

# Package: lcmsPlot (via r-universe)

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**Type** Package

**Title** Comprehensive Liquid Chromatography-Mass Spectrometry (LC-MS)  
data visualisation package

**Version** 1.0.0

**Description** lcmsPlot is an R package designed for visualising Liquid Chromatography-Mass Spectrometry (LC-MS) data with publication-ready high-quality plots. The package enables users to generate and customise chromatograms, mass traces, spectra, and more with fine-tuned aesthetics and annotation options.

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## Contents

lcmsPlot-package . . . . .	3
+,lcmsPlotClass,function-method . . . . .	4
create_xcms_raw_list . . . . .	4
ExternalDataSource-class . . . . .	5
get_XCMSnExp_object_example . . . . .	6
iterate_plot_batches . . . . .	6
lcmsPlot . . . . .	7
lcmsPlotClass-class . . . . .	8
lcmsPlotDataContainer-class . . . . .	9
lp_arrange . . . . .	10
lp_chromatogram . . . . .	11
lp_compound_discoverer . . . . .	13
lp_facets . . . . .	14
lp_get_plot . . . . .	15
lp_grid . . . . .	16
lp_intensity_map . . . . .	17
lp_labels . . . . .	18
lp_layout . . . . .	19
lp_legend . . . . .	20
lp_mass_trace . . . . .	21
lp_peak_density . . . . .	22
lp_rt_diff_plot . . . . .	23
lp_rt_line . . . . .	24
lp_spectra . . . . .	25
lp_total_ion_current . . . . .	27
MsDialPeaksSource . . . . .	27
MZmineFeatureListsSource . . . . .	29
next_plot . . . . .	30

show,ExternalDataSource-method . . . . .	31
show,lcmsPlotClass-method . . . . .	32
show,lcmsPlotDataContainer-method . . . . .	33
show,XcmsRawList-method . . . . .	34
XcmsRawList . . . . .	34
XcmsRawList-class . . . . .	35

<b>Index</b>	<b>36</b>
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lcmsPlot-package	<i>lcmsPlot: Comprehensive Liquid Chromatography-Mass Spectrometry (LC-MS) data visualisation package</i>
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## Description

lcmsPlot offers flexible and powerful visualisation of raw and processed LC-MS data. It supports chromatograms, mass spectra, and other plot types, combining high performance with broad customisation. Designed for large datasets, it facilitates assessment of signal quality, feature comparison across samples, and generation of publication-ready figures.

## Details

### Main features

- Unified, intuitive, and ggplot2-like interface.
- Plot from different types of sources, such as raw files (e.g. mzML), XCMS objects (e.g., XCMSnExp), or Compound Discoverer results.
- Plot chromatograms, mass traces, spectra, and more.
- Combine different types of LC-MS data plots (e.g. chromatograms with spectra).
- Arrange plots in different ways according to metadata factors.
- Large-scale plotting through iterative batching.

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## See Also

Useful links:

- <https://github.com/computational-metabolomics/lcmsPlot>
- Report bugs at <https://github.com/computational-metabolomics/lcmsPlot/issues>

---

`+,lcmsPlotClass,function-method`

*Apply a function to an lcmsPlotClass object using the infix + operator*

---

### Description

This provides a convenient infix style for applying transformations to lcmsPlotClass objects.

### Usage

```
## S4 method for signature 'lcmsPlotClass,function'  
e1 + e2
```

### Arguments

`e1` An instance of class lcmsPlotClass.  
`e2` A function that takes an lcmsPlotClass object and returns another.

### Value

An instance of class lcmsPlotClass.

### Examples

```
raw_files <- dir(  
  system.file("cdf", package = "faahK0"),  
  full.names = TRUE,  
  recursive = TRUE)[1:5]  
  
p <- lcmsPlot(raw_files) +  
  lp_chromatogram(aggregation_fun = "max") +  
  lp_arrange(group_by = "sample_id") +  
  lp_legend(position = "bottom") +  
  lp_labels(legend = "Sample")
```

---

`create_xcms_raw_list` *Create an XcmsRawList from raw LC-MS files*

---

### Description

Reads one or more raw MS files using `xcms::xcmsRaw()` and wraps the results in an `XcmsRawList`. Optionally runs the reads in parallel via `BiocParallel`.

**Usage**

```
create_xcms_raw_list(  
  paths,  
  profstep = 1,  
  mslevel = NULL,  
  scanrange = NULL,  
  BPPARAM = NULL  
)
```

**Arguments**

paths	A character vector of file paths to raw MS files (e.g. .mzML, .mzXML, .CDF).
profstep	A numeric value passed to <code>xcms::xcmsRaw()</code> controlling the mass bin size used to build the profile matrix. Defaults to 0, which skips profile matrix generation. Only set this to a positive value if you need the profile matrix for peak detection.
mslevel	A numeric value passed to <code>xcms::xcmsRaw()</code> indicating which MS level to load. NULL loads all levels. Defaults to NULL.
scanrange	A length-2 integer vector passed to <code>xcms::xcmsRaw()</code> restricting the range of scans to read. NULL reads all scans. Defaults to NULL.
BPPARAM	A <code>BiocParallelParam</code> object controlling parallel execution. When NULL (default) files are read sequentially.

**Value**

An `XcmsRawList` object with one `xcmsRaw` element per file.

**Examples**

```
paths <- dir(  
  system.file("cdf", package = "faahKO"),  
  full.names = TRUE,  
  recursive = TRUE  
)  
xl <- create_xcms_raw_list(paths)
```

---

ExternalDataSource-class

*External data source wrapper*

---

**Description**

An S4 class representing an external data source such as MZmine. This class acts as an adapter for external data sources by converting data to a common format (XCMS).

**Slots**

name A character value indicating the name of the data source.

metadata A data.frame representing the sample metadata.

peaks A data.frame representing the exported peaks.

---

```
get_XCMSnExp_object_example
```

*Get an XCMSnExp example object from the faahKO dataset*

---

**Description**

Get an XCMSnExp example object from the faahKO dataset

**Usage**

```
get_XCMSnExp_object_example(indices = c(1, 2, 3), should_group_peaks = FALSE)
```

**Arguments**

indices A numeric vector of sample indices to select from the faahKO CDF files. Defaults to the first three samples.

should\_group\_peaks

A logical value indicating whether to group the detected peaks.

