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Title Analysis of Metabolomics GC/MS Data

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Collate Oclasses.R clusterAlignment.R init.R multipleAlignment.R
peaksAlignment.R progressiveAlignment.R betweenAlignment.R dp.R
metrics.R parse.R peaksDataset.R gatherInfo.R plotFragments.R
rmaFitUnit.R addXCMSPeaks.R retFatMatrix.R exportSpectra.R
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Contents

addAMDISPeaks	2
addChromaTOFPeaks	4
addXCMSPeaks	5
betweenAlignment	7
calcTimeDiffs	8
clusterAlignment	9
corPrt	11
deDuper	12
distToLib	13
dp	13
dynRT	15
eitherMatrix-class	16
exportSpectra	16
gatherInfo	17
headToTailPlot	18
importSpec	19
imputePeaks	20
matchSpec	21
multipleAlignment-class	22
ndpRT	24
normDotProduct	25
parseChromaTOF	27
parseELU	28
peaksAlignment-class	29
peaksDataset	32
plotAlignedFrag	33
plotAlignment,peaksAlignment-method	34
plotChrom,peaksDataset-method	36
plotClustAlignment,clusterAlignment-method	38
plotFrag	39
plotImage	40
progressiveAlignment-class	42
retFatMatrix	43
rmaFitUnit	45
show,multipleAlignment-method	46
Index	48

addAMDISPeaks	<i>Add AMDIS peak detection results</i>
---------------	---

Description

Reads ASCII ELU-format files (output from AMDIS) and attaches them to an already created peaksDataset object

Usage

```
addAMDISPeaks(object, fns = dir(, "[Eu][Ll][Uu]"), verbose = TRUE, ...)
```

Arguments

object	a peaksDataset object.
fns	character vector of same length as object@rawdata (user ensures the order matches)
verbose	whether to give verbose output, default TRUE
...	arguments passed on to parseELU

Details

Repeated calls to parseELU to add peak detection results to the original peaksDataset object.

Value

peaksDataset object

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[parseELU](#), [peaksDataset](#)

Examples

```
# need access to CDF (raw data) and ELU files
require(gcspikelite)
gcmsPath<-paste(find.package("gcspikelite"), "data", sep="/")

# full paths to file names
cdfFiles<-dir(gcmsPath, "CDF", full=TRUE)
eluFiles<-dir(gcmsPath, "ELU", full=TRUE)

# create a 'peaksDataset' object and add AMDIS peaks to it
pd<-peaksDataset(cdfFiles[1], mz=seq(50, 550), rtrange=c(7.5, 8.5))
pd<-addAMDISPeaks(pd, eluFiles[1])
```

addChromaTOFPeaks *Add ChromaTOF peak detection results*

Description

Reads ASCII tab-delimited format files (output from ChromaTOF) and attaches them to an already created peaksDataset object

Usage

```
addChromaTOFPeaks(  
  object,  
  fns = dir( "[Tt][Xx][Tx]" ),  
  rtDivide = 60,  
  verbose = TRUE,  
  ...  
)
```

Arguments

object	a peaksDataset object.
fns	character vector of same length as object@rawdata (user ensures the order matches)
rtDivide	number giving the amount to divide the retention times by.
verbose	whether to give verbose output, default TRUE
...	arguments passed on to parseChromaTOF

Details

Repeated calls to parseChromaTOF to add peak detection results to the original peaksDataset object.

Value

peaksDataset object

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[parseChromaTOF](#), [peaksDataset](#)

Examples

```
# need access to CDF (raw data) and ChromaTOF files
require(gcspikelite)
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")

# full paths to file names
cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
# [not run] cTofFiles<-dir(gcmsPath,"txt",full=TRUE)

# create a 'peaksDataset' object and add ChromaTOF peaks to it
pd<-peaksDataset(cdfFiles[1],mz=seq(50,550),rtrange=c(7.5,8.5))
# [not run] pd<-addChromTOFPeaks(pd,...)
```

addXCMSPeaks

addXCMSPeaks

Description

Add xcms/CAMERA peak detection results

Usage

```
addXCMSPeaks(
  files,
  object,
  settings = list(),
  minintens = 100,
  minfeat = 6,
  BPPARAM = bpparam(),
  multipleMF = FALSE,
  multipleMFParam = list(fwhm = c(5, 10, 15), mz.abs = 0.2, rt.abs = 2)
)
```

Arguments

files	list of chromatogram files
object	a peakDataset object
settings	see findPeaks-matchedFilter findPeaks-centWave
minintens	minimum ion intensity to be included into a pseudospectra
minfeat	minimum number of ion to be created a pseudospectra
BPPARAM	a parameter class specifying if and how parallel processing should be performed

multipleMF	logical Try to remove redundant peaks, in this case where there are any peaks within an absolute m/z value of 0.2 and within 3 s for any one sample in the xcmsSet (the largest peak is kept)
multipleMFParam	list. It contains the settings for the peak-picking. mz_abs represent the the mz range; rt_abs represent thert range
mz.abs	mz range
rt.abs	rt range

Details

Reads the raw data using xcms, group each extracted ion according to their retention time using CAMERA and attaches them to an already created peaksDataset object

Repeated calls to xcmsSet and annotate to perform peak-picking and deconvolution. The peak detection results are added to the original peaksDataset object. Two peak detection alorithms are available: continuous wavelet transform (peakPicking=c('cwt')) and the matched filter approach (peakPicking=c('mF')) described by Smith et al (2006). For further information consult the xcms package manual.

Value

peaksDataset object

Author(s)

Riccardo Romoli <riccardo.romoli@unifi.it>

See Also

[peaksDataset](#) [findPeaks-matchedFilter](#) [findPeaks-centWave](#) [xcmsRaw-class](#)

Examples

```
files <- list.files(path = paste(find.package("gcspikelite"), "data",
                               sep = "/"), "CDF", full = TRUE)
data <- peaksDataset(files[1:2], mz = seq(50, 550), rtrange = c(7.5, 8.5))
## create settings object
mfp <- xcms::MatchedFilterParam(fwhm = 10, snthresh = 5)
cwt <- xcms::CentWaveParam()
data <- addXCMSPeaks(files[1:2], data, settings = mfp, multipleMF = FALSE)
data
```

betweenAlignment *Data Structure for "between" alignment of many GCMS samples*

Description

This function creates a "between" alignment (i.e. comparing merged peaks)

Usage

```
betweenAlignment(
  pD,
  cAList,
  pAList,
  impList,
  filterMin = 1,
  gap = 0.7,
  D = 10,
  usePeaks = TRUE,
  df = 30,
  verbose = TRUE,
  metric = 2,
  type = 2,
  penalty = 0.2,
  compress = FALSE
)
```

Arguments

pD	a peaksDataset object
cAList	list of clusterAlignment objects, one for each experimental group
pAList	list of progressiveAlignment objects, one for each experimental group
impList	list of imputation lists
filterMin	minimum number of peaks within a merged peak to be kept in the analysis
gap	gap parameter
D	retention time penalty parameter
usePeaks	logical, whether to use peaks (if TRUE) or the full 2D profile alignment (if FALSE)
df	distance from diagonal to calculate similarity
verbose	logical, whether to print information
metric	numeric, different algorithm to calculate the similarity matrix between two mass spectrum. metric=1 call normDotProduct(); metric=2 call ndpRT(); metric=3 call corPrt()
type	numeric, two different type of alignment function
penalty	penalization applied to the matching between two mass spectra if $(t_1 - t_2) > D$
compress	logical whether to compress the similarity matrix into a sparse format.

