
    pyogenes Cas9 (SpCas9) nickase"),
  pam_side="3prime",
  spacer_length=20)

Cas9H840A <- CrisprNickase("Cas9H840A",
  nickingStrand="original",
  pams=c("(3)NGG", "(3)NAG", "(3)NGA"),
  weights=c(1, 0.2593, 0.0694),
  metadata=list(description="H840A-mutated Streptococcus
    pyogenes Cas9 (SpCas9) nickase"),
  pam_side="3prime",
  spacer_length=20)
```

### Description

CrisprNuclease object for the RNA-targeting Csm complex from *Streptococcus thermophilus*

**Usage**

```
data(Csm, package="crisprBase")
```

**Format**

CrisprNuclease object.

**Details**

The specific Csm complex is an RNA-targeting nuclease derived from *Streptococcus thermophilus*. There is no preferred PAM sequences, and the default (optimal) spacer length is 32nt. See <https://doi.org/10.1038/s41587-022-01649-9>.

---

enAsCas12a

*enAsCas12a CrisprNuclease object*

---

**Description**

CrisprNuclease object for the Enhanced *Acidaminococcus* Cas12a (AsCas12a) nuclease.

**Usage**

```
data(enAsCas12a, package="crisprBase")
```

**Format**

CrisprNuclease object.

**Details**

The enAsCas12a nuclease recognizes an extended set of PAM sequences beyond the canonical TTTV sequence for AsCas12a. Spacer sequences must be located downstream of PAM sequences.

---

extractPamFromTarget

*Extract PAM sequences from target sequences*

---

**Description**

Extract PAM sequences from target sequences (protospacer + PAM) using information stored in a CrisprNuclease object.

**Usage**

```
extractPamFromTarget(targets, object)
```

**Arguments**

targets            Character vector of target sequences.  
object            CrisprNuclease corresponding to the target sequences.

**Value**

Character vector of PAM sequences of length equal to that of the targets character vector.

**Author(s)**

Jean-Philippe Fortin

**Examples**

```
data(SpCas9, AsCas12a, package="crisprBase")
# Extracting PAM sequences from Cas9 protospacers:
targets <- c("AGGTGCTGATTGTAGTGCTGCGG",
            "AGGTGCTGATTGTAGTGCTGAGG")
extractPamFromTarget(targets, SpCas9)
# Extracting PAM sequences from Cas12a targets:
targets <- c("TTTAAGGTGCTGATTGTAGTGCTGTGT",
            "TTTCAGGTGCTGATTGTAGTGCTGAAA")
extractPamFromTarget(targets, AsCas12a)
```

---

extractProtospacerFromTarget

*Extract protospacer sequences from target sequences*

---

**Description**

Extract protospacer sequences from target sequences (protospacer + PAM) using information stored in a CrisprNuclease object.

**Usage**

```
extractProtospacerFromTarget(targets, object)
```

**Arguments**

targets            Character vector of targets sequences.  
object            CrisprNuclease corresponding to the targets sequences.

**Value**

Character vector of protospacer sequences of length equal to that of the targets character vector.

**Author(s)**

Jean-Philippe Fortin

**Examples**

```
data(SpCas9, AsCas12a, package="crisprBase")
# Extracting protospacer sequences from Cas9 targets:
targets <- c("AGGTGCTGATTGTAGTGCTCGG",
            "AGGTGCTGATTGTAGTGCTGAGG")
extractProtospacerFromTarget(targets, SpCas9)
# Extracting protospacer sequences from Cas12a targets:
targets <- c("TTTAAGGTGCTGATTGTAGTGCTGTGT",
            "TTTCAGGTGCTGATTGTAGTGCTGAAA")
extractProtospacerFromTarget(targets, AsCas12a)
```

---

`getAvailableCrisprNucleases`

*Return list of available CrisprNuclease objects in crisprBase*

---

**Description**

Return list of available CrisprNuclease objects in crisprBase.

**Usage**

```
getAvailableCrisprNucleases()
```

**Value**

Character vector of available CrisprNuclease objects found in crisprBase.

**Author(s)**

Jean-Philippe Fortin

**Examples**

```
getAvailableCrisprNucleases()
```

---

getCutSiteFromPamSite *Return cut site coordinates from PAM site coordinates*

---

### Description

Return cut site coordinates from PAM site coordinates.

### Usage

```
getCutSiteFromPamSite(pam_site, strand, nuclease = NULL)
```

### Arguments

pam_site	Coordinate of the first nucleotide of the PAM sequence.
strand	Either "+" or "-".
nuclease	A <a href="#">CrisprNuclease</a> object.

### Value

numeric vector of cut sites

### Author(s)

Jean-Philippe Fortin

### Examples