**Value**

An XCMSnExp object containing raw data, detected chromatographic peaks, and, if requested, grouped features.

**Examples**

```
get_XCMSnExp_object_example(indices = 1:5)
```

---

```
iterate_plot_batches Iterate on the batches of plots
```

---

**Description**

iterate\_plot\_batches iterates over batches of plots defined by the batch\_size parameter passed to the lcmsPlot constructor function.

**Usage**

```
iterate_plot_batches(object, iter_fn)

## S4 method for signature 'lcmsPlotClass,function'
iterate_plot_batches(object, iter_fn)
```

**Arguments**

**object**            An instance of class `lcmsPlotClass`.  
**iter\_fn**            The function to apply to each item being iterated on.

**Value**

NULL (called for its side effect).

**Examples**

```
raw_files <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE)[1:5]

p <- lcmsPlot(raw_files, batch_size = 2) +
  lp_chromatogram(features = rbind(c(
    mzmin = 334.9,
    mzmax = 335.1,
    rtmin = 2700,
    rtmax = 2900))) +
  lp_arrange(group_by = "sample_id") +
  lp_legend(position = "bottom") +
  lp_labels(legend = "Sample")

pdf(tempfile(fileext = ".pdf"))
iterate_plot_batches(p, function(plot_obj) {
  print(plot_obj)
})
dev.off()
```

---

`lcmsPlot`*Create an `lcmsPlotClass` object*

---

**Description**

The `lcmsPlotClass` class allows a unified approach for the management of LC-MS data for the purpose of visualisation. It includes the options for customising the plot, the LC-MS data, and the underlying plot object. The `lcmsPlot` function is the main entry point and the preferred approach to creating `lcmsPlotClass` objects.

**Usage**

```
lcmsPlot(
  dataset,
  sample_id_column = "sample_id",
  metadata = NULL,
  batch_size = NULL,
  BPPARAM = NULL
)
```

**Arguments**

dataset	An object of type XCMSnExp, MsExperiment, MZmineSource, or character. If a character vector is supplied, it will be interpreted as a list of mzML paths.
sample_id_column	A character value indicating which column should be used as the sample ID. By default it is "sample_id".
metadata	A data.frame containing the samples metadata in case it is not provided in the dataset object.
batch_size	A numeric value indicating the number of samples per batch. This parameter is necessary when plotting multiple batches using the <code>iterate_plot_batches</code> or <code>next_plot</code> functions.
BPPARAM	A BiocParallelParam object for enabling parallelism. See <a href="#">BiocParallelParam</a> for more information.

**Value**

An instance of `lcmsPlotClass`. It will create the necessary internal structures related to the data (data slot) and options (options slot).

**Examples**

```
raw_files <- dir(
  system.file("cdf", package = "faahKO"),
  full.names = TRUE,
  recursive = TRUE)[1:5]

p <- lcmsPlot(raw_files)
```

---

lcmsPlotClass-class    *Managing LC-MS data for visualisation*

---

**Description**

The `lcmsPlotClass` class allows a unified approach for the management of LC-MS data for the purpose of visualisation. It includes the options for customising the plot, the LC-MS data, and the underlying plot object.

**Slots**

- options A list to store the plot options.
- data An instance of class lcmsPlotDataContainer.
- history A list to store the applied layers to generate a plot; for internal use.
- plot A patchwork object representing the underlying plot object.

**General information**

The lcmsPlotClass class has been designed to be the entry point for all data and outputs related to the lcmsPlot package. The class abstracts away the data handling, making it easier to use lcmsPlot with existing data wrappers like MsExperiment or XCMSnExp.

**Preferred usage**

The lcmsPlotClass class can be used directly to instantiate an object, however the preferred approach is to use the lcmsPlot function.

---

 lcmsPlotDataContainer-class

*A unified storing mechanism for LC-MS data*

---

**Description**

The lcmsPlotDataContainer class allows the storage of different types of LC-MS data. This class can be used independently from the plotting utilities, however the preferred approach is to use it with the lcmsPlotClass class.

**Slots**

- data\_obj The data object. One of: XCMSnExp, MsExperiment, MChromatograms, XChromatograms, XChromatogram, XcmsRawList, or character representing mzML paths.
- metadata A data.frame containing the sample metadata.
- chromatograms A data.frame containing the chromatograms.
- mass\_traces A data.frame containing the mass traces.
- spectra A data.frame containing the spectra.
- peak\_density A data.frame containing peak density curve data and optional feature-group rectangles, as produced by lp\_peak\_density().
- total\_ion\_current A data.frame containing the total ion current.
- intensity\_maps A data.frame containing the 2D intensity maps representing the distribution of detected peaks across m/z and RT.
- rt\_diff A data.frame containing the raw and adjusted RT values.
- feature\_metadata A data.frame containing feature/compound annotations attached to datasets through a column called feature\_metadata\_id.
- detected\_peaks A data.frame containing the detected peaks from an XCMSnExp or MsExperiment object.

---

`lp_arrange`*Define the arrangement of chromatograms*

---

### Description

The `lp_arrange` function specifies how chromatograms should be arranged when visualised. It determines the grouping metadata factor through the `group_by` parameter.

### Usage

```
lp_arrange(group_by)
```

### Arguments

<code>group_by</code>	A character value determining the column to group by in the samples metadata.
-----------------------	---

### Value

A function that takes an `lcmsPlot` object and returns a modified version with the specified arrangement options stored in `options$arrangement`. It is intended for use with the `+` operator, which incrementally layers new data or visual components onto the `lcmsPlot` object.

### Examples

```
raw_files <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE)[1:5]

## Plots chromatograms overlaid without specifying a grouping factor
p <- lcmsPlot(raw_files) +
  lp_chromatogram(aggregation_fun = "max")
p

## Plots chromatograms overlaid specifying a grouping factor
## (e.g., sample_id)
p <- p + lp_arrange(group_by = "sample_id")
p
```

---

lp\_chromatogram      *Define the chromatograms to plot*

---

### Description

The lp\_chromatogram function allows the generation of different types of chromatograms.