Details

betweenAlignment objects gives the data structure which stores the result of an alignment across several "pseudo" datasets. These pseudo datasets are constructed by merging the "within" alignments.

Value

betweenAlignment object

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[multipleAlignment](#)

Examples

```
require(gcspikelite)
## see 'multipleAlignment'
```

calcTimeDiffs

Calculate retention time shifts from profile alignments

Description

This function takes the set of all pairwise profile alignments and use these to estimate retention time shifts between each pair of samples. These will then be used to normalize the retention time penalty of the signal peak alignment.

Usage

```
calcTimeDiffs(pd, ca.full, verbose = TRUE)
```

Arguments

pd	a peaksDataset object
ca.full	a clusterAlignment object, fit with
verbose	logical, whether to print out information

Details

Using the set of profile alignments,

Value

list of same length as `ca.full@alignments` with the matrices giving the retention time penalties.

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[peaksAlignment](#), [clusterAlignment](#)

Examples

```
require(gcspikelite)

# paths and files
gcmsPath <- paste(find.package("gcspikelite"), "data", sep="/")
cdfFiles <- dir(gcmsPath, "CDF", full=TRUE)
eluFiles <- dir(gcmsPath, "ELU", full=TRUE)

# read data, peak detection results
pd <- peaksDataset(cdfFiles[1:2], mz=seq(50, 550), rtrange=c(7.5, 8.5))
pd <- addAMDISPeaks(pd, eluFiles[1:2])

# pairwise alignment using all scans
fullca <- clusterAlignment(pd, usePeaks=FALSE, df=100)

# calculate retention time shifts
timedf <- calcTimeDiffs(pd, fullca)
```

clusterAlignment	<i>Data Structure for a collection of all pairwise alignments of GCMS runs</i>
------------------	--

Description

Store the raw data and optionally, information regarding signal peaks for a number of GCMS runs

Usage

```
clusterAlignment(  
  pD,  
  runs = 1:length(pD@rawdata),  
  timedf = NULL,  
  usePeaks = TRUE,  
  verbose = TRUE,  
  ...  
)
```

Arguments

pD	a peaksDataset object.
runs	vector of integers giving the samples to calculate set of pairwise alignments over.
timedf	list (length = the number of pairwise alignments) of matrices giving the expected time differences expected at each pair of peaks used with usePeaks=TRUE, passed to peaksAlignment
usePeaks	logical, TRUE uses peakdata list, FALSE uses rawdata list for computing similarity.
verbose	logical, whether to print out info.
...	other arguments passed to peaksAlignment

Details

clusterAlignment computes the set of pairwise alignments.

Value

clusterAlignment object

Author(s)

Mark Robinson, Riccardo Romoli

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[peaksDataset](#), [peaksAlignment](#)

Examples

```
require(gcspikelite)

# paths and files
gcmsPath <- paste(find.package("gcspikelite"), "data", sep="/")
cdfFiles <- dir(gcmsPath, "CDF", full=TRUE)
eluFiles <- dir(gcmsPath, "ELU", full=TRUE)

# read data, peak detection results
pd <- peaksDataset(cdfFiles[1:2], mz=seq(50,550), rtrange=c(7.5,8.5))
pd <- addAMDISPeaks(pd, eluFiles[1:2])

ca <- clusterAlignment(pd, gap=0.5, D=0.05, df=30, metric=1, type=1)
```

corPrt

Retention Time Penalized Correlation

Description

This function calculates the similarity of all pairs of peaks from 2 samples, using the spectra similarity and the retention time differences

Usage

```
corPrt(d1, d2, t1, t2, D, penalty = 0.2)
```

Arguments

d1	data matrix for sample 1
d2	data matrix for sample 2
t1	vector of retention times for sample 1
t2	vector of retention times for sample 2
D	retention time window for the matching
penalty	penalization applied to the matching between two mass spectra if $(t1-t2)>D$

Details

Computes the Pearson correlation between every pair of peak vectors in the retention time window (D) and returns the similarity matrix.

Value

matrix of similarities

Author(s)

Riccardo Romoli

See Also[peaksAlignment](#)**Examples**

```
## Not Run
require(gcspikelite)
files <- list.files(path = paste(find.package("gcspikelite"), "data",
                               sep = "/"), "CDF", full = TRUE)
data <- peaksDataset(files[1:2], mz = seq(50, 550), rtrange = c(7.5, 8.5))
## create settings object
mfp <- xcms::MatchedFilterParam(fwhm = 10, snthresh = 5)
cwt <- xcms::CentWaveParam()
data <- addXCMSPeaks(files[1:2], data, settings = mfp, multipleMF = FALSE)
data
## review peak picking
plotChrom(data, rtrange=c(7.5, 10.5), runs=c(1:2))

r <- corPrt(data@peaksdata[[1]], data@peaksdata[[2]],
            data@peaksrt[[1]], data@peaksrt[[2]], D = 50, penalty = 0.2)
## End (Not Run)
```

deDuper

deDuper

Description

Duplicate peak removal function

Usage

```
deDuper(object, mz.abs = 0.1, rt.abs = 2)
```

Arguments

object	xcms object
mz.abs	mz range
rt.abs	rt range

Details

Remove redundant peaks, in this case where there are any peaks within an absolute m/z value of 0.2 and within 3 s for any one sample in the xcmsSet (the largest peak is kept)

Value

an object of xcms class

Author(s)

r

distToLib	<i>distToLib</i>
-----------	------------------

Description

The function calculate the distance between each mas spec in the msp file and the aligned mass spec from each sampe

Usage

```
distToLib(mspLib, outList)
```

Arguments

mspLib	a .msp file from NIST
outList	an object from gatherInfo()

Details

Return the distance matrix

Value

the distance matrix between the mass spec and the aligned spec

Author(s)

Riccardo Romoli

dp	<i>Dynamic programming algorithm, given a similarity matrix</i>
----	---

Description

This function calls C code for a bare-bones dynamic programming algorithm, finding the best cost path through a similarity matrix.

Usage

```
dp(M, gap = 0.5, big = 1e+10, verbose = FALSE)
```

Arguments

M	similarity matrix
gap	penalty for gaps
big	large value used for matrix margins
verbose	logical, whether to print out information

Details

This is a pretty standard implementation of a bare-bones dynamic programming algorithm, with a single gap parameter and allowing only simple jumps through the matrix (up, right or diagonal).

Value

list with element match with the set of pairwise matches.

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[normDotProduct](#)

Examples

```
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
pd<-peaksDataset(cdfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))
pd<-addAMDISPeaks(pd,eluFiles[1:2])

# similarity matrix
r<-normDotProduct(pd@peaksdata[[1]],pd@peaksdata[[2]])

# dynamic-programming-based matching of peaks
v<-dp(r,gap=.5)
```

dynRT

dynRT

Description

Dynamic Retention Time Based Alignment algorithm, given a similarity matrix

Usage

```
dynRT(S)
```

Arguments

S similarity matrix

Details

This function align two chromatograms finding the maximum similarity among the mass spectra

Value

list containing the matched peaks between the two chromatograms. The number represent position of the spectra in the S matrix

Author(s)

riccardo.romoli@unifi.it

Examples

```
require(gcspikelite)
files <- list.files(path = paste(find.package("gcspikelite"), "data",
                               sep = "/"), "CDF", full = TRUE)
data <- peaksDataset(files[1:2], mz = seq(50, 550), rtrange = c(7.5, 8.5))
## create settings object
mfp <- xcms::MatchedFilterParam(fwhm = 10, snthresh = 5)
cwt <- xcms::CentWaveParam()
data <- addXCMSPeaks(files[1:2], data, settings = mfp, multipleMF = FALSE)
data
## review peak picking
plotChrom(data, rtrange=c(7.5, 10.5), runs=c(1:2))
## similarity
r <- ndpRT(data@peaksdata[[1]], data@peaksdata[[2]], data@peaksrt[[1]],
           data@peaksrt[[2]], D = 50)
## dynamic retention time based alignment algorithm
v <- dynRT(S = r)
```

eitherMatrix-class *A class description*

Description

A class description

exportSpectra *exportSpectra*

Description

Write the mass spectrum into a .msp file to be used in NIST search.