```
data(SpCas9, package="crisprBase")
getCutSiteFromPamSite(pam_site=100, strand="+", nuclease=SpCas9)
getCutSiteFromPamSite(pam_site=100, strand="-", nuclease=SpCas9)
```

---

getCutSiteRanges *Construct a cut site GRanges from a list of PAM sites*

---

### Description

Construct a cut site GRanges from a list of PAM sites using information stored in a CrisprNuclease object.

**Usage**

```
getCutSiteRanges(
  gr = NULL,
  seqnames = NULL,
  pam_site = NULL,
  strand = NULL,
  nuclease = NULL
)
```

**Arguments**

<code>gr</code>	GRanges object of width 1 specifying the coordinates of the first nucleotide of the PAM sequences.
<code>seqnames</code>	Character vector of genomic sequence names. Ignored if <code>gr</code> is not NULL.
<code>pam_site</code>	Numeric vector specifying the coordinates of the first nucleotide of the PAM sequences corresponding to the PAM sequences. Ignored if <code>gr</code> is not NULL.
<code>strand</code>	Character vector specifying the strand of the PAM. Ignored if <code>gr</code> is not NULL.
<code>nuclease</code>	CrisprNuclease object.

**Value**

GRanges object representing genomic coordinates of the cut sites.

**Author(s)**

Jean-Philippe Fortin

**Examples**

```
data(SpCas9, AsCas12a, package="crisprBase")
library(GenomicRanges)
gr <- GRanges("chr10",
              IRanges(start=c(100,120), width=1),
              strand=c("+","-"))
getCutSiteRanges(gr, nuclease=SpCas9)
getCutSiteRanges(gr, nuclease=AsCas12a)
```

---

getEditingSiteFromPamSite

*Return optimal editing site coordinates from PAM site coordinates*

---

**Description**

Return optimal editing site coordinates from PAM site coordinates.

**Usage**

```

getEditingSiteFromPamSite(
  pam_site,
  strand,
  baseEditor = NULL,
  substitution = NULL
)

```

**Arguments**

pam_site	Coordinate of the first nucleotide of the PAM sequence.
strand	Either "+" or "-".
baseEditor	A <a href="#">BaseEditor</a> object.
substitution	String indicating which substitution should be used to estimate the optimal editing position. E.g. "C2T" will return the optimal editing position for C to T editing.

**Value**

numeric vector of editing sites.

**Author(s)**

Jean-Philippe Fortin

**Examples**

```

data(BE4max, package="crisprBase")
getEditingSiteFromPamSite(pam_site=100, strand="+", baseEditor=BE4max, "C2T")

```

---

getPamRanges

*Construct a PAM GRanges from a list of PAM sites*

---

**Description**

Construct a PAM GRanges from a list of PAM sites using information stored in a CrisprNuclease object.

**Usage**

```

getPamRanges(
  gr = NULL,
  seqnames = NULL,
  pam_site = NULL,
  strand = NULL,
  nuclease = NULL
)

```

**Arguments**

<code>gr</code>	GRanges object of width 1 specifying the coordinates of the first nucleotide of the PAM sequences.
<code>seqnames</code>	Character vector of genomic sequence names. Ignored if <code>gr</code> is not NULL.
<code>pam_site</code>	Numeric vector specifying the coordinates of the first nucleotide of the PAM sequences corresponding to the PAM sequences. Ignored if <code>gr</code> is not NULL.
<code>strand</code>	Character vector specifying the strand of the PAM. Ignored if <code>gr</code> is not NULL.
<code>nuclease</code>	CrisprNuclease object.

**Value**

GRanges object representing genomic coordinates of PAM sequences.

**Author(s)**

Jean-Philippe Fortin

**Examples**

```
data(SpCas9, AsCas12a, package="crisprBase")
library(GenomicRanges)
gr <- GRanges("chr10",
              IRanges(start=c(100,120), width=1),
              strand=c("+","-"))
getPamRanges(gr, nuclease=SpCas9)
getPamRanges(gr, nuclease=AsCas12a)
```

---

`getProtospacerRanges` *Construct a protospacer GRanges from a list of PAM sites*

---

**Description**

Construct a protospacer GRanges from a list of PAM sites using information stored in a CrisprNuclease object.

**Usage**