### Usage

```
lp_chromatogram(
  features = NULL,
  sample_ids = NULL,
  ppm = 10,
  rt_tol = 10,
  line_type = "solid",
  highlight_peaks = FALSE,
  highlight_peaks_color = NULL,
  highlight_peaks_mode = "polygon",
  highlight_peaks_factor = "sample_id",
  aggregation_fun = "max",
  rt_type = "uncorrected",
  rt_unit = "second",
  intensity_unit = "absolute",
  fill_gaps = FALSE,
  na.rm = FALSE,
  highlight_apices = list(column = NULL, top_n = NULL)
)
```

### Arguments

features	Specifies which features to generate the chromatogram for. This can be either: a matrix with columns mz and rt (optional); a matrix with columns mzmin, mzmax, rtmin (optional), rtmax (optional); a data.frame with columns sample_id, mz and rt (optional); a data.frame with columns sample_id, mzmin, mzmax, rtmin (optional), rtmax (optional); a character vector representing the grouped peaks (feature) names as returned by xcms::groupnames - requires the data to be an XCMSnExp or MsExperiment object with grouped peaks.
sample_ids	A character vector specifying the sample IDs to include in the plot. If NULL, the function uses the sample IDs specified in the lcmsPlot object.
ppm	A numeric value specifying the mass accuracy (in ppm) used when generating chromatograms. Ignored when the features parameter specifies both mzmin and mzmax.
rt_tol	A numeric value specifying the RT tolerance used when generating chromatograms. Ignored when the features parameter specifies both rtmin and rtmax.

<code>line_type</code>	A character value specifying the line type (from <code>ggplot2</code> ). One of: "solid", "dashed", "dotted", "dotdash", "longdash", "twodash".
<code>highlight_peaks</code>	A logical value indicating whether to highlight the detected peaks; the input data must be an <code>XCMSnExp</code> or <code>MsExperiment</code> object.
<code>highlight_peaks_color</code>	A character value indicating the color of the highlighted peaks.
<code>highlight_peaks_mode</code>	A character value indicating how the peaks should be highlighted. One of "polygon" (fills the area under the curve), "rectangle" (draws a bounding box from <code>rtmin</code> to <code>rtmax</code> up to the peak apex), or "point" (marks the peak apex with a point). Defaults to "polygon".
<code>highlight_peaks_factor</code>	A character value indicating the factor from the metadata that determines the color. By default it colors by <code>sample_id</code> .
<code>aggregation_fun</code>	A character value indicating which aggregation function to use for the spectra intensities; one of <code>max</code> or <code>sum</code> . Only applicable to summary chromatograms.
<code>rt_type</code>	A character value indicating what type of RT to use for the chromatograms. One of <code>uncorrected</code> (default), <code>corrected</code> , or <code>both</code> ; the input data must be an <code>XCMSnExp</code> or <code>MsExperiment</code> object. If both is chosen, this will give access to a metadata column called <code>rt_adjusted</code> that can be used to differentiate the two RT types (e.g., through faceting).
<code>rt_unit</code>	A character value indicating the unit to use for the RT axis; one of "minute" or "second".
<code>intensity_unit</code>	A character value indicating the unit to use for the intensity axis; one of "absolute" or "relative".
<code>fill_gaps</code>	A logical value indicating whether to fill gaps in RT with 0 intensity.
<code>na.rm</code>	A logical value. When <code>TRUE</code> , data points whose intensity is <code>NA</code> are removed before plotting. Defaults to <code>FALSE</code> .
<code>highlight_apices</code>	A logical value indicating whether to highlight apices with the corresponding RT values in a chromatogram.

## Value

This function returns another function that takes an `lcmsPlot` object and produces a modified version containing the generated chromatograms in its data slot. It is designed to be used with the `+` operator, which serves as a layering mechanism. Each use of `+` incrementally enriches the `lcmsPlot` object by adding new data or visual components.

## Summary chromatograms

In this type of chromatogram, the intensities of the spectra from each scan in an LC–MS dataset are either summed to produce the total ion current (TIC) chromatogram or the most intense peak is selected to produce the base peak chromatogram (BPC). To create such chromatograms do not

specify the features parameter as that will create the chromatograms for the selected features. In this context, the main parameter is aggregation\_fun which can take either sum (TIC) or max (BPC).

### Feature chromatograms

A feature is a combination of retention time (RT) and m/z. Feature chromatograms can be created by specifying the features parameter.

### Examples

```
raw_files <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE)[1:5]

p <- lcmsPlot(raw_files) +
  lp_chromatogram(aggregation_fun = "max") +
  lp_arrange(group_by = "sample_id")

p
```

---

lp\_compound\_discoverer

*Define the options to use when plotting LC-MS data coming from Compound Discoverer results.*

---

### Description

Define the options to use when plotting LC-MS data coming from Compound Discoverer results.

### Usage

```
lp_compound_discoverer(compounds_query = NULL, rt_extend = 10)
```

### Arguments

compounds\_query

A character value indicating the expression used to filter compounds from the Compound Discoverer results. The expression is evaluated on the compound table and can reference the following columns:

**name** Compound name.

**formula** Chemical formula of the compound.

**adduct** Ion adduct (e.g. [M+H]<sup>+</sup>, [M-H]<sup>-</sup>).

**rt** Retention time of the compound (in seconds).

**rtmin** Minimum retention time of the compound peak.

**rtmax** Maximum retention time of the compound peak.

**mz** Mass-to-charge ratio (m/z) of the detected ion.  
**maxo** Maximum observed peak intensity.  
**into** Integrated peak area reported by Compound Discoverer.  
**rt\_extend** A numeric value indicating how much (in seconds) the retention time window should be extended on each side of the compound peak when extracting and plotting chromatograms.

### Value

A function that takes an `lcmsPlot` object and returns a modified version with the specified Compound Discoverer options stored in `options$compound_discoverer`. It is intended for use with the `+` operator, which incrementally layers new data or visual components onto the `lcmsPlot` object.