Usage

```
exportSpectra(object, outList, spectra, normalize = TRUE)
```

Arguments

object	an object of class "peaksDataset"
outList	an object created using the gatherInfo() function
spectra	numeric. The number of the mass spectra to be printed. It correspond to the number of the peak in the plot() and the number of the peak in the gatherInfo() list.
normalize	logical. If the mass spectra has to be normalized to 100

Details

Write the mass spectrum into a .msp file to be used in NIST search.

Value

a .msp file

Author(s)

riccardo.romoli@unifi.com

gatherInfo

*Gathers abundance informations from an alignment***Description**

Given an alignment table (indices of matched peaks across several samples) such as that within a `progressiveAlignment` or `multipleAlignment` object, this routines goes through the raw data and collects the abundance of each fragment peak, as well as the retention times across the samples.

Usage

```
gatherInfo(
  pD,
  obj,
  newind = NULL,
  method = c("apex"),
  findmzind = TRUE,
  useTIC = FALSE,
  top = NULL,
  intensity.cut = 0.05
)
```

Arguments

<code>pD</code>	a <code>peaksDataset</code> object, to get the abundance data from
<code>obj</code>	either a <code>multipleAlignment</code> or <code>progressiveAlignment</code> object
<code>newind</code>	list giving the
<code>method</code>	method used to gather abundance information, only <code>apex</code> implemented currently.
<code>findmzind</code>	logical, whether to take a subset of all <code>m/z</code> indices
<code>useTIC</code>	logical, whether to use total ion current for abundance summaries
<code>top</code>	only use the top <code>top</code> peaks
<code>intensity.cut</code>	percentage of the maximum intensity

Details

This procedure loops through the the table of matched peaks and gathers the

Value

Returns a list (of lists) for each row in the alignment table. Each list has 3 elements:

<code>mz</code>	a numerical vector of the <code>m/z</code> fragments used
<code>rt</code>	a numerical vector for the exact retention time of each peak across all samples
<code>data</code>	matrix of fragment intensities. If <code>useTIC = TRUE</code> , this matrix will have a single row

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[imputePeaks](#)

Examples

```
require(gcspikelite)

## paths and files
gcmsPath <- paste(find.package("gcspikelite"), "data", sep = "/")
cdfFiles <- dir(gcmsPath, "CDF", full = TRUE)
eluFiles <- dir(gcmsPath, "ELU", full = TRUE)

## read data, peak detection results
pd <- peaksDataset(cdfFiles[1:2], mz = seq(50, 550), rtrange = c(7.5, 8.5))
pd <- addAMDISPeaks(pd, eluFiles[1:2])

## multiple alignment
ma <- multipleAlignment(pd, c(1,1), wn.gap = 0.5, wn.D = 0.05, bw.gap = 0.6,
                        bw.D = 0.2, usePeaks = TRUE, filterMin = 1, df = 50,
                        verbose = TRUE, metric = 1, type = 1)

## gather apex intensities
d <- gatherInfo(pd, ma)

## table of retention times
nm <- list(paste("MP", 1:length(d), sep = ""), c("S1", "S2"))
rts <- matrix(unlist(sapply(d, .subset, "rt")), byrow = TRUE, nc = 2,
             dimnames = nm)
```

headToTailPlot

Head to tail plot

Description

The head-to-tail-plot for the mass spectra

Usage

```
headToTailPlot(specFromLib, specFromList)
```

Arguments

specFromLib the mass spectra obtained from the .msp file
specFromList the mass spectra obtained from [gatherInfo](#)

Details

Head-to-tail-plot to visually compare the mass spectra

Value

the plot

Author(s)

Riccardo Romoli

importSpec

importSpec

Description

Read the mass spectra from an external msp file

Usage

```
importSpec(file)
```

Arguments

file a .msp file from NIST search library database

Details

Read the mass spectra from an external file in msp format. The format is used in NIST search library database.

Value

list containing the mass spectra

Author(s)

riccardo.romoli@unifi.it

`imputePeaks`*Imputation of locations of peaks that were undetected*

Description

Using the information within the peaks that are matched across several runs, we can impute the location of the peaks that are undetected in a subset of runs

Usage

```
imputePeaks(pD, obj, typ = 1, obj2 = NULL, filterMin = 1, verbose = TRUE)
```

Arguments

<code>pD</code>	a peaksDataset object
<code>obj</code>	the alignment object, either <code>multipleAlignment</code> or <code>progressiveAlignment</code> , that is used to infer the unmatched peak locations
<code>typ</code>	type of imputation to do, 1 for simple linear interpolation (default), 2 only works if <code>obj2</code> is a <code>clusterAlignment</code> object
<code>obj2</code>	a <code>clusterAlignment</code> object
<code>filterMin</code>	minimum number of peaks within a merged peak to impute
<code>verbose</code>	logical, whether to print out information

Details

If you are aligning several samples and for a (small) subset of the samples in question, a peak is undetected, there is information within the alignment that can be useful in determining where the undetected peak is, based on the surrounding matched peaks. Instead of moving forward with missing values into the data matrices, this procedure goes back to the raw data and imputes the location of the apex (as well as the start and end), so that we do not need to bother with post-hoc imputation or removing data because of missing components.

We realize that imputation is prone to error and prone to attributing intensity from neighbouring peaks to the unmatched peak. We argue that this is still better than having to deal with these in statistical models after that fact. This may be an area of future improvement.

Value

list with 3 elements apex, start and end, each masked matrices giving the scan numbers of the imputed peaks.

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[multipleAlignment](#), [progressiveAlignment](#), [peaksDataset](#)

Examples

```
require(gcspikelite)

## paths and files
gcmsPath <- paste(find.package("gcspikelite"), "data", sep = "/")
cdfFiles <- dir(gcmsPath,"CDF", full = TRUE)
eluFiles <- dir(gcmsPath,"ELU", full = TRUE)

## read data, peak detection results
pd <- peaksDataset(cdfFiles[1:3], mz = seq(50,550), rtrange = c(7.5,8.5))
pd <- addAMDISPeaks(pd, eluFiles[1:3])

## alignments
ca <- clusterAlignment(pd, gap = 0.5, D = 0.05, df = 30, metric = 1, type =
  1, compress = FALSE)
pa <- progressiveAlignment(pd, ca, gap = 0.6, D = 0.1, df = 30,
  compress = FALSE)

v <- imputePeaks(pd, pa, filterMin = 1)
```

matchSpec

matchSpec

Description

Calculate the distance between a reference mass spectrum

Usage

```
matchSpec(spec1, outList, whichSpec)
```

Arguments

spec1	reference mass spectrum
outList	the return of gatherInfo
whichSpec	the entry number of outList

Details

Calculate the distance between a reference mass spectrum and one from the sample

Value

the distance between the reference mass spectrum and the others

Author(s)