```
getProtospacerRanges(
  gr = NULL,
  seqnames = NULL,
  pam_site = NULL,
  strand = NULL,
  nuclease = NULL,
  spacer_len = NULL
)
```

**Arguments**

gr	GRanges object of width 1 specifying the coordinates of the first nucleotide of the PAM sequences.
seqnames	Character vector of genomic sequence names. Ignored if gr is not NULL.
pam_site	Numeric vector specifying the coordinates of the first nucleotide of the PAM sequences corresponding to the protospacers. Ignored if gr is not NULL.
strand	Character vector specifying the strand of the protospacer. Ignored if gr is not NULL.
nuclease	CrisprNuclease object.
spacer_len	Non-negative integer to overwrite the default spacer length stored in the CrisprNuclease object. s

**Value**

GRanges object representing genomic coordinates of protospacer sequences.

**Author(s)**

Jean-Philippe Fortin

**Examples**

```
data(SpCas9, AsCas12a, package="crisprBase")
library(GenomicRanges)
gr <- GRanges("chr10",
              IRanges(start=c(100,120), width=1),
              strand=c("+", "-"))
getProtospacerRanges(gr, nuclease=SpCas9)
getProtospacerRanges(gr, nuclease=AsCas12a)
```

---

getTargetRanges

*Construct a target GRanges from a list of PAM sites*

---

**Description**

Construct a target (protospacer + PAM) GRanges from a list of PAM sites using information stored in a CrisprNuclease object.

**Usage**

```
getTargetRanges(
  gr = NULL,
  seqnames = NULL,
  pam_site = NULL,
  strand = NULL,
```

```

nuclease = NULL,
spacer_len = NULL
)

```

### Arguments

<code>gr</code>	GRanges object of width 1 specifying the coordinates of the first nucleotide of the PAM sequences.
<code>seqnames</code>	Character vector of genomic sequence names. Ignored if <code>gr</code> is not NULL.
<code>pam_site</code>	Numeric vector specifying the coordinates of the first nucleotide of the PAM sequences corresponding to the targets. Ignored if <code>gr</code> is not NULL.
<code>strand</code>	Character vector specifying the strand of the target. Ignored if <code>gr</code> is not NULL.
<code>nuclease</code>	CrisprNuclease object.
<code>spacer_len</code>	Non-negative integer to overwrite the default spacer length stored in the CrisprNuclease object.

### Value

GRanges object representing genomic coordinates of the target sequences.

### Author(s)

Jean-Philippe Fortin

### Examples

```

data(SpCas9, AsCas12a, package="crisprBase")
library(GenomicRanges)
gr <- GRanges("chr10",
              IRanges(start=c(100,120), width=1),
              strand=c("+", "-"))
getTargetRanges(gr, nuclease=SpCas9)
getTargetRanges(gr, nuclease=AsCas12a)

```

---

MAD7

*MAD7 CrisprNuclease object*


---

### Description

CrisprNuclease object for the MAD7 nuclease (Cas12a-like nuclease)

### Usage

```
data(MAD7, package="crisprBase")
```

**Format**

CrisprNuclease object.

**Details**

The MAD7 nuclease recognizes YTTV PAM sequences. Spacer sequences must be located downstream of PAM sequences.

---

motifs

*An S4 class to represent a nuclease.*

---

**Description**

Return motif string representations of recognition sites.

Return length of the recognition sites sequences.

**Usage**

```
motifs(object, ...)
```

```
motifLength(object, ...)
```

```
nucleaseName(object)
```

```
targetType(object)
```

```
weights(object, ...)
```

```
nucleaseName(object) <- value
```

```
targetType(object) <- value
```

```
weights(object) <- value
```

```
cutSites(object, ...)
```

```
isCutting(object)
```

```
isRnase(object)
```

```
isDnase(object)
```

```
Nuclease(  
  nucleaseName,  
  targetType = c("DNA", "RNA"),  
  motifs = NULL,
```

```
    cutSites = NULL,
    weights = rep(1, length(motifs)),
    metadata = list()
)

## S4 method for signature 'Nuclease'
show(object)

## S4 method for signature 'Nuclease'
nucleaseName(object)

## S4 replacement method for signature 'Nuclease'
nucleaseName(object) <- value

## S4 method for signature 'Nuclease'
targetType(object)

## S4 replacement method for signature 'Nuclease'
targetType(object) <- value

## S4 method for signature 'Nuclease'
weights(object, expand = FALSE)

## S4 replacement method for signature 'Nuclease'
weights(object) <- value

## S4 method for signature 'Nuclease'
isCutting(object)

## S4 method for signature 'Nuclease'
isRnase(object)

## S4 method for signature 'Nuclease'
isDnase(object)

## S4 method for signature 'Nuclease'
motifs(
  object,
  primary = FALSE,
  strand = c("+", "-"),
  expand = FALSE,
  as.character = FALSE
)