### Examples

```
## Not run:
lcmsPlot("cd_example.cdResult") +
  lp_compound_discoverer(
    compounds_query = 'name %in% c("Proline", "Betaine")',
    rt_extend = 5
  ) +
  lp_chromatogram(highlight_peaks = TRUE) +
  lp_grid(rows = "sample_id", cols = "name", free_x = TRUE) +
  lp_labels(title = "Compound Discoverer example", legend = "Sample") +
  lp_legend(position = "bottom")

## End(Not run)
```

---

lp\_facets                      *Define the plot's faceting*

---

### Description

The `lp_facets` function arranges plots into a grid based on a metadata factor, creating a series of smaller plots (facets).

### Usage

```
lp_facets(facets, ncol = NULL, nrow = NULL, free_x = FALSE, free_y = FALSE)
```

### Arguments

**facets** A character vector of factors from the sample metadata to use for faceting.  
**ncol** A numeric value indicating the number of columns in the layout.  
**nrow** A numeric value indicating the number of rows in the layout.  
**free\_x** A logical value indicating whether the x-axis scales are allowed to vary across panels.  
**free\_y** A logical value indicating whether the y-axis scales are allowed to vary across panels.

**Value**

A function that takes an `lcmsPlot` object and returns a modified version with the specified faceting options stored in `options$facets`. It is intended for use with the `+` operator, which incrementally layers new data or visual components onto the `lcmsPlot` object.

**Examples**

```
raw_files <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE)[1:5]

## Plots chromatograms overlaid
p <- lcmsPlot(raw_files) +
  lp_chromatogram(aggregation_fun = "max")
p

## Using lp_facets we create facets for each sample_id
p <- p + lp_facets(facets = "sample_id")
p
```

---

`lp_get_plot`*Get the underlying plot object.*

---

**Description**

Get the underlying plot object.

**Usage**

```
lp_get_plot()
```

**Value**

A function that takes an `lcmsPlot` object and returns a modified version with the rendered plot stored in the `plot` slot. It is intended for use with the `+` operator, which incrementally layers new data or visual components onto the `lcmsPlot` object.

**Examples**

```
raw_files <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE)[1:4]

## Create faceted chromatogram plots with a reference RT line
p <- lcmsPlot(raw_files) +
  lp_chromatogram(features = rbind(c(
    mzmin = 334.9,
```

```

      mzmax = 335.1,
      rtmin = 2700,
      rtmax = 2900))) +
lp_facets(facets = 'sample_id', ncol = 4) +
lp_rt_line(intercept = 2800, line_type = 'solid', color = 'red')
p

## Extract the ggplot object and apply a theme
p <- p +
  lp_get_plot() +
  ggplot2::theme_bw()
p

```

---

lp\_grid

*Define a gridded plot*


---

### Description

The `lp_grid` function arranges plots into a matrix of panels defined by row and column faceting metadata factors.

### Usage

```
lp_grid(rows, cols, free_x = FALSE, free_y = FALSE)
```

### Arguments

<code>rows</code>	A character value indicating the factors that represent rows.
<code>cols</code>	A character value indicating the factors that represent columns.
<code>free_x</code>	A logical value indicating whether the x-axis scales are allowed to vary across panels.
<code>free_y</code>	A logical value indicating whether the y-axis scales are allowed to vary across panels.

### Value

A function that takes an `lcmsPlot` object and returns a modified version with the specified grid options stored in `options$grid`. It is intended for use with the `+` operator, which incrementally layers new data or visual components onto the `lcmsPlot` object.

### Examples

```

raw_files <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE
)[1:4]

```

```
## Create metadata for the samples
metadata <- data.frame(
  sample_id = sub("\\\\.CDF", "", basename(raw_files)),
  factor1 = c("S", "S", "C", "C"),
  factor2 = c("T", "U", "T", "U")
)

## Create feature chromatograms for the specified samples
p <- lcmsPlot(raw_files, metadata = metadata) +
  lp_chromatogram(features = rbind(c(
    mzmin = 334.9,
    mzmax = 335.1,
    rtmin = 2700,
    rtmax = 2900)))
p

## Arrange chromatograms in a grid split by experimental factors
## Rows correspond to `factor1` and columns correspond to `factor2`
p <- p + lp_grid(rows = "factor1", cols = "factor2")
p
```

---

lp_intensity_map	<i>Define a 2D intensity map</i>
------------------	----------------------------------

---

## Description

The `lp_intensity_map` function produces an intensity map in which signal intensity is represented at each  $m/z$  / RT coordinate.

## Usage

```
lp_intensity_map(
  mz_range,
  rt_range,
  sample_ids = NULL,
  density = FALSE,
  x_dim = "rt",
  y_dim = "mz",
  fill_scale = NULL
)
```

## Arguments

<code>mz_range</code>	A numeric value indicating the $m/z$ range of the map.
<code>rt_range</code>	A numeric value indicating the RT range of the map.
<code>sample_ids</code>	A character vector specifying the sample IDs to include in the plot. If <code>NULL</code> , the function uses the sample IDs specified in the <code>lcmsPlot</code> object.
<code>density</code>	A logical value indicating whether to show a density plot.

x_dim	A character value indicating which dimension to place on the x-axis. One of "rt" (default) or "mz".
y_dim	A character value indicating which dimension to place on the y-axis. One of "mz" (default) or "rt".
fill_scale	A ggplot2 scale object to use for the fill aesthetic (e.g. scale_fill_gradient(low = "white", high = "red")). When NULL (default), uses scale_fill_viridis_c for intensity and scale_fill_viridis_d for density plots.

### Value

This function returns another function that takes an lcmsPlot object and produces a modified version containing the generated 2D intensity map in its data slot. It is designed to be used with the + operator, which serves as a layering mechanism. Each use of + incrementally enriches the lcmsPlot object by adding new data or visual components.

### Examples

```
raw_files <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE)[1]

p <- lcmsPlot(raw_files) +
  lp_intensity_map(
    mz_range = c(200, 600),
    rt_range = c(4200, 4500),
    density = TRUE)

p
```

---

lp\_labels

*Define the labels of the plot*

---

### Description

The lp\_labels function allows the specification of the plot title and the legend title.

### Usage

```
lp_labels(title = NULL, legend = NULL)
```

### Arguments

title	A character value indicating the plot title.
legend	A character value indicating the legend's title.