Riccardo Romoli

multipleAlignment-class

Data Structure for multiple alignment of many GCMS samples

Description

Store the raw data and optionally, information regarding signal peaks for a number of GCMS runs

Usage

```
multipleAlignment(  
  pd,  
  group,  
  bw.gap = 0.8,  
  wn.gap = 0.6,  
  bw.D = 0.2,  
  wn.D = 0.05,  
  filterMin = 1,  
  lite = FALSE,  
  usePeaks = TRUE,  
  df = 50,  
  verbose = TRUE,  
  timeAdjust = FALSE,  
  doImpute = FALSE,  
  metric = 2,  
  type = 2,  
  penalty = 0.2,  
  compress = FALSE  
)
```

Arguments

pd	a peaksDataset object
group	factor variable of experiment groups, used to guide the alignment algorithm
bw.gap	gap parameter for "between" alignments

wn.gap	gap parameter for "within" alignments
bw.D	distance penalty for "between" alignments. When type = 2 represent the retention time window expressed in seconds
wn.D	distance penalty for "within" alignments. When type = 2 represent the retention time window expressed in seconds
filterMin	minimum number of peaks within a merged peak to be kept in the analysis
lite	logical, whether to keep "between" alignment details (default, FALSE)
usePeaks	logical, whether to use peaks (if TRUE) or the full 2D profile alignment (if FALSE)
df	distance from diagonal to calculate similarity
verbose	logical, whether to print information
timeAdjust	logical, whether to use the full 2D profile data to estimate retention time drifts (Note: time required)
doImpute	logical, whether to impute the location of unmatched peaks
metric	numeric, different algorithm to calculate the similarity matrix between two mass spectrum. metric=1 call normDotProduct(); metric=2 call ndpRT(); metric=3 call corPrt()
type	numeric, two different type of alignment function
penalty	penalization applied to the matching between two mass spectra if $(t_1 - t_2) > D$
compress	logical whether to compress the similarity matrix into a sparse format.

Details

multipleAlignment is the data structure giving the result of an alignment across several GCMS runs. Multiple alignments are done progressively. First, all samples with the same tg\$Group label will be aligned (denoted a "within" alignment). Second, each group will be summarized into a pseudo-data set, essentially a spectrum and retention time for each matched peak of the within-alignment. Third, these "merged peaks" are aligned in the same progressive manner, here called a "between" alignment.

Value

multipleAlignment object

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[peaksDataset](#), [betweenAlignment](#), [progressiveAlignment](#)

Examples

```
require(gcspikelite)

## paths and files
gcmsPath <- paste(find.package("gcspikelite"), "data", sep = "/")
cdfFiles <- dir(gcmsPath, "CDF", full = TRUE)
eluFiles <- dir(gcmsPath, "ELU", full = TRUE)

## read data, peak detection results
pd <- peaksDataset(cdfFiles[1:2], mz = seq(50, 550), rtrange = c(7.5, 8.5))
pd <- addAMDISPeaks(pd, eluFiles[1:2])

## multiple alignment
ma <- multipleAlignment(pd, c(1, 1), wn.gap = 0.5, wn.D = 0.05, bw.gap = 0.6,
                        bw.D = 0.2, usePeaks = TRUE, filterMin = 1, df = 50,
                        verbose = TRUE, metric = 1, type = 1)
```

ndpRT

Retention Time Penalized Normalized Dot Product

Description

This function calculates the similarity of all pairs of peaks from 2 samples, using the spectra similarity and the retention time differences

Usage

```
ndpRT(s1, s2, t1, t2, D)
```

Arguments

s1	data matrix for sample 1
s2	data matrix for sample 2
t1	vector of retention times for sample 1
t2	vector of retention times for sample 2
D	retention time window for the matching

Details

Computes the normalized dot product between every pair of peak vectors in the retention time window (D) and returns a similarity matrix.

Value

matrix of similarities

Author(s)

Riccardo Romoli

See Also[peaksAlignment](#)**Examples**

```
## Not Run
require(gcspikelite)
files <- list.files(path = paste(find.package("gcspikelite"), "data",
                               sep = "/"), "CDF", full = TRUE)
data <- peaksDataset(files[1:2], mz = seq(50, 550), rtrange = c(7.5, 8.5))
## create settings object
mfp <- xcms::MatchedFilterParam(fwhm = 10, snthresh = 5)
cwt <- xcms::CentWaveParam()
data <- addXCMSPeaks(files[1:2], data, settings = mfp, multipleMF = FALSE)
data
## review peak picking
plotChrom(data, rtrange = c(7.5, 10.5), runs = c(1:2))

r <- ndpRT(data@peaksdata[[1]], data@peaksdata[[2]],
           data@peaksrt[[1]], data@peaksrt[[2]], D = 50)
## End (Not Run)
```

normDotProduct

Normalized Dot Product

Description

This function calculates the similarity of all pairs of peaks from 2 samples, using the spectra similarity

Usage

```
normDotProduct(
  x1,
  x2,
  t1 = NULL,
  t2 = NULL,
  df = max(ncol(x1), ncol(x2)),
  D = 1e+05,
  timedf = NULL,
  verbose = FALSE
)
```

Arguments

x1	data matrix for sample 1
x2	data matrix for sample 2
t1	vector of retention times for sample 1
t2	vector of retention times for sample 2
df	distance from diagonal to calculate similarity
D	retention time penalty
timedf	matrix of time differences to normalize to. if NULL, 0 is used.
verbose	logical, whether to print out information

Details

Efficiently computes the normalized dot product between every pair of peak vectors and returns a similarity matrix. C code is called.

Value

matrix of similarities

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[dp](#), [peaksAlignment](#)

Examples

```
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"), "data", sep="/")
cdfFiles<-dir(gcmsPath, "CDF", full=TRUE)
eluFiles<-dir(gcmsPath, "ELU", full=TRUE)

# read data, peak detection results
pd<-peaksDataset(cdfFiles[1:2], mz=seq(50, 550), rtrange=c(7.5, 8.5))
pd<-addAMDISPeaks(pd, eluFiles[1:2])

r<-normDotProduct(pd@peaksdata[[1]], pd@peaksdata[[2]])
```

parseChromaTOF	<i>Parser for ChromaTOF files</i>
----------------	-----------------------------------

Description

Reads ASCII ChromaTOF-format files from AMDIS (Automated Mass Spectral Deconvolution and Identification System)

Usage

```
parseChromaTOF(  
  fn,  
  min.pc = 0.01,  
  mz = seq(85, 500),  
  rt.cut = 0.008,  
  rtrange = NULL,  
  skip = 1,  
  rtDivide = 60  
)
```

Arguments

fn	ChromaTOF filename to read.
min.pc	minimum percent of maximum intensity.
mz	vector of mass-to-charge bins of raw data table.
rt.cut	the difference in retention time, below which peaks are merged together.
rtrange	retention time range to parse peaks from, can speed up parsing if only interested in a small region (must be numeric vector of length 2)
skip	number of rows to skip at beginning of the ChromaTOF
rtDivide	multiplier to divide the retention times by (default: 60)

Details

parseChromaTOF will typically be called by [addChromaTOFPeaks](#), not called directly.

Peaks that are detected within `rt.cut` are merged together. This avoids peaks which are essentially overlapping.

Fragments that are less than `min.pc` of the maximum intensity fragment are discarded.

Value

list with components `peaks` (table of spectra – rows are mass-to-charge and columns are the different detected peaks) and `tab` (table of features for each detection), according to what is stored in the ChromaTOF file.