## S4 method for signature 'Nuclease'
motifLength(object)

## S4 method for signature 'Nuclease'
```

```
cutSites(object, strand = c("+", "-", "both"), combine = TRUE, middle = FALSE)
```

### Arguments

object	<a href="#">Nuclease</a> object.
...	Additional arguments for class-specific methods
value	New value to pass to the setter functions.
nucleaseName	Name of the nuclease.
targetType	String specifying target type ("DNA" or "RNA").
motifs	Character vector of recognition sequence motifs written from 5' to 3' written in Rebase convention. If the point of cleavage has been determined, the precise site is marked with ^. Only letters in the IUPAC code are accepted. For nucleases that cleave away from their recognition sequence, the cleavage sites are indicated in parentheses. See details for more information.
cutSites	Matrix with 2 rows (+ and - strand, respectively) specifying the cleavage coordinates relative to the first nucleotide of the motif sequence. Each column corresponds to a motif specified in the motifs slot.
weights	Optional numeric vector specifying relative weights for the recognition motifs to specify cleavage probabilities.
metadata	Optional list providing global metadata information.
expand	Should sequences be expanded to only contain ATCG nucleotides? FALSE by default.
primary	Should only the motif with the highest weight be returned? FALSE by default. Only relevant if weights are stored in the <a href="#">Nuclease</a> object.
strand	Strand to allow reverse complementation of the motif. "+" by default.
as.character	Should the motif sequences be returned as a character vector? FALSE by default.
combine	Should only unique values be considered? TRUE by default.
middle	For staggered cuts, should the middle point between the cut on the forward strand and the cut on the reverse strand be considered as the cut site? FALSE by default.

### Value

A [Nuclease](#) object

### Functions

- [Nuclease\(\)](#): Create a [Nuclease](#) object

### Slots

nucleaseName Name of the nuclease.  
targetType Character string indicating target type ("DNA" or "RNA").

**motifs** DNASTringSet of recognition sequence motifs written from 5' to 3'.

**cutSites** Matrix with 2 rows (+ and - strand, respectively) specifying the cleavage coordinates relative to the first nucleotide of the motif sequence. Each column corresponds to a motif specified in the motifs slot.

**weights** Optional numeric vector specifying relative weights for the motifs corresponding to cleavage probabilities.

**metadata** Optional string providing a description of the nuclease.

### Constructors

Use the constructor `link{Nuclease}` to create a Nuclease object.

### Accessors

**nucleaseName:** To get the name of the nuclease.

**targetType:** To get the target type ("DNA" or "RNA").

**metadata:** To get the metadata list of the nuclease.

**motifs:** To get the recognition motif nucleotide sequences.

**weights:** To get nuclease weights.

**cutSites:** To get nuclease cut sites.

### See Also

See the [CrisprNuclease](#) for CRISPR-specific nucleases.

### Examples

```
EcoRI <- Nuclease("EcoRI",
                 motifs=c("G^AATTC"),
                 metadata=list(description="EcoRI restriction enzyme"))
```

---

nickaseName	<i>An S4 class to represent a nickase</i>
-------------	---

---

### Description

An S4 class to represent a nickase

**Usage**

```
nickaseName(object)

nickaseName(object) <- value

nickingStrand(object)

nickingStrand(object) <- value

Nickase(
  nickaseName,
  nickingStrand = c("original", "opposite"),
  motifs = NULL,
  cutSites = NULL,
  weights = rep(1, length(motifs)),
  metadata = list()
)

## S4 method for signature 'Nickase'
show(object)

## S4 method for signature 'Nickase'
nickaseName(object)

## S4 replacement method for signature 'Nickase'
nickaseName(object) <- value

## S4 method for signature 'Nickase'
nickingStrand(object)

## S4 replacement method for signature 'Nickase'
nickingStrand(object) <- value

## S4 method for signature 'Nickase'
weights(object, expand = FALSE)

## S4 replacement method for signature 'Nickase'
weights(object) <- value

## S4 method for signature 'Nickase'
isCutting(object)

## S4 method for signature 'Nickase'
motifs(
  object,
  primary = FALSE,
  strand = c("+", "-"),
  expand = FALSE,
```

```

    as.character = FALSE
  )