**Value**

A function that takes an `lcmsPlot` object and returns a modified version with the specified label options stored in `options$labels`. It is intended for use with the `+` operator, which incrementally layers new data or visual components onto the `lcmsPlot` object.

**Examples**

```
raw_files <- dir(
  system.file("cdf", package = "faahKO"),
  full.names = TRUE,
  recursive = TRUE)[1:5]

## Create a chromatogram plot by grouping samples into batches
## By default, the legend is derived from the grouping variable
p <- lcmsPlot(raw_files, batch_size = 2) +
  lp_chromatogram(features = rbind(c(
    mzmin = 334.9,
    mzmax = 335.1,
    rtmin = 2700,
    rtmax = 2900))) +
  lp_arrange(group_by = "sample_id")
p

## Customise the legend label
p <- p + lp_labels(legend = "Sample")
p
```

---

lp\_layout

*Define the plot layout*

---

**Description**

Define the plot layout

**Usage**

```
lp_layout(design = NULL)
```

**Arguments**

`design` Specification of the location of areas in the layout See [https://patchwork.data-imaginist.com/reference/wrap\\_plots.html](https://patchwork.data-imaginist.com/reference/wrap_plots.html)

**Value**

A function that takes an `lcmsPlot` object and returns a modified version with the specified layout options stored in `options$layout`. It is intended for use with the `+` operator, which incrementally layers new data or visual components onto the `lcmsPlot` object.

**Examples**

```

data_obj <- get_XCMSnExp_object_example(indices = 1)

## Plot chromatograms and spectra for selected samples and features
p <- lcmsPlot(data_obj, sample_id_column = 'sample_name') +
  lp_chromatogram(
    features = rbind(
      c(mzmin = 334.9, mzmax = 335.1, rtmin = 2700, rtmax = 2900),
      c(mzmin = 278.99721, mzmax = 279.00279, rtmin = 2740, rtmax = 2840)
    ),
    sample_ids = 'ko15',
    highlight_peaks = TRUE
  ) +
  lp_spectra(mode = "closest_apex", ms_level = 1) +
  lp_facets(facets = "feature_id", ncol = 2)

## Customise panel layout to place chromatogram above spectra
p <- p + lp_layout(design = "C\nS\nS")

```

---

lp\_legend

*Define the legend layout*


---

**Description**

Define the legend layout

**Usage**

```
lp_legend(position = NULL)
```

**Arguments**

**position** A character value indicating the legend's position. One of "top", "right", "bottom", "left", or "inside".

**Value**

A function that takes an `lcmsPlot` object and returns a modified version with the specified legend options stored in `options$legend`. It is intended for use with the `+` operator, which incrementally layers new data or visual components onto the `lcmsPlot` object.

**Examples**

```

raw_files <- dir(
  system.file("cdf", package = "faahKO"),
  full.names = TRUE,
  recursive = TRUE)[1:5]

## Create a chromatogram plot grouped by sample with a custom legend label

```

```
p <- lcmsPlot(raw_files, batch_size = 2) +
  lp_chromatogram(features = rbind(c(
    mzmin = 334.9,
    mzmax = 335.1,
    rtmin = 2700,
    rtmax = 2900))) +
  lp_arrange(group_by = "sample_id") +
  lp_labels(legend = "Sample")
p

## Move the legend below the plot
p <- p + lp_legend(position = "bottom")
p
```

---

lp\_mass\_trace

*Define the mass trace to plot*

---

## Description

The `lp_mass_trace` function enables the generation of mass traces, which are graphical representations commonly used in mass spectrometry data analysis. A mass trace plots individual data points defined by their retention time and corresponding mass-to-charge ratio ( $m/z$ ), making it easier to visualise how specific ions behave over the course of a chromatographic run.

## Usage

```
lp_mass_trace()
```

## Value

This function returns another function that takes an `lcmsPlot` object and produces a modified version containing the generated mass traces in its data slot. It is designed to be used with the `+` operator, which serves as a layering mechanism. Each use of `+` incrementally enriches the `lcmsPlot` object by adding new data or visual components.

## Examples

```
raw_files <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE)[1:5]

## Create chromatograms of a specific feature
p <- lcmsPlot(raw_files) +
  lp_chromatogram(features = rbind(c(
    mzmin = 334.9,
    mzmax = 335.1,
    rtmin = 2700,
    rtmax = 2900))) +
```

```
lp_arrange(group_by = "sample_id")

## Add mass traces
p <- p + lp_mass_trace()

p
```

---

lp\_peak\_density

*Plot the peak density for one or more m/z features*

---

## Description

The `lp_peak_density` function produces a peak density plot. For each supplied feature (m/z bin):

- The x axis shows retention time.
- The y axis shows sample indices (1 to n), positioned within the density range.
- Detected peaks are plotted as coloured points at each sample's position.
- The kernel density estimate of peak apex RTs is drawn as a line.
- When `min_fraction` is supplied, the density-descent grouping algorithm is simulated and feature groups that pass the threshold are highlighted with semi-transparent rectangles.

## Usage

```
lp_peak_density(
  features = NULL,
  bw = 30,
  min_fraction = NULL,
  min_samples = 1L,
  sample_groups = NULL,
  max_features = 50L,
  rt_unit = "second"
)
```

## Arguments

<code>features</code>	A matrix or data frame with columns <code>mzmin</code> and <code>mzmax</code> (required) and <code>rtmin</code> , <code>rtmax</code> (optional). Each row defines one m/z bin. When omitted, the features are taken from <code>lp_chromatogram</code> if it has already been called on the same object.
<code>bw</code>	A numeric value specifying the kernel density bandwidth in seconds. Passed to <code>stats::density()</code> . Defaults to 30.
<code>min_fraction</code>	A numeric value in $[0, 1]$ . When supplied, simulates the <code>PeakDensityParam</code> grouping algorithm and draws semi-transparent rectangles for feature groups where at least this fraction of samples (per group) contain a peak. Defaults to <code>NULL</code> (no simulation).

min_samples	An integer specifying the minimum absolute number of samples per group required to define a feature group. Only used when min_fraction is set. Defaults to 1L.
sample_groups	A vector of length equal to the number of samples assigning each sample to a group (as in PeakDensityParam). Defaults to NULL, which treats all samples as a single group.
max_features	An integer specifying the maximum number of feature group rectangles to draw per m/z bin. Defaults to 50L.
rt_unit	A character value indicating the unit for the RT axis; one of "second" (default) or "minute".