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[addAMDISPeaks](#)

Examples

```
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
tofFiles<-dir(gcmsPath,"tof",full=TRUE)

# parse ChromaTOF file
cTofList<-parseChromaTOF(tofFiles[1])
```

parseELU

Parser for ELU files

Description

Reads ASCII ELU-format files from AMDIS (Automated Mass Spectral Deconvolution and Identification System)

Usage

```
parseELU(f, min.pc = 0.01, mz = seq(50, 550), rt.cut = 0.008, rtrange = NULL)
```

Arguments

f	ELU filename to read.
min.pc	minimum percent of maximum intensity.
mz	vector of mass-to-charge bins of raw data table.
rt.cut	the difference in retention time, below which peaks are merged together.
rtrange	retention time range to parse peaks from, can speed up parsing if only interested in a small region (must be numeric vector of length 2)

Details

parseELU will typically be called by [addAMDISPeaks](#), not called directly.

Peaks that are detected within `rt.cut` are merged together. This avoids peaks which are essentially overlapping.

Fragments that are less than `min.pc` of the maximum intensity fragment are discarded.

Value

list with components `peaks` (table of spectra – rows are mass-to-charge and columns are the different detected peaks) and `tab` (table of features for each detection), according to what is stored in the ELU file.

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[addAMDISPeaks](#)

Examples

```
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# parse ELU file
eluList<-parseELU(eluFiles[1])
```

peaksAlignment-class *Data Structure for pairwise alignment of 2 GCMS samples*

Description

Store the raw data and optionally, information regarding signal peaks for a number of GCMS runs

Usage

```

peaksAlignment(
  d1,
  d2,
  t1,
  t2,
  gap = 0.5,
  D = 50,
  timedf = NULL,
  df = 30,
  verbose = TRUE,
  usePeaks = TRUE,
  compress = TRUE,
  metric = 2,
  type = 2,
  penalty = 0.2
)

```

Arguments

d1	matrix of MS intensities for 1st sample (if doing a peak alignment, this contains peak apexes/areas; if doing a profile alignment, this contains scan intensities. Rows are m/z bins, columns are peaks/scans.
d2	matrix of MS intensities for 2nd sample
t1	vector of retention times for 1st sample
t2	vector of retention times for 2nd sample
gap	gap penalty for dynamic programming algorithm. Not used if type=2
D	time window (on same scale as retention time differences, t1 and t2. Default scale is seconds.)
timedf	list (length = the number of pairwise alignments) of matrices giving the expected time differences expected at each pair of peaks used with usePeaks=TRUE.
df	integer, how far from the diagonal to go to calculate the similarity of peaks. Smaller value should run faster, but be careful not to choose too low.
verbose	logical, whether to print out info.
usePeaks	logical, TRUE uses peakdata list, FALSE uses rawdata list for computing similarity.
compress	logical, whether to compress the similarity matrix into a sparse format.
metric	numeric, different algorithm to calculate the similarity matrix between two mass spectrum. metric=1 call normDotProduct(); metric=2 call ndpRT(); metric=3 call corPrt()
type	numeric, two different type of alignment function
penalty	penalization applied to the matching between two mass spectra if (t1-t2)>D

Details

peaksAlignment is a hold-all data structure of the raw and peak detection data.

Value

peaksAlignment object

Author(s)

Mark Robinson, Riccardo Romoli

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[peaksDataset](#), [clusterAlignment](#)

Examples

```
## see clusterAlignment, it calls peaksAlignment

## Not Run:
files <- list.files(path = paste(find.package("gcspikelite"), "data",
                               sep = "/"), "CDF", full = TRUE)
data <- peaksDataset(files[1:2], mz = seq(50, 550), rtrange = c(7.5, 8.5))
## create settings object
mfp <- xcms::MatchedFilterParam(fwhm = 10, snthresh = 5)
cwt <- xcms::CentWaveParam(snthresh = 3, ppm = 3000, peakwidth = c(3, 40),
                           prefilter = c(3, 100), fitgauss = FALSE, integrate = 2, noise = 0,
                           extendLengthMSW = TRUE, mzCenterFun = "wMean")
data <- addXCMSPeaks(files[1:2], data, settings = mfp)
data
plotChrom(data, rtrange=c(7.5, 10.5), runs=c(1:2))

## align two chromatogram
pA <- peaksAlignment(data@peaksdata[[1]], data@peaksdata[[2]],
                    data@peaksrt[[1]], data@peaksrt[[2]], D = 50,
                    metric = 3, compress = FALSE, type = 2, penalty = 0.2)

plotAlignment(pA)
pA@v$match

par(mfrow=c(2,1))
plot(data@peaksdata[[1]][,15], type = 'h', main = paste(data@peaksrt[[1]][[15]]))
plot(data@peaksdata[[2]][,17], type = 'h',
     main = paste(data@peaksrt[[2]][[17]]))
## End (Not Run)
```

peaksDataset

Data Structure for raw GCMS data and peak detection results

Description

Store the raw data and optionally, information regarding signal peaks for a number of GCMS runs

Usage

```
peaksDataset(  
  fns = dir(, "[Cc][Dd][Ff]"),  
  verbose = TRUE,  
  mz = seq(50, 550),  
  rtDivide = 60,  
  rtrange = NULL  
)
```

Arguments

fns	character vector, filenames of raw data in CDF format.
verbose	logical, if TRUE then iteration progress information is output.
mz	vector giving bins of raw data table.
rtDivide	number giving the amount to divide the retention times by.
rtrange	retention time range to limit data to (must be numeric vector of length 2)

Details

peaksDataset is a hold-all data structure of the raw and peak detection data.

Value

peaksDataset object

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

Examples

```
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"), "data", sep="/")
cdfFiles<-dir(gcmsPath, "CDF", full=TRUE)
eluFiles<-dir(gcmsPath, "ELU", full=TRUE)

# read data
pd<-peaksDataset(cdfFiles[1:2], mz=seq(50, 550), rtrange=c(7.5, 8.5))
show(pd)
```

plotAlignedFrag *plotAlignedFrag*

Description

Plot the aligned mass spectra

Usage

```
plotAlignedFrag(
  object,
  outList,
  specID,
  fullRange = TRUE,
  normalize = TRUE,
  ...
)
```

Arguments

object	where to keep the mass range of the experiment
outList	where to keep the mass spectra; both abundance than m/z
specID	a vector containing the index of the spectra to be plotted. Is referred to outList
fullRange	if TRUE uses the mass range of the whole experiment, otherwise uses only the mass range of each plotted spectrum
normalize	if TRUE normalize the intensity of the mass peak to 100, the most abundant is 100% and the other peaks are scaled consequentially
...	further arguments passed to the 'plot' command

Details

Plot the deconvoluted and aligned mass spectra collected using gatherInfo()

Author(s)

Riccardo Romoli (riccardo.romoli@unifi.it)