  ## S4 method for signature 'Nickase'
  motifLength(object)

  ## S4 method for signature 'Nickase'
  cutSites(object, combine = TRUE)

```

### Arguments

object	<a href="#">Nickase</a> object.
value	New value to pass to the setter functions.
nickaseName	Name of the nickase.
nickingStrand	String specifying with strand with respect to the motif sequence (5' to 3') is nicked. Must be either "original" (default) or "opposite".
motifs	Character vector of recognition sequence motifs written from 5' to 3' written in Rebase convention. If the point of cleavage has been determined, the precise site is marked with ^. Only letters in the IUPAC code are accepted. For nickases that cleave away from their recognition sequence, the cleavage sites are indicated in parentheses. See details for more information.
cutSites	Vector specifying the cleavage coordinates relative to the first nucleotide of the motif sequence. Each column corresponds to a motif specified in the motifs slot.
weights	Optional numeric vector specifying relative weights for the recognition motifs to specify cleavage probabilities.
metadata	Optional list providing global metadata information.
expand	Should sequences be expanded to only contain ATCG nucleotides? FALSE by default.
primary	Should only the motif with the highest weight be returned? FALSE by default. Only relevant if weights are stored in the <a href="#">Nickase</a> object.
strand	Strand to allow reverse complementation of the motif. "+" by default.
as.character	Should the motif sequences be returned as a character vector? FALSE by default.
combine	Should only unique values be considered? TRUE by default.

### Value

A [Nickase](#) object

### Functions

- `Nickase()`: Create a [Nickase](#) object

**Slots**

nickaseName Name of the nickase

motifs DNASTringSet of recognition sequence motifs written from 5' to 3'.

nickingStrand String specifying with strand with respect to the motif sequence (5' to 3') is nicked. Must be either "original" (default) or "opposite".

cutSites Vector specifying the cleavage coordinates relative to the first nucleotide of the motif sequence. Each column corresponds to a motif specified in the motifs slot.

weights Optional numeric vector specifying relative weights for the motifs corresponding to cleavage probabilities.

metadata Optional string providing a description of the nickase.

**Constructors**

Use the constructor `link{Nickase}` to create a Nickase object.

**Accessors**

nickaseName: To get the name of the nickase.

nickingStrand: To get the nicking strand.

metadata: To get the metadata list of the nickase

motifs: To get the recognition motif nucleotide sequences.

weights: To get nickase weights.

cutSites: To get nickase cut sites.

**See Also**

See the [CrisprNickase](#) for CRISPR-specific nickases.

**Examples**

```
Nb.BsmI <- Nickase("Nb.BsmI",  
                 motifs=c("GAATG^C"),  
                 nickingStrand="opposite",  
                 metadata=list(description="Nb.BsmI nicking enzyme."))
```

---

plotEditingWeights      *Quick plot to visualize editing weights*

---

### Description

Quick plot to visualize editing weights from a BaseEditor object.

### Usage

```
plotEditingWeights(  
  baseEditor,  
  discardEmptyRows = TRUE,  
  substitutions = NULL,  
  ylim = c(0, 1),  
  ...  
)
```

### Arguments

baseEditor	A <a href="#">BaseEditor</a> object.
discardEmptyRows	Should rows that have all weight equal to 0 be discarded? TRUE by default.
substitutions	Character vector specifying substitutions to be plotted. If NULL (default), all substitutions are shown.
ylim	Vector of length 2 specifying the y plot limits.
...	Additional arguments to be passed to plot

### Value

Nothing. A plot is generated as a side effect.

### Examples

```
if (interactive()){  
  data(BE4max, package="crisprBase")  
  plotEditingWeights(BE4max)  
}
```

---

restrictionEnzymes      *List of Nuclease objects representing common restriction enzymes*

---

**Description**

List of Nuclease objects representing common restriction enzymes from REBASE database.

**Usage**

```
data(restrictionEnzymes, package="crisprBase")
```

**Format**

List of Nuclease objects.

**Details**

List of Nuclease objects representing common restriction enzymes from REBASE database.

---

SaCas9      *SaCas9 CrisprNuclease object*

---

**Description**

CrisprNuclease object for the wildtype Staphylococcus aureus Cas9 (SaCas9) nuclease.

**Usage**

```
data(SaCas9, package="crisprBase")
```

**Format**

CrisprNuclease object.

**Details**

The AsCas9 nuclease recognizes NNGRRT PAM sequences. Spacer sequences must be located upstream of PAM sequences. Editing weights were obtained from [doi:10.1038/nature14299](https://doi.org/10.1038/nature14299).

---

spacerLength	<i>An S4 class to represent a CRISPR nuclease.</i>
--------------	--

---

**Description**

An S4 class to represent a CRISPR nuclease.

**Usage**

```
spacerLength(object, ...)  
targetLength(object, ...)  
pamLength(object, ...)  
spacerGap(object)  
hasSpacerGap(object)  
spacerGap(object) <- value  
spacerLength(object) <- value  
pamSide(object, ...)  
pamSide(object) <- value  
pams(object, ...)  
pamIndices(object, ...)  
spacerIndices(object, ...)  
prototypeSequence(object, ...)  
  