### Value

This function returns another function that takes an `lcmsPlot` object and produces a modified version containing the generated peak density data in its data slot. It is designed to be used with the `+` operator, which serves as a layering mechanism.

### Examples

```
data_obj <- get_XCMSnExp_object_example(
  indices = 1:3,
  should_group_peaks = TRUE)
p <- lcmsPlot(data_obj, sample_id_column = "sample_name") +
  lp_peak_density(
    features = rbind(c(mzmin = 334.9, mzmax = 335.1,
                      rtmin = 2700, rtmax = 2900)),
    bw = 30,
    min_fraction = 0.5)
p
```

---

lp_rt_diff_plot	<i>Generate the retention time difference plot between raw and adjusted datasets</i>
-----------------	--

---

### Description

The `lp_rt_diff_plot` function generates the data necessary to plot the difference between the raw and retention time adjusted datasets. Only applicable to `XCMSnExp` and `MsExperiment` objects.

### Usage

```
lp_rt_diff_plot()
```

### Value

This function returns another function that takes an `lcmsPlot` object and produces a modified version containing the generated retention time differences, between raw and adjusted, in its data slot. It is designed to be used with the `+` operator, which serves as a layering mechanism. Each use of `+` incrementally enriches the `lcmsPlot` object by adding new data or visual components.

**Examples**

```
data_obj <- get_XCMSnExp_object_example(
  indices = 1:3,
  should_group_peaks = TRUE)
p <- lcmsPlot(data_obj, sample_id_column = "sample_name") +
  lp_rt_diff_plot()
p
```

---

lp\_rt\_line

*Define a vertical line on a retention time value*

---

**Description**

Define a vertical line on a retention time value

**Usage**

```
lp_rt_line(intercept, line_type = "dashed", color = "black")
```

**Arguments**

intercept	A numeric value indicating the retention time axis (x-axis) intercept.
line_type	A character value indicating the line type. One of "solid", "dashed", "dotted", "dotdash", "longdash", "twodash".
color	A character value indicating the line color.

**Value**

A function that takes an lcmsPlot object and returns a modified version with the specified RT line options stored in options\$rt\_lines. It is intended for use with the + operator, which incrementally layers new data or visual components onto the lcmsPlot object.

**Examples**

```
raw_files <- dir(
  system.file("cdf", package = "faahKO"),
  full.names = TRUE,
  recursive = TRUE)[1:4]

## Create chromatogram plots faceted by sample
p <- lcmsPlot(raw_files) +
  lp_chromatogram(features = rbind(c(
    mzmin = 334.9,
    mzmax = 335.1,
    rtmin = 2700,
    rtmax = 2900))) +
  lp_facets(facets = 'sample_id', ncol = 4)
p
```

```
## Add a vertical retention time reference line
p <- p + lp_rt_line(intercept = 2800, line_type = 'solid', color = 'red')
p
```

---

lp\_spectra

*Define the spectra to plot*


---

## Description

The `lp_spectra` function enables the generation of spectra, which are graphical representations of ions detected at each mass-to-charge ratio ( $m/z$ ) with their corresponding absolute or relative intensities.

## Usage

```
lp_spectra(
  sample_ids = NULL,
  mode = "closest_apex",
  ms_level = 1,
  rt = NULL,
  scan_index = NULL,
  interval = 3,
  spectral_match_db = NULL,
  match_target_index = NULL,
  peak_label_size = 3,
  intensity_breaks_by = 20,
  auto_facet = TRUE
)
```

## Arguments

<code>sample_ids</code>	A character vector specifying the sample IDs to include in the plot. If <code>NULL</code> , the function uses the sample IDs specified in the <code>lcmsPlot</code> object or the <code>lp_chromatogram</code> function.
<code>mode</code>	The method to choose the scan from which to extract the spectra. One of: <code>closest</code> , the closest scan to the specified RT - <code>rt</code> parameter); <code>closest_apex</code> , the closest scan to a detected peak; <code>across_peak</code> , selects scans across a detected peak at a certain interval specified in the <code>interval</code> parameter. <code>mode</code> is not applicable to standalone spectra.
<code>ms_level</code>	The MS level to consider for the scan.
<code>rt</code>	When <code>mode = "closest"</code> , the RT to consider.
<code>scan_index</code>	The exact scan index to consider for extracting a spectrum. <code>scan_index</code> and <code>mode</code> are mutually exclusive.
<code>interval</code>	When <code>mode = "across_peak."</code> The RT interval to consider.

spectral_match_db	The database containing reference spectra used for matching and comparison with the input spectra.
match_target_index	The index, ranked by descending match score, identifying which reference spectrum to display in the mirror plot.
peak_label_size	A numeric value controlling the font size of m/z labels annotated on spectral peaks. Defaults to 3.
intensity_breaks_by	A numeric value specifying the step size (in percent) between y-axis intensity breaks. Defaults to 20.
auto_facet	A logical value. When TRUE (default), a facet is automatically added to separate spectra from different samples or scans. Set to FALSE to suppress automatic faceting.

### Value

This function returns another function that takes an `lcmsPlot` object and produces a modified version containing the generated spectra in its data slot. It is designed to be used with the `+` operator, which serves as a layering mechanism. Each use of `+` incrementally enriches the `lcmsPlot` object by adding new data or visual components.

### Spectra associated with chromatograms

A spectrum is obtained from a scan at a specific retention time (RT). Therefore, when plotting a chromatogram together with its associated spectra, it is common to mark the RT with a vertical line on the chromatogram to indicate where the spectra were acquired. See the example below on how to generate these types of spectra.

### Standalone spectra

Standalone spectra can also be generated, provided no chromatograms are present (i.e., `lp_chromatogram` has not been used).