Examples

```
files <- list.files(path = paste(find.package("gcspikelite"), "data",
                               sep = "/"), "CDF", full = TRUE)
data <- peaksDataset(files[1:4], mz = seq(50, 550), rtrange = c(7.5, 8.5))
## create settings object
mfp <- xcms::MatchedFilterParam(fwhm = 10, snthresh = 5)
cwt <- xcms::CentWaveParam(snthresh = 3, ppm = 3000, peakwidth = c(3, 40),
  prefilter = c(3, 100), fitgauss = FALSE, integrate = 2, noise = 0,
  extendLengthMSW = TRUE, mzCenterFun = "wMean")
data <- addXCMSPeaks(files[1:4], data, settings = mfp)
data
## multiple alignment
ma <- multipleAlignment(data, c(1,1,2,2), wn.gap = 0.5, wn.D = 0.05,
  bw.gap = 0.6, bw.D = 0.2, usePeaks = TRUE, filterMin = 1, df = 50,
  verbose = TRUE, metric = 2, type = 2)

## gather apex intensities
gip <- gatherInfo(data, ma)
gip[[33]]
plotAlignedFragments(object = data, outList = gip, specID = 33)
```

plotAlignment, peaksAlignment-method
plotAlignment

Description

Plotting functions for GCMS data objects

Usage

```
## S4 method for signature 'peaksAlignment'
plotAlignment(
  object,
  xlab = "Peaks - run 1",
  ylab = "Peaks - run 2",
  plotMatches = TRUE,
  matchPch = 19,
  matchLwd = 3,
  matchCex = 0.5,
  matchCol = "black",
  col = colorpanel(50, "white", "green", "navyblue"),
  breaks = seq(0, 1, length = 51),
  ...
)
```

Arguments

object	a clusterAlignment object
xlab	x-axis label
ylab	y-axis label
plotMatches	logical, whether to plot matches
matchPch	match plotting character
matchLwd	match line width
matchCex	match character expansion factor
matchCol	match colour
col	vector of colours for colour scale
breaks	vector of breaks for colour scale
...	further arguments passed to image

Details

Plot an object of [peaksAlignment](#)

The similarity matrix is plotted and optionally, the set of matching peaks. clusterAlignment objects are just a collection of all pairwise peakAlignment objects.

Value

plot an object of class [peaksAlignment](#)

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[peaksAlignment](#) [plotAlignment](#)

Examples

```
require(gcspikelite)
files <- list.files(path = paste(find.package("gcspikelite"), "data",
                               sep = "/"), "CDF", full = TRUE)
data <- peaksDataset(files[1:2], mz = seq(50, 550), rtrange = c(7.5, 8.5))
## create settings object
mfp <- xcms::MatchedFilterParam(fwhm = 10, snthresh = 5)
cwt <- xcms::CentWaveParam(snthresh = 3, ppm = 3000, peakwidth = c(3, 40),
                           prefilter = c(3, 100), fitgauss = FALSE, integrate = 2, noise = 0,
                           extendLengthMSW = TRUE, mzCenterFun = "wMean")
```

```

data <- addXCMSPeaks(files[1:2], data, settings = mfp)
data
## image plot
plotChrom(data, rtrange = c(7.5,8.5), plotPeaks = TRUE, plotPeakLabels =TRUE)

## align two chromatogram
pA <- peaksAlignment(data@peaksdata[[1]], data@peaksdata[[2]],
                     data@peaksrt[[1]], data@peaksrt[[2]], D = 50,
                     compress = FALSE, type = 1, metric = 1,
                     gap = 0.5)
plotAlignment(pA)

```

plotChrom,peaksDataset-method

Plotting functions for GCMS data objects

Description

Store the raw data and optionally, information regarding signal peaks for a number of GCMS runs

Usage

```

## S4 method for signature 'peaksDataset'
plotChrom(
  object,
  runs = 1:length(object@rawdata),
  mzind = 1:nrow(object@rawdata[[1]]),
  mind = NULL,
  plotSampleLabels = TRUE,
  calcGlobalMax = FALSE,
  peakCex = 0.8,
  plotPeaks = TRUE,
  plotPeakBoundaries = FALSE,
  plotPeakLabels = FALSE,
  plotMergedPeakLabels = TRUE,
  mlwd = 3,
  usePeaks = TRUE,
  plotAcrossRuns = FALSE,
  overlap = F,
  rtrange = NULL,
  cols = NULL,
  thin = 1,
  max.near = median(object@rawrt[[1]]),
  how.near = 50,
  scale.up = 1,
  ...
)

```

Arguments

object	a peaksDataset object.
runs	set of run indices to plot
mzind	set of mass-to-charge indices to sum over (default, all)
mind	matrix of aligned indices
plotSampleLabels	logical, whether to display sample labels
calcGlobalMax	logical, whether to calculate an overall maximum for scaling
peakCex	character expansion factor for peak labels
plotPeaks	logical, whether to plot hashes for each peak
plotPeakBoundaries	logical, whether to display peak boundaries
plotPeakLabels	logical, whether to display peak labels
plotMergedPeakLabels	logical, whether to display 'merged' peak labels
mlwd	line width of lines indicating the alignment
usePeaks	logical, whether to plot alignment of peaks (otherwise, scans)
plotAcrossRuns	logical, whether to plot across peaks when unmatched peak is given
overlap	logical, whether to plot TIC/XICs overlapping
rtrange	vector of length 2 giving start and end of the X-axis
cols	vector of colours (same length as the length of runs)
thin	when usePeaks=FALSE, plot the alignment lines every thin values
max.near	where to look for maximum
how.near	how far away from max.near to look
scale.up	a constant factor to scale the TICs
...	further arguments passed to the plot

Details

Each TIC is scale to the maximum value (as specified by the how.near and max.near values). The many parameters gives considerable flexibility of how the TICs can be visualized.

Value

plot the chromatograms

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also[peaksDataset](#)**Examples**

```
require(gcspikelite)

## paths and files
gcmsPath <- paste(find.package("gcspikelite"), "data", sep="/")
cdfFiles <- dir(gcmsPath, "CDF", full=TRUE)
eluFiles <- dir(gcmsPath, "ELU", full=TRUE)

## read data
pd <- peaksDataset(cdfFiles[1:3], mz=seq(50,550), rrange=c(7.5,8.5))

## image plot
plotChrom(pd, rrange = c(7.5,8.5), plotPeaks = TRUE,
          plotPeakLabels = TRUE)
```

plotClustAlignment, clusterAlignment-method
plotClustAlignment

Description

Plotting functions for GCMS data objects

Usage

```
## S4 method for signature 'clusterAlignment'
plotClustAlignment(object, alignment = 1, ...)
```

Arguments

<code>object</code>	clusterAlignment object.
<code>alignment</code>	the set of alignments to plot
<code>...</code>	further arguments passed to image. See also plotAlignment

Details

For clusterAlignment objects, the similarity matrix is plotted and optionally, the set of matching peaks. clusterAlignment objects are just a collection of all pairwise peakAlignment objects.

Value

plot the pairwise alignment

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[plotAlignment](#)

Examples

```
require(gcspikelite)

# paths and files
gcmsPath <- paste(find.package("gcspikelite"), "data", sep="/")
cdfFiles <- dir(gcmsPath, "CDF", full=TRUE)
eluFiles <- dir(gcmsPath, "ELU", full=TRUE)

# read data, peak detection results
pd <- peaksDataset(cdfFiles[1:2], mz=seq(50,550), rtrange=c(7.5,8.5))
pd <- addAMDISPeaks(pd, eluFiles[1:2])

ca <- clusterAlignment(pd, gap=0.5, D=0.05, df=30, metric=1, type=1)
plotClustAlignment(ca, run = 1)
plotClustAlignment(ca, run = 2)
plotClustAlignment(ca, run = 3)
```

plotFrag

plotFrag

Description

Plot the mass spectra from the profile matrix

Usage

```
plotFrag(object, sample, specID, normalize = TRUE, ...)
```

Arguments

object	an object of class "peaksDataset" where to keep the mass spectra; both abundance (y) than m/z (x)
sample	character, the sample from were to plot the mass spectra
specID	numerical, a vector containing the index of the spectra to be plotted.

normalize logical, if TRUE normalize the intensity of the mass peak to 100, the most abundant is 100 consequentially