CrisprNuclease(  
  nucleaseName,  
  targetType = c("DNA", "RNA"),  
  pams = NA_character_,  
  weights = rep(1, length(pams)),  
  metadata = list(),  
  pam_side = NA_character_,  
  spacer_gap = 0L,  
  spacer_length = NA_integer_  
)  
  
## S4 method for signature 'CrisprNuclease'
```

```

show(object)

## S4 method for signature 'CrisprNuclease'
pamLength(object)

## S4 method for signature 'CrisprNuclease'
spacerLength(object)

## S4 replacement method for signature 'CrisprNuclease'
spacerLength(object) <- value

## S4 method for signature 'CrisprNuclease'
pamSide(object)

## S4 replacement method for signature 'CrisprNuclease'
pamSide(object) <- value

## S4 method for signature 'CrisprNuclease'
spacerGap(object)

## S4 replacement method for signature 'CrisprNuclease'
spacerGap(object) <- value

## S4 method for signature 'CrisprNuclease'
hasSpacerGap(object)

## S4 method for signature 'CrisprNuclease'
targetLength(object)

## S4 method for signature 'CrisprNuclease'
pams(object, primary = TRUE, ignore_pam = FALSE, as.character = FALSE)

## S4 method for signature 'CrisprNuclease'
pamIndices(object)

## S4 method for signature 'CrisprNuclease'
spacerIndices(object)

## S4 method for signature 'CrisprNuclease'
prototypeSequence(object, primary = TRUE)

```

### Arguments

object	<a href="#">CrisprNuclease</a> object.
...	Additional arguments for class-specific methods
value	For spacerLength<- and gapLength<-, must be a non-negative integer. For pamSide, must be either '5prime' or '3prime'.
nucleaseName	Name of the CRISPR nuclease.

targetType	String specifying target type ("DNA" or "RNA").
pams	Character vector of PAM sequence motifs written from 5' to 3'. If the point of cleavage has been determined, the precise site is marked with ^. Only letters in the IUPAC code are accepted. For nucleases that cleave away from their recognition sequence, the cleavage sites are indicated in parentheses. See details for more information.
weights	Optional numeric vector specifying relative weights of the PAM sequences to specify cleavage probabilities.
metadata	Optional list providing global metadata information.
pam_side	String specifying the side of the PAM sequence with respect to the protospacer sequence. Must be either '3prime' (e.g. Cas9) or '5prime' (e.g. Cas12a)
spacer_gap	Integer specifying the length (in nucleotides) between the spacer sequence and the PAM sequence (e.g. 0 for Cas9 and Cas12a).
spacer_length	Integer specifying the length of the spacer sequence
primary	Should only the PAM sequence with the heighest weight be returned? If no cleavage weights are stored in the <a href="#">CrisprNuclease</a> object, all sequences are returned. TRUE by default.
ignore_pam	Should all possible k-mer sequences for a given PAM length be returned, ir-respectively of the PAM sequence motifs stored in the <a href="#">CrisprNuclease</a> object? FALSE by default.
as.character	Should the PAM sequences be returned as a character vector? FALSE by default.

**Value**

A [CrisprNuclease](#) object

**Functions**

- [CrisprNuclease\(\)](#): Create a [CrisprNuclease](#) object

**Slots**

pam_side	String specifying the side of the PAM sequence with respect to the protospacer sequence. Must be either '3prime' (e.g. SpCas9) or '5prime' (e.g. AsCas12a)
spacer_length	Integer specifying the length of the spacer sequence
spacer_gap	Integer specifying the length (in nucleotides) between the spacer sequence and the PAM sequence (e.g. 0 for SpCas9 and AsCas12a).

**Constructors**

Use the constructor `link{CrisprNuclease}` to create a [CrisprNuclease](#) object.

**Accessors**

- nucleaseName: To get the name of the CRISPR nuclease.
- spacerLength: To return the length of the spacer sequence.
- targetLength: To return the length of the target sequence (protospacer + pam).
- pamLength: To return the length of the PAM sequence.
- pamSide: To return the side of the PAM sequence with respect to the spacer sequence.
- spacerGap: To return the length of the gap between the PAM and spacer sequences.
- pams: To return the list of PAM sequences.

**Setters**

- spacerGap<-: To change the length of the gap between the PAM and spacer sequences.
- pamSide<-: To change the side of the PAM sequence with respect to the protospacer sequence.
- spacerLength<-: To change the length of the spacer sequence.

**Utility functions for genomic arithmetics**

- pamIndices: To return the relative coordinates of the PAM sequence within the target sequence.
- spacerIndices: To return the relative coordinates of the spacer sequence within the target sequence.

**Examples**