### Examples

```
raw_files <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE)[1]

p <- lcmsPlot(raw_files) +
  lp_chromatogram(features = rbind(c(
    mzmin = 334.9,
    mzmax = 335.1,
    rtmin = 2700,
    rtmax = 2900))) +
  lp_spectra(mode = "closest", rt = 2785)
p
```

---

lp\_total\_ion\_current *Define the total ion current (TIC)*

---

### Description

The `lp_total_ion_current` generates summary data for the total ion current (TIC) of the selected samples.

### Usage

```
lp_total_ion_current(sample_ids = NULL, type = "boxplot")
```

### Arguments

`sample_ids` A character vector specifying the sample IDs to include in the plot. If `NULL`, the function uses the sample IDs specified in the `lcmsPlot` object.

`type` A character value indicating the type of plot; one of "boxplot", "violin", "jitter".

### Value

This function returns another function that takes an `lcmsPlot` object and produces a modified version containing the generated total ion current (TIC) in its data slot. It is designed to be used with the `+` operator, which serves as a layering mechanism. Each use of `+` incrementally enriches the `lcmsPlot` object by adding new data or visual components.

### Examples

```
data_obj <- get_XCMSnExp_object_example()

p <- lcmsPlot(data_obj, sample_id_column = "sample_name") +
  lp_total_ion_current(type = "violin") +
  lp_arrange(group_by = "sample_id")

p
```

---

MsDialPeaksSource *Create an MS-DIAL data source from peak lists*

---

### Description

This function reads MS-DIAL peak list files and sample metadata and converts them into a format compatible with `xcms`-style peak tables.

### Usage

```
MsDialPeaksSource(peaks_paths, sample_paths, metadata_path = NULL)
```

## Arguments

- `peaks_paths` A character vector of paths to MS-DIAL peak list files (CSV or TSV). The ordering should match the one in the metadata (i.e., the samples).
- `sample_paths` A character vector of paths to raw sample files (e.g., mzML).
- `metadata_path` A character value (optional) indicating the path to a metadata file (CSV or TSV). If NULL, a metadata data.frame is created from `sample_paths`.

## Value

An object of class `ExternalDataSource`.

## Examples

```
## Create temporary example files
tmp_dir <- tempdir()

## Fake raw sample paths
sample_paths <- file.path(
  tmp_dir,
  c("sample1.mzML", "sample2.mzML")
)

## Create minimal MS-DIAL peak list files (one per sample)
peak_list_paths <- file.path(
  tmp_dir,
  c("sample1_peaks.csv", "sample2_peaks.csv")
)

msdial_peaks_1 <- tibble::tibble(
  "Precursor m/z" = c(100.1, 200.2),
  "RT (min)" = c(5.0, 10.0),
  "Area" = c(10000, 20000),
  "Height" = c(500, 800),
  "RT left(min)" = c(4.8, 9.8),
  "RT right (min)" = c(5.2, 10.2)
)

msdial_peaks_2 <- tibble::tibble(
  "Precursor m/z" = c(150.3, 250.4),
  "RT (min)" = c(6.0, 12.0),
  "Area" = c(15000, 25000),
  "Height" = c(600, 900),
  "RT left(min)" = c(5.8, 11.8),
  "RT right (min)" = c(6.2, 12.2)
)

utils::write.csv(msdial_peaks_1, peak_list_paths[1], row.names = FALSE)
utils::write.csv(msdial_peaks_2, peak_list_paths[2], row.names = FALSE)

## Create the data source
ds <- MsDialPeaksSource(
```

```
    peaks_paths = peak_list_paths,  
    sample_paths = sample_paths  
  )
```

---

MZmineFeatureListsSource

*Create an MZmine data source from feature lists*

---

## Description

This function reads MZmine feature list files (supports version 2 and above) and sample metadata and converts them into a format compatible with xcms-style peak tables.

## Usage

```
MZmineFeatureListsSource(  
  feature_lists_paths,  
  sample_paths,  
  metadata_path = NULL  
)
```

## Arguments

`feature_lists_paths` A character vector of paths to MZmine feature list files (CSV or TSV).

`sample_paths` A character vector of paths to raw sample files (e.g., mzML).

`metadata_path` A character value (optional) indicating the path to a metadata file (CSV or TSV). If NULL, a metadata data frame is created from `sample_paths`.

## Value

An object of class `ExternalDataSource`.

## Examples

```
## Create temporary example files  
tmp_dir <- tempdir()  
  
## Fake sample paths  
sample_paths <- file.path(  
  tmp_dir,  
  c("sample1.mzML", "sample2.mzML")  
)  
  
## Create a minimal MZmine 2 feature list CSV  
feature_list_path <- file.path(tmp_dir, "mzmine_features.csv")  
  
mzmine_features <- tibble::tibble(  
  # ...  
)
```

```

"row m/z" = c(100.1, 200.2),
"sample1.mzML Feature status" = c("DETECTED", "DETECTED"),
"sample1.mzML Feature m/z" = c(100.1, 200.2),
"sample1.mzML Feature RT" = c(300, 600),
"sample1.mzML Peak area" = c(10000, 20000),
"sample1.mzML Peak height" = c(500, 800),
"sample1.mzML Feature m/z min" = c(99.9, 199.9),
"sample1.mzML Feature m/z max" = c(100.3, 200.4),
"sample1.mzML Feature RT start" = c(290, 590),
"sample1.mzML Feature RT end" = c(310, 610),
check.names = FALSE
)

utils::write.csv(mzmine_features, feature_list_path, row.names = FALSE)

## Create the data source
ds <- MZmineFeatureListsSource(
  feature_lists_paths = feature_list_path,
  sample_paths = sample_paths
)

```

---

next\_plot

*Move to the next plot object (batch-mode)*


---

## Description

next\_plot progresses an lcmsPlotClass object to the next plot in a batch-processing sequence. This is typically used when multiple plots are generated and inspected iteratively, such as when navigating large LC–MS datasets in a batched workflow. The batch size is defined in the lcmsPlot function’s argument batch\_size.

## Usage

```

next_plot(object)

## S4 method for signature 'lcmsPlotClass'
next_plot(object)

```

## Arguments

object            An instance of class lcmsPlotClass.

## Value

An instance of class lcmsPlotClass.