... other parameter passed to the plot() function

Details

Plot the deconvoluted mass spectra from the profile matrix

Author(s)

riccardo.romoli@unifi.it

Examples

```
files <- list.files(path = paste(find.package("gcspikelite"), "data",
                               sep = "/"), "CDF", full = TRUE)
data <- peaksDataset(files[1:2], mz = seq(50, 550), rtrange = c(7.5, 8.5))
## create settings object
mfp <- xcms::MatchedFilterParam(fwhm = 10, snthresh = 5)
cwt <- xcms::CentWaveParam(snthresh = 3, ppm = 3000, peakwidth = c(3, 40),
                           prefilter = c(3, 100), fitgauss = FALSE, integrate = 2, noise = 0,
                           extendLengthMSW = TRUE, mzCenterFun = "wMean")
data <- addXCMSPeaks(files[1:2], data, settings = mfp)
data
## align two chromatogram
pA <- peaksAlignment(data@peaksdata[[1]], data@peaksdata[[2]],
                    data@peaksrt[[1]], data@peaksrt[[2]], D = 50,
                    metric = 3, compress = FALSE, type = 2, penalty = 0.2)
pA@v$match
## plot the mass spectra
par(mfrow=c(2,1))
plotFrag(object=data, sample=1, specID=10)
plotFrag(object=data, sample=2, specID=12)
```

plotImage

Plot of images of GCMS data

Description

Image plots (i.e. 2D heatmaps) of raw GCMS profile data

Usage

```
## S4 method for signature 'peaksDataset'
plotImage(
  object,
  run = 1,
  rtrange = c(11, 13),
```

```
    main = NULL,  
    mzrange = c(50, 200),  
    SCALE = log2,  
    ...  
  )
```

Arguments

object	a peaksDataset object
run	index of the run to plot an image for
rtrange	vector of length 2 giving start and end of the X-axis (retention time)
main	main title (auto-constructed if not specified)
mzrange	vector of length 2 giving start and end of the Y-axis (mass-to-charge ratio)
SCALE	function called to scale the data (default: log2)
...	further arguments passed to the image command

Details

For peakDataset objects, each TIC is scale to the maximum value (as specified by the `how.near` and `max.near` values). The many parameters gives considerable flexibility of how the TICs can be visualized.

For peakAlignment objects, the similarity matrix is plotted and optionally, the set of matching peaks. clusterAlignment objects are just a collection of all pairwise peakAlignment objects.

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[plot](#), [peaksDataset](#)

Examples

```
require(gcspikelite)  
  
# paths and files  
gcmsPath<-paste(find.package("gcspikelite"), "data", sep="/")  
cdfFiles<-dir(gcmsPath, "CDF", full=TRUE)  
eluFiles<-dir(gcmsPath, "ELU", full=TRUE)  
  
# read data  
pd<-peaksDataset(cdfFiles[1], mz=seq(50, 550), rtrange=c(7.5, 8.5))
```

```
# image plot
plotImage(pd,run=1,rtrange=c(7.5,8.5),main="")
```

progressiveAlignment-class

Data Structure for progressive alignment of many GCMS samples

Description

Performs a progressive peak alignment (clustalw style) of multiple GCMS peak lists

Usage

```
progressiveAlignment(
  pD,
  cA,
  D = 50,
  gap = 0.5,
  verbose = TRUE,
  usePeaks = TRUE,
  df = 30,
  compress = FALSE,
  type = 2
)
```

Arguments

pD	a peaksDataset object
cA	a clusterAlignment object
D	retention time penalty
gap	gap parameter
verbose	logical, whether to print information
usePeaks	logical, whether to use peaks (if TRUE) or the full 2D profile alignment (if FALSE)
df	distance from diagonal to calculate similarity
compress	logical, whether to store the similarity matrices in sparse form
type	numeric, two different type of alignment function

Details

The progressive peak alignment we implemented here for multiple GCMS peak lists is analogous to how `clustalw` takes a set of pairwise sequence alignments and progressively builds a multiple alignment. More details can be found in the reference below.

Value

progressiveAlignment object

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[peaksDataset](#), [multipleAlignment](#)

Examples

```
require(gcspikelite)
files <- list.files(path = paste(find.package("gcspikelite"), "data",
                               sep = "/"), "CDF", full = TRUE)
data <- peaksDataset(files[1:2], mz = seq(50, 550), rtrange = c(7.5, 8.5))
## create settings object
mfp <- xcms::MatchedFilterParam(fwhm = 10, snthresh = 5)
cwt <- xcms::CentWaveParam(snthresh = 3, ppm = 3000, peakwidth = c(3, 40),
                           prefilter = c(3, 100), fitgauss = FALSE, integrate = 2, noise = 0,
                           extendLengthMSW = TRUE, mzCenterFun = "wMean")
data <- addXCMSPeaks(files[1:2], data, settings = mfp)
data
ca <- clusterAlignment(data, gap = 0.5, D = 0.05, df = 30, metric = 1,
                       type = 1, compress = FALSE)
pa <- progressiveAlignment(data, ca, gap = 0.6, D = 0.1, df = 30,
                           type = 1, compress = FALSE)
```

retFatMatrix

retFatMatrix

Description

Build a fat data matrix

Usage

```
retFatMatrix(object, data, minFilter = round(length(object@files)/3 * 2))
```

Arguments

object	peakDataset object
data	a gatherInfo() object
minFilter	the minimum number for a feature to be returned in the data matrix. Default is 2/3 of the samples

Details

This function allows to extract the data from an object created using `gatherInfo` and build a data matrix using the area of the deconvoluted and aligned peaks. The row are the samples while the column represent the different peaks.

Value

A fat data matrix containing the area of the deconvoluted and aligned peaks. The row are the samples while the column represent the different peaks

Author(s)

Riccardo Romoli <riccardo.romoli@unifi.it>

See Also

[gatherInfo](#)

Examples

```
require(gcspikelite)
files <- list.files(path = paste(find.package("gcspikelite"), "data",
                               sep = "/"), "CDF", full = TRUE)
data <- peaksDataset(files[1:2], mz = seq(50, 550), rtrange = c(7.5, 8.5))
## create settings object
mfp <- xcms::MatchedFilterParam(fwhm = 10, snthresh = 5)
cwt <- xcms::CentWaveParam(snthresh = 3, ppm = 3000, peakwidth = c(3, 40),
                           prefilter = c(3, 100), fitgauss = FALSE, integrate = 2, noise = 0,
                           extendLengthMSW = TRUE, mzCenterFun = "wMean")
data <- addXCMSPeaks(files[1:2], data, settings = mfp)
data
ma <- multipleAlignment(pd = data, group = c(1,1),
                       filterMin = 1, metric = 2, type = 2)
outList <- gatherInfo(data, ma)
mtxD <- retFatMatrix(object = data, data = outList, minFilter = 1)
```

`rmaFitUnit`*Fits a robust linear model (RLM) for one metabolite*

Description

Using `r1m` from MASS, this procedure fits a linear model using all the fragments

Usage

```
rmaFitUnit(  
  u,  
  maxit = 5,  
  mzEffect = TRUE,  
  cls = NULL,  
  fitSample = TRUE,  
  fitOrCoef = c("coef", "fit"),  
  TRANSFORM = log2  
)
```

Arguments

<code>u</code>	a metabolite unit (list object with vectors <code>mz</code> and <code>rt</code> for <code>m/z</code> and retention times, respectively and a data element giving the <code>fragmentxsample</code> intensity matrix)
<code>maxit</code>	maximum number of iterations (default: 5)
<code>mzEffect</code>	logical, whether to fit <code>m/z</code> effect (default: TRUE)
<code>cls</code>	class variable
<code>fitSample</code>	whether to fit individual samples (alternative is fit by group)
<code>fitOrCoef</code>	whether to return a vector of coefficients (default: "coef"), or an <code>r1m</code> object ("fit")
<code>TRANSFORM</code>	function to transform the raw data to before fitting (default: <code>log2</code>)

Details

Fits a robust linear model.