```
SpCas9 <- CrisprNuclease("SpCas9",
  pams=c("(3/3)NGG", "(3/3)NAG", "(3/3)NGA"),
  weights=c(1, 0.2593, 0.0694),
  metadata=list(description="Wildtype Streptococcus
    pyogenes Cas9 (SpCas9) nuclease"),
  pam_side="3prime",
  spacer_length=20)
```

---

 SpCas9

*SpCas9 CrisprNuclease object*


---

**Description**

CrisprNuclease object for the wildtype Streptococcus pyogenes Cas9 (SpCas9) nuclease.

**Usage**

```
data(SpCas9, package="crisprBase")
```

**Format**

CrisprNuclease object.

**Details**

The SpCas9 nuclease recognizes NGG PAM sequences. Spacer sequences must be located upstream of PAM sequences.

---

SpG

*SpG CrisprNuclease object*

---

**Description**

CrisprNuclease object for the engineered *Streptococcus pyogenes* Cas9 SpG nuclease.

**Usage**

```
data(SpG, package="crisprBase")
```

**Format**

CrisprNuclease object.

**Details**

The SpG nuclease recognizes NGN PAM sequences. Spacer sequences must be located upstream of PAM sequences. Editing weights were obtained from [doi:10.1038/s41467-022-28884-7](https://doi.org/10.1038/s41467-022-28884-7).

# Index

## \* datasets

- AsCas12a, [3](#)
  - BE4max, [6](#)
  - CasRx, [7](#)
  - Csm, [10](#)
  - enAsCas12a, [11](#)
  - MAD7, [19](#)
  - restrictionEnzymes, [28](#)
  - SaCas9, [28](#)
  - SpCas9, [32](#)
  - SpG, [33](#)
- annotateMismatches, [2](#)
- AsCas12a, [3](#)
- BaseEditor, [5](#), [16](#), [27](#)
- BaseEditor (baseEditorName), [4](#)
- BaseEditor-class (baseEditorName), [4](#)
- baseEditorName, [4](#)
- baseEditorName, BaseEditor-method (baseEditorName), [4](#)
- baseEditorName<- (baseEditorName), [4](#)
- baseEditorName<- , BaseEditor-method (baseEditorName), [4](#)
- BE4max, [6](#)
- CasRx, [7](#)
- CrisprNickase, [9](#), [26](#)
- CrisprNickase (CrisprNickase-class), [7](#)
- CrisprNickase-class, [7](#)
- CrisprNuclease, [5](#), [14](#), [23](#), [30](#), [31](#)
- CrisprNuclease (spacerLength), [29](#)
- CrisprNuclease-class (spacerLength), [29](#)
- Csm, [10](#)
- cutSites (motifs), [20](#)
- cutSites, Nickase-method (nickaseName), [23](#)
- cutSites, Nuclease-method (motifs), [20](#)
- editingStrand (baseEditorName), [4](#)
- editingStrand, BaseEditor-method (baseEditorName), [4](#)
- editingStrand<- (baseEditorName), [4](#)
- editingStrand<- , BaseEditor-method (baseEditorName), [4](#)
- editingWeights (baseEditorName), [4](#)
- editingWeights, BaseEditor-method (baseEditorName), [4](#)
- enAsCas12a, [11](#)
- extractPamFromTarget, [11](#)
- extractProtospacerFromTarget, [12](#)
- getAvailableCrisprNucleases, [13](#)
- getCutSiteFromPamSite, [14](#)
- getCutSiteRanges, [14](#)
- getEditingSiteFromPamSite, [15](#)
- getPamRanges, [16](#)
- getProtospacerRanges, [17](#)
- getTargetRanges, [18](#)
- hasSpacerGap (spacerLength), [29](#)
- hasSpacerGap, CrisprNickase-method (CrisprNickase-class), [7](#)
- hasSpacerGap, CrisprNuclease-method (spacerLength), [29](#)
- isCutting (motifs), [20](#)
- isCutting, Nickase-method (nickaseName), [23](#)
- isCutting, Nuclease-method (motifs), [20](#)
- isDnase (motifs), [20](#)
- isDnase, Nuclease-method (motifs), [20](#)
- isRnase (motifs), [20](#)
- isRnase, Nuclease-method (motifs), [20](#)
- MAD7, [19](#)
- motifLength (motifs), [20](#)
- motifLength, Nickase-method (nickaseName), [23](#)