## Examples

```
raw_files <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE)[1:5]

p <- lcmsPlot(raw_files, batch_size = 2) +
  lp_chromatogram(features = rbind(c(
    mzmin = 334.9,
    mzmax = 335.1,
    rtmin = 2700,
    rtmax = 2900))) +
  lp_arrange(group_by = "sample_id") +
  lp_legend(position = "bottom") +
  lp_labels(legend = "Sample")

p <- next_plot(p)
p
```

---

show,ExternalDataSource-method

*Show a summary of an instance of class ExternalDataSource*

---

## Description

Show a summary of an instance of class ExternalDataSource

## Usage

```
## S4 method for signature 'ExternalDataSource'
show(object)
```

## Arguments

object            An instance of class ExternalDataSource.

## Value

Invisible NULL

## Examples

```
## Create dummy metadata
metadata <- data.frame(
  sample_id = c("S1", "S2"),
  sample_path = c("sample1.mzML", "sample2.mzML")
)

## Create dummy peaks
```

```
peaks <- data.frame(
  mz = c(100.1, 150.2),
  rt = c(300, 450),
  rtmin = c(290, 440),
  rtmax = c(310, 460),
  into = c(10000, 15000),
  maxo = c(2000, 2500),
  sample_index = c(1, 2)
)

## Create ExternalDataSource object
eds <- new(
  "ExternalDataSource",
  name = "Example data source",
  metadata = metadata,
  peaks = peaks
)

## Show summary
eds
```

---

show,lcmsPlotClass-method

*Plot the lcmsPlotClass object*

---

## Description

Display an instance of lcmsPlotClass class to the selected device.

## Usage

```
## S4 method for signature 'lcmsPlotClass'
show(object)
```

## Arguments

object            An instance of class lcmsPlotClass.

## Value

Invisible NULL

## Examples

```
raw_files <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE)[1:5]

## Shows summary information as the plot has not been built yet
```

```
p <- lcmsPlot(raw_files)
p

## Shows the actual plot
p <- lcmsPlot(raw_files) +
  lp_chromatogram(aggregation_fun = "max") +
  lp_arrange(group_by = "sample_id") +
  lp_legend(position = "bottom") +
  lp_labels(legend = "Sample")

p
```

---

show,lcmsPlotDataContainer-method

*Show a summary of an instance of class lcmsPlotDataContainer*

---

## Description

Show a summary of an instance of class lcmsPlotDataContainer

## Usage

```
## S4 method for signature 'lcmsPlotDataContainer'
show(object)
```

## Arguments

object            An instance of class lcmsPlotDataContainer.

## Value

Invisible NULL

## Examples

```
raw_files <- dir(
  system.file("cdf", package = "faahKO"),
  full.names = TRUE,
  recursive = TRUE)[1:5]

data_obj <- new("lcmsPlotDataContainer",
  data_obj = raw_files,
  metadata = tibble::tibble(),
  chromatograms = tibble::tibble(),
  mass_traces = tibble::tibble(),
  spectra = tibble::tibble(),
  peak_density = tibble::tibble(),
  total_ion_current = tibble::tibble(),
  intensity_maps = tibble::tibble(),
  rt_diff = tibble::tibble(),
```

```
feature_metadata = tibble::tibble(),
detected_peaks = tibble::tibble()
data_obj
```

---

show, XcmsRawList-method

*Show a summary of an XcmsRawList object*

---

### Description

Show a summary of an XcmsRawList object

### Usage

```
## S4 method for signature 'XcmsRawList'
show(object)
```

### Arguments

object            An instance of XcmsRawList.

### Value

Invisible NULL.

---

XcmsRawList

*Create an XcmsRawList object*

---

### Description

Wraps one or more xcmsRaw objects into an XcmsRawList container for use as the dataset argument of lcmsPlot.

### Usage

```
XcmsRawList(...)
```

### Arguments

...            One or more xcmsRaw objects, or a single list of xcmsRaw objects.

### Value

An instance of XcmsRawList.

**Examples**

```
paths <- dir(
  system.file("cdf", package = "faahK0"),
  full.names = TRUE,
  recursive = TRUE
)[c(1, 2)]
raw1 <- xcms::xcmsRaw(paths[1])
raw2 <- xcms::xcmsRaw(paths[2])
x1 <- XcmsRawList(raw1, raw2)
```

---

XcmsRawList-class	<i>A list of xcmsRaw objects representing multiple samples</i>
-------------------	--

---

**Description**

XcmsRawList is a container for one or more xcmsRaw objects, where each element corresponds to a single sample. It is the recommended way to pass in-memory raw LC-MS data to lcmsPlot.

**Slots**

data A list of xcmsRaw objects, one per sample.

# Index

`+`, `lcmsPlotClass`, function-method, 4

`BiocParallelParam`, 8

`create_xcms_raw_list`, 4

`ExternalDataSource`-class, 5

`get_XCMSnExp_object_example`, 6

`iterate_plot_batches`, 6

`iterate_plot_batches`, `lcmsPlotClass`, function-method  
(`iterate_plot_batches`), 6

`lcmsPlot`, 7

`lcmsPlot`-package, 3

`lcmsPlotClass`-class, 8

`lcmsPlotDataContainer`-class, 9

`lp_arrange`, 10

`lp_chromatogram`, 11

`lp_compound_discoverer`, 13

`lp_facets`, 14

`lp_get_plot`, 15

`lp_grid`, 16

`lp_intensity_map`, 17

`lp_labels`, 18

`lp_layout`, 19

`lp_legend`, 20

`lp_mass_trace`, 21

`lp_peak_density`, 22

`lp_rt_diff_plot`, 23

`lp_rt_line`, 24

`lp_spectra`, 25

`lp_total_ion_current`, 27

`MsDialPeaksSource`, 27

`MZmineFeatureListsSource`, 29

`next_plot`, 30

`next_plot`, `lcmsPlotClass`-method  
(`next_plot`), 30

`show`, `ExternalDataSource`-method, 31

`show`, `lcmsPlotClass`-method, 32

`show`, `lcmsPlotDataContainer`-method, 33

`show`, `XcmsRawList`-method, 34

`stats::density()`, 22

`XcmsRawList`, 34

`XcmsRawList`-class, 35