Value

list giving elements of fragment and sample coefficients (if `fitOrCoef="coef"`) or a list of elements from the fitting process (if `fitOrCoef="fit"`)

Author(s)

Mark Robinson

References

Mark D Robinson (2008). Methods for the analysis of gas chromatography - mass spectrometry data *PhD dissertation* University of Melbourne.

See Also

[peaksAlignment](#), [clusterAlignment](#)

Examples

```
require(gcspikelite)

# paths and files
gcmsPath<-paste(find.package("gcspikelite"),"data",sep="/")
cdfFiles<-dir(gcmsPath,"CDF",full=TRUE)
eluFiles<-dir(gcmsPath,"ELU",full=TRUE)

# read data, peak detection results
pd<-peaksDataset(cdfFiles[1:2],mz=seq(50,550),rtrange=c(7.5,8.5))
pd<-addAMDISPeaks(pd,eluFiles[1:2])

# pairwise alignment using all scans
fullca<-clusterAlignment(pd, usePeaks = FALSE, df = 100)

# calculate retention time shifts
timedf<-calcTimeDiffs(pd, fullca)
```

show,multipleAlignment-method

*Store the raw data and optionally, information regarding signal peaks
for a number of GCMS runs*

Description

multipleAlignment is the data structure giving the result of an alignment across several GCMS runs. Multiple alignments are done progressively. First, all samples with the same `tg$Group` label will be aligned (denoted a "within" alignment). Second, each group will be summarized into a pseudo-data set, essentially a spectrum and retention time for each matched peak of the within-alignment. Third, these "merged peaks" are aligned in the same progressive manner, here called a "between" alignment.

Usage

```
## S4 method for signature 'multipleAlignment'  
show(object)
```

Arguments

object multipleAlignment object

Author(s)

Mark Robinson

Index

- * **classes**
 - [betweenAlignment](#), [7](#)
 - [clusterAlignment](#), [9](#)
 - [multipleAlignment-class](#), [22](#)
 - [peaksAlignment-class](#), [29](#)
 - [peaksDataset](#), [32](#)
 - [plotAlignment](#), [peaksAlignment-method](#), [34](#)
 - [plotChrom](#), [peaksDataset-method](#), [36](#)
 - [plotClustAlignment](#), [clusterAlignment-method](#), [38](#)
 - [plotImage](#), [40](#)
 - [progressiveAlignment-class](#), [42](#)
- * **gatherInfo()**
 - [plotAlignedFragments](#), [33](#)
- * **manip**
 - [addAMDISPeaks](#), [2](#)
 - [addChromaTOFPeaks](#), [4](#)
 - [addXCMSPeaks](#), [5](#)
 - [calcTimeDiffs](#), [8](#)
 - [corPrt](#), [11](#)
 - [dp](#), [13](#)
 - [gatherInfo](#), [17](#)
 - [imputePeaks](#), [20](#)
 - [ndpRT](#), [24](#)
 - [normDotProduct](#), [25](#)
 - [parseChromaTOF](#), [27](#)
 - [parseELU](#), [28](#)
 - [rmaFitUnit](#), [45](#)
- * **plot()**
 - [plotAlignedFragments](#), [33](#)
- [addAMDISPeaks](#), [2](#), [28](#), [29](#)
- [addChromaTOFPeaks](#), [4](#), [27](#)
- [addXCMSPeaks](#), [5](#)
- [betweenAlignment](#), [7](#), [23](#)
- [betweenAlignment-class](#)
 - [\(betweenAlignment\)](#), [7](#)
- [betweenAlignment-method](#)
 - [\(betweenAlignment\)](#), [7](#)
- [betweenAlignment-show](#)
 - [\(betweenAlignment\)](#), [7](#)
- [calcTimeDiffs](#), [8](#)
- [clusterAlignment](#), [9](#), [9](#), [31](#), [46](#)
- [clusterAlignment-class](#)
 - [\(clusterAlignment\)](#), [9](#)
- [clusterAlignment-plot](#)
 - [\(clusterAlignment\)](#), [9](#)
- [clusterAlignment-show](#)
 - [\(clusterAlignment\)](#), [9](#)
- [corPrt](#), [11](#)
- [deDuper](#), [12](#)
- [distToLib](#), [13](#)
- [dp](#), [13](#), [26](#)
- [dynRT](#), [15](#)
- [eitherMatrix-class](#), [16](#)
- [exportSpectra](#), [16](#)
- [gatherInfo](#), [17](#), [19](#), [21](#), [44](#)
- [headToTailPlot](#), [18](#)
- [importSpec](#), [19](#)
- [imputePeaks](#), [18](#), [20](#)
- [matchSpec](#), [21](#)
- [multipleAlignment](#), [8](#), [21](#), [43](#)
- [multipleAlignment](#)
 - [\(multipleAlignment-class\)](#), [22](#)
- [multipleAlignment-class](#), [22](#)
- [multipleAlignment-class](#),
 - [\(multipleAlignment-class\)](#), [22](#)
- [multipleAlignment-method](#)
 - [\(multipleAlignment-class\)](#), [22](#)
- [multipleAlignment-show](#),
 - [\(multipleAlignment-class\)](#), [22](#)

ndpRT, 24
normDotProduct, 14, 25

parseChromaTOF, 5, 27
parseELU, 3, 28
peaksAlignment, 9, 10, 12, 25, 26, 35, 46
peaksAlignment (peaksAlignment-class), 29
peaksAlignment-class, 29
peaksAlignment-plot (peaksAlignment-class), 29
peaksAlignment-show (peaksAlignment-class), 29
peaksDataset, 3, 5, 6, 10, 21, 23, 31, 32, 38, 41, 43
peaksDataset-class (peaksDataset), 32
peaksDataset-plot (peaksDataset), 32
peaksDataset-show (peaksDataset), 32
plot, 41
plot, clusterAlignment, ANY-method (clusterAlignment), 9
plot, clusterAlignment-method (clusterAlignment), 9
plot, peaksAlignment, ANY-method (peaksAlignment-class), 29
plot, peaksAlignment-method (peaksAlignment-class), 29
plot, peaksDataset, ANY-method (peaksDataset), 32
plot, peaksDataset-method (peaksDataset), 32
plotAlignedFragments, 33
plotAlignment, 35, 39
plotAlignment, peaksAlignment-method, 34
plotChrom, peaksDataset-method, 36
plotClustAlignment, clusterAlignment-method, 38
plotFragments, 39
plotImage, 40
plotImage, peaksDataset-method (plotImage), 40
progressiveAlignment, 21, 23
progressiveAlignment (progressiveAlignment-class), 42
progressiveAlignment-class, 42
progressiveAlignment-show (progressiveAlignment-class), 42

retFatMatrix, 43
rmaFitUnit, 45

show, (betweenAlignment), 7
show, clusterAlignment-method (clusterAlignment), 9
show, multipleAlignment-method, 46
show, peaksAlignment-method (peaksAlignment-class), 29
show, peaksDataset-method (peaksDataset), 32
show, progressiveAlignment-method (progressiveAlignment-class), 42