- motifLength, Nuclease-method (motifs), [20](#)

- motifs, 20
- motifs, Nickase-method (nickaseName), 23
- motifs, Nuclease-method (motifs), 20
- Nickase, 25
- Nickase (nickaseName), 23
- Nickase-class (nickaseName), 23
- nickaseName, 23
- nickaseName, Nickase-method (nickaseName), 23
- nickaseName<- (nickaseName), 23
- nickaseName<-, Nickase-method (nickaseName), 23
- nickingStrand (nickaseName), 23
- nickingStrand, Nickase-method (nickaseName), 23
- nickingStrand<- (nickaseName), 23
- nickingStrand<-, Nickase-method (nickaseName), 23
- Nuclease, 22
- Nuclease (motifs), 20
- Nuclease-class (motifs), 20
- nucleaseName (motifs), 20
- nucleaseName, Nuclease-method (motifs), 20
- nucleaseName<- (motifs), 20
- nucleaseName<-, Nuclease-method (motifs), 20
- pamIndices (spacerLength), 29
- pamIndices, CrisprNickase-method (CrisprNickase-class), 7
- pamIndices, CrisprNuclease-method (spacerLength), 29
- pamLength (spacerLength), 29
- pamLength, CrisprNickase-method (CrisprNickase-class), 7
- pamLength, CrisprNuclease-method (spacerLength), 29
- pams (spacerLength), 29
- pams, CrisprNickase-method (CrisprNickase-class), 7
- pams, CrisprNuclease-method (spacerLength), 29
- pamSide (spacerLength), 29
- pamSide, CrisprNickase-method (CrisprNickase-class), 7
- pamSide, CrisprNuclease-method (spacerLength), 29
- pamSide<- (spacerLength), 29
- pamSide<-, CrisprNickase-method (CrisprNickase-class), 7
- pamSide<-, CrisprNuclease-method (spacerLength), 29
- plotEditingWeights, 27
- prototypeSequence (spacerLength), 29
- prototypeSequence, CrisprNickase-method (CrisprNickase-class), 7
- prototypeSequence, CrisprNuclease-method (spacerLength), 29
- restrictionEnzymes, 28
- SaCas9, 28
- setEditingWeights (baseEditorName), 4
- show, BaseEditor-method (baseEditorName), 4
- show, CrisprNickase-method (CrisprNickase-class), 7
- show, CrisprNuclease-method (spacerLength), 29
- show, Nickase-method (nickaseName), 23
- show, Nuclease-method (motifs), 20
- spacerGap (spacerLength), 29
- spacerGap, CrisprNickase-method (CrisprNickase-class), 7
- spacerGap, CrisprNuclease-method (spacerLength), 29
- spacerGap<- (spacerLength), 29
- spacerGap<-, CrisprNickase-method (CrisprNickase-class), 7
- spacerGap<-, CrisprNuclease-method (spacerLength), 29
- spacerIndices (spacerLength), 29
- spacerIndices, CrisprNickase-method (CrisprNickase-class), 7
- spacerIndices, CrisprNuclease-method (spacerLength), 29
- spacerLength, 29
- spacerLength, CrisprNickase-method (CrisprNickase-class), 7
- spacerLength, CrisprNuclease-method (spacerLength), 29
- spacerLength<- (spacerLength), 29
- spacerLength<-, CrisprNickase-method (CrisprNickase-class), 7
- spacerLength<-, CrisprNuclease-method (spacerLength), 29

SpCas9, [32](#)  
SpG, [33](#)

targetLength (spacerLength), [29](#)  
targetLength, CrisprNickase-method  
(CrisprNickase-class), [7](#)  
targetLength, CrisprNuclease-method  
(spacerLength), [29](#)  
targetType (motifs), [20](#)  
targetType, Nuclease-method (motifs), [20](#)  
targetType<- (motifs), [20](#)  
targetType<-, Nuclease-method (motifs),  
[20](#)

weights (motifs), [20](#)  
weights, Nickase-method (nickaseName), [23](#)  
weights, Nuclease-method (motifs), [20](#)  
weights<- (motifs), [20](#)  
weights<- , Nickase-method (nickaseName),  
[23](#)  
weights<- , Nuclease-method (motifs), [20](